Relationship of resistin levels with endometrial cancer risk


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Cancer of endometrium (CAE) is the most common gynecologic malignancy in industrialized nations. Increased resistin levels, an adipocytokine produced by adipose tissue and macrophages, have been considered as a risk factor in gastric, colon and breast cancer, recently. No studies associating resistin levels with endometrial cancer have been done so far. The purpose of this case-control study was to determine the relationship between serum circulating resistin levels and resistin gene -420C>G (rs3219175) variant in endometrial cancer patients. 37 Caucasian female patients and 39 healthy controls were enrolled in this study. Difference in resistin levels between age and BMI matched patients group (mean 24.2 ng/ml) and control subjects (mean 10.1 ng/ml) were statistically significant (p < 0.01). We also determined single nucleotide polymorphism -420C>G (rs3219175) within resistin gene and no significant association between resistin levels and investigated polymorphism was found. Furthermore, no significant association between higher resistin levels and diabetes mellitus 2, body mass index, smoking or age have been observed within studied groups. To our knowledge, this is the first study examining the relationship between serum resistin levels and endometrial cancer and our results show, that patients with endometrial cancer have significantly increased circulating levels of resistin compared to control subjects.

Key words: Endometrial cancer, resistin, SNP -420C>G

Cancer of endometrium (CAE), the neoplastic growth of endometrial epithelial cells, is the most common gynecologic malignancy in industrialized nations. Worldwide, it is diagnosed approximately 142 000 new endometrial carcinomas and approximately 42 000 women die of the disease every year. In the Czech Republic, the incidence has slightly increasing trend and in 2007 reached number 33/100 000 women [1]. In numbers, approximately 1796 new cases have been diagnosed and 567 women died in Czech Republic in 2007. CAE incidence rises dramatically following the menopause with climax in 6th decennium. Therefore increasing incidence trend could be explained by prolonged life-span and increased exposure to risk factors. Well-known risk factors, which are involved in carcinogenesis of CAE are obesity, hypertension, diabetes mellitus 2, nulliparity, polycystic ovarian disease, use of estrogen medications that lead to unopposed estrogen stimulation of the endometrium [2] and newly also inflammatory processes are considered as another important feature proposed to influence cancer development. Although, this risk factors are known for many years, the molecular mechanisms of endometrial cancer development and contribution degree of known risk factors in this process, especially obesity and diabetes mellitus 2, are still not fully understood. In last decade, the discovery that adipose tissue is not only energy-storage organ but active metabolic tissue, could answer many questions and shed a light on links between obesity, diabetes mellitus, cancer and other clinical features. The white adipose tissue acts as an active endocrine organ secreting multiple metabolically important proteins – ‘adipocytokines’, involved in physiological functions such as energy homeostasis, immunity and inflammation [3,4]. One of the relatively new identified adipokine is gene encoding for resistin (RETN). After its initial discovery in mice, it was suggested to be the link between obesity and type 2 diabetes. Steppan et al. [5] demonstrated that resistin expression was increased in obese animals, and moreover, administration of recombinant resistin to normal animals produced insulin resistance [5,6]. When considering resistin involvement in pathophysiology of obesity and diabetes mellitus 2 in humans, resistin studies have produced contradictory findings. Although some studies reported increase of resistin in obesity and diabetes type 2, most did not show correlation between resistin circulating levels and body mass index or insulin resist-
ance [7-10]. This contrast could be caused by fact, that human and mouse RETN genes share only 53% homology, therefore their functions may differ between them [11] and other roles of RETN may exist in human. Although the exact function of resistin in human is still not known, increased resistin levels have been associated with coronary heart disease [12-15] and considered as a risk factor in gastric, colon and breast cancer [16-19], recently. The purpose of this case-control study was to determine the relationship between serum circulating resistin levels and resistin gene -420C>G (rs3219175) variant in endometrial cancer patients.

Patients and methods

Subjects. Thirty seven Caucasian female patients and 39 healthy controls were enrolled in this study. Recruited patients were diagnosed for endometrial cancer and treated at a University Hospital Brno at Department of Obstetrics and Gynecology. Complete anamnesis, anthropometric measurements and medical documentation including blood tests and histology of endometrial tumor have been obtained. Healthy women with no previous history of any kind of cancer, with similar age and body mass index (BMI) were recruited as controls. For correct statistical analysis of resistin we matched 25 patients and 25 control subjects by age and BMI. All participants gave their written informed consent before they entered the study. This study was approved by the Committee for Ethics of Medical Experiments on Human Subjects, Faculty of Medicine, Masaryk University, Brno and was performed in adherence to the Declaration of Helsinki Guidelines.

