

The aqueous garlic, onion and leek extracts release nitric oxide from S-nitrosoglutathione and prolong relaxation of aortic rings

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Abstract. Garlic, onion and leek have beneficial effects in treatment of numerous health disorders. The aim of the present study was to investigate underlying molecular mechanisms. To test the potency of the aqueous garlic, onion and leek extracts to release NO from GSNO we have measured NO oxidation product, NO₂⁻, by the Griess reagent method. Further, we studied the ability of garlic extract to relax noradrenaline-precontracted rat aortic rings in the presence of GSNO and effects of garlic extract on electrical properties of rat heart intracellular chloride channels. We have observed that: i) garlic, onion and leek extracts released NO from GSNO in the order: garlic > onion > leek; ii) the ability of garlic extract to release NO was pH-dependent (8.0 > 7.4 > 6.0) and potentiated by thiols (Cys >> GSH = N-acetyl-cysteine > oxidized glutathione) at concentration 100 μmol/l; iii) the garlic extract (0.045 mg/ml) prolonged relaxation time of aortic rings induced by GSNO (50 nmol/l) and inhibited intracellular chloride channels.

We suggest that NO-releasing properties of the garlic, onion and leek extracts and their interaction with Cys and GSH are involved in NO-signalling pathway which contributes to some of its numerous beneficial biological effects.

Key words: Garlic – Nitric oxide – Cysteine – Aorta relaxation – Chloride channels

Abbreviations: BLM, bilayer lipid membrane; GSH, reduced glutathione; GSNO, nitrosoglutathione; GSSG, oxidised glutathione; NA, noradrenaline; NAC, N-acetylcysteine.

Introduction

Garlic (*Allium sativum* L.) and its various forms have numerous beneficial effects in treatment of different health disorders. It reduces cardiovascular risk, decreases oxidized low-density lipoproteins, abnormal platelet aggregation and high blood pressure (Singh and Singh 2008; Iciek et al. 2009; Ginter and Simko 2010). Garlic enhances immune functions and has antibacterial, antifungal, antiviral and antioxidative activities and may have anticarcinogenic effect. It is commonly used for treat-

ment of throat infections, digestive tract disorders and fungal infections (Singh and Singh 2008; Iciek et al. 2009; Ginter and Simko 2010). Medicinal properties of garlic have been attributed to organosulfur compounds (Rose et al. 2005; Jacob et al. 2008; Iciek et al. 2009). Recently it was observed that human red blood cells convert garlic-derived organic polysulphides into H₂S, a gasotransmitter, which influences numerous biological processes (Benavides et al. 2007; Lowicka and Beltowski 2007). Beneficial biological effects of onion and leek were also reported (Dorant et al. 1996; Rahimi et al. 2010).

It has been proposed that low-molecular weight thiols such as glutathione, which react with nitric oxide, are potential candidates for a nitric oxide-carrier molecule in living organism. Endogenous nitrosothiols, e.g. S-nitrosoglutathione (GSNO), may act as intermediates in the storage

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and/or transport of nitric oxide (Ng et al. 2007). H₂S gas and H₂S donors NaHS or Na₂S were found to release NO from NO-stores nitrosothiols, biological membranes and L1210 cells and NaHS inhibited intracellular chloride channels (Ondrias et al. 2008; Teng et al. 2008; Malekova et al. 2009; Bertova et al. 2010). Interaction of H₂S with nitroso-compounds potentiated aortic rings relaxation (Ondrias et al. 2008; Bertova et al. 2010). NO plays a central role in diverse signalling pathways. Besides its binding to a soluble guanylate cyclase it interacts directly or indirectly with many other proteins by post-translational modifications that can alter their function (Stamler 1994; Zhang and Hogg 2005). S-nitrosothiol, e.g. GSNO, and NO transfer reactions between protein and peptide cysteines have been proposed to regulate many biological processes (Stamler 1994; Zhang and Hogg 2005; Benhar et al. 2008).

To understand the numerous biological effects of garlic, onion and leek, and their possible involvement in NO- and GSNO-regulated biological processes, we studied properties of their extracts to release NO from NO-donor GSNO.

Materials and Methods

Chemicals

All chemicals were purchased from Sigma-Aldrich. Garlic and onion bulbs or leek bundles were obtained from local sources. The pieces of garlic, onion or leek weighting 6 g were pressed through 1.2 mm diameters pores to 20 ml buffer (in mmol/l: 160 KCl, 1 MgCl₂, 0.1 diethylene triamine-pentaacetic acid (DTPA), 50 HEPES/TRIS, 7.4 pH) to obtain a crude homogenate. The garlic, onion or leek