Genotype analysis and serum level measurements. Blood samples were obtained from all individuals in EDTA anticoagulant and DNA was isolated by the salting-out method. Purity and concentration was measured on nanodrop spectrophotometer. Genotyping of -420C>G SNP (rs1862513) within resistin gene sequence was performed by polymerase chain reaction (PCR) and consequent restriction fragment length polymorphism (RFLP). Primers (forward primer 5’ – TTT TGT CAT GGT TGC ATC AGC – 3‘; reverse primer 5’ – GGG CTC AGC TAA CCA AAT – 3‘) were designed within resistin gene and its promoter sequence, carried in SNPper database (http://snpper.chip.org) which corresponds to contig NG_023447.1 (www.ncbi.nlm.nih.gov). 330-bp fragment was amplified by standard Taqman polymerase under following amplification conditions: total volume of 25 μl containing 150 ng genomic DNA, initial denaturation 94°C for 3 minutes, then 31 cycles of 94°C for 30 seconds, 55°C for 15 seconds, 70°C for 30 seconds, and 70°C for 10 minutes and consequently digested by endonuclease restriction enzyme BbsI (NEB, USA) under manufacturers recommendations. Results have been evaluated by single strand conformation polymorphism (SSCP) method.

Blood drawings for biochemical examinations were performed in the morning after 12 hour fasting. After serum separation were samples frozen at -20 degrees. Serum resistin level was measured by Enzyme-linked immunosorbent assay (ELISA), using human resistin ELISA kit (R&D Systems, USA). Intra- and inter-assay precisions were less than 5.3 and 9.2 %, respectively. The minimum detectable dose of resistin was 0.055 ng/mL.

 Statistical analysis. Basic descriptive statistics were used for general characterization of measured data. Considering quite low amount of patients and controls, median (and 5th and 95th percentile) was used as main characteristic. For correct usage of statistical method was necessary to test the normality of each quantitative trait. This was performed by histogram (Phi-Square) and Kolmogorov-Smirnov test. All evaluated parameters were significantly different from normal distribution. Thus we decided to use only non-parametric methods. For testing of dichotomic parameters (e.g. diabetes, smoking) was used Mann-Whitney test. To examine relations between continuous and categorical parameters with more than two categories (genotype), Kruskall-Wallis ANOVA was performed. P-values below the conventionally agreed level of significance (p < 0.05) were considered statistically significant. For determine adiposity were set cohorts stratified by baseline BMI = 25 which allowed match patients and controls for BMI and age (tested by M-W test). All results were carried out using software Statistica (version 9.0; StatSoft inc. (2009)).

Results

Standard robust summary statistics were used to describe distribution patterns in the primary data (mean, minimum, maximum). These are shown in Table1. The mean age of all participants was 49±13.5 years whereas patients had in average 57 years and controls 42 years. After creation of matched groups, the mean age changed quite slightly to 52 years (with range 26 – 61) and 48 years (with range 38 – 67) in group of patients and controls respectively. Homogeneity of age between groups was approved by Mann-Whitney test (p = 0.07). For matched groups was also performed testing of BMI. Computed mean value of BMI was 31.5 and 30.9 for patients and controls respectively. Difference in BMI between groups was not observed (p = 0.36). When we compared concentration of resistin between matched groups, the mean level was 24.2 ng/ml and 10.1 ng/ml in group of patients and controls respectively. The test proved statistically significant difference between groups (p < 0.01). It was necessary to test relationship between serum resistin level and BMI and between resistin level and age for each group separately. As

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<th>Table 1. Basic characteristics of patients and control subjects</th>
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measure of this relationship was applied Spearman correlation metric. Results are in Table 2.

We investigated whether the difference in serum resistin level between patients and control subjects is associated with resistin natural variant -420C>G polymorphism. All patients and control subjects were genotyped for mentioned polymorphism. These genotypes were divided into three groups (CC, CG, GG) and resistin dependency was tested by Kruskall-Wallis Anova (nonparametric one-way Anova). Dependency of serum resistin levels on genotype was not statistically significant (for control subjects p=0.39; patients p=0.72). Genotype distribution between patients and control group was tested by phi-square test and genotype distribution did not show any significant differences (χ2=0.567; p=0.75). For determination of similarity between genotypes associated with resistin levels (within each group separately) was used Tukey’s HSD test (Table 3). The most different are GC and CC genotypes, within patients group (p=0.75) and control group respectively (p=0.36), both of them are not significant. We have also found non-significant relationship between resistin levels and diabetes mellitus type 2 (p=0.707) hypertension (p=0.948) or smoking (p=0.272).

**Discussion**

Resistin is a member of the adipokine family and in initial studies has been reported, that resistin can cause insulin resistance and decrease adipocyte differentiation in rodents [5,6]. However, similar studies attempting to associate serum resistin concentration and occurrence of endometrial cancer remained after adjustment for BMI and age.

Osawa et al. found correlation between G/G natural variant of -420C>G polymorphism and higher level of circulating resistin in Japan diabetes mellitus type 2 patients. According to authors, this could be explained by specific recognition of -420G by Sp1/3, which increases RETN promoter activity, leading to enhanced serum resistin levels [24]. Also other studies on indicating that -420C>G variant within resistin gene promoter could be connected with higher serum levels [25, 26]. In contrary, we did not observe any association between serum resistin levels and -420G RETN variant. Moreover, the investigated polymorphism did not express any deviation in allele distribution from Hardy-Weinberg equilibrium within patients and controls group. Although our results do not correspond with previously mentioned studies, they are in compliance with recent work of Onum et al. on Japan cohort. Authors investigated other polymorphism within RETN gene named -358A, which is compound with -420G and both of them are required to confer higher serum resistin levels. In Caucasians, the association between SNP-420 and plasma resistin is not strong, and A at -358 may not exist, suggesting that SNP-358 could explain this ethnic difference [27]. Unfortunately, we did not investigate -358A polymorphism, so we can not confirm Onumi’s hypothesis. In addition, measured resistin levels in our study did not show any correlation with smoking, diabetes mellitus type 2 or hypertension.
In other words, based on our results resistin appears to have diabetes mellitus type 2, hypertension, smoking, age, adiposity and resistin SNP -420C>G independent link to endometrial cancer. Although the underlying mechanisms of this association are not clear, there are some suggestions why are higher serum resistin levels associated with gastric, colon, breast cancer and newly based on our findings also with endometrial cancer.

It has been published that even resistin is produced by adipose tissue, macrophages are more important source of resistin in humans [28]. Bokarewa et al. found that resistin exert potent pro-inflammatory properties by up-regulating pro-inflammatory cytokines, probably via NF-kB pathway [29].

In addition, recent data indicate that stimulation of macrophages in vitro with endotoxin or pro-inflammatory cytokines leads to a marked increase in resistin production [30]. In other clinical studies, resistin have been associated with inflammatory markers apparently independent on BMI, but related to risk factors of atherosclerosis [31, 32]. Together with facts that up-regulated pro-inflammatory cytokines and chemokines in tumors [33-35] and inflammation itself is recently recognized as risk factor contributing on cancer development and progression [36-39], resistin seems to be a novel secreted protein in relation to inflammation in humans and may present molecular link between inflammation and carcinogenesis.

Several limitations need to be considered when interpreting our findings. Firstly, this initial study was done on relatively small cohort therefore we agree that larger studies to validate our results need to be done. In this study we use only two single measurements of baseline levels of resistin, from one collected serum, which may subject to error regarding its ability to reflect the true status of circulating resistin. However, previous studies suggest that blood levels of resistin have a relatively low intra-subject variation over time [32] and this may reliably reflect the long-term resistin levels in adult women. Even likely, we can not elucidate whether or not higher serum resistin levels are indeed caused by production of resistin in endometrial cancer cells. All this facts need to be taken in account when considering our findings.

To conclude, the present study provides evidence that endometrial cancer patients had significantly higher serum levels of resistin compared to those without endometrial cancer. Converging lines of evidence from this study and earlier in vitro and case-control studies offer the background biological context for considering resistin as a novel link between inflammation and cancer. Further studies to fully understand the mechanisms underlying resistin's effect and contribution on cancer development/progression need to be done. It is on our best behalf to keep studying well-known and newly discovered risk factors as well, and their role in process of carcinogenesis.

References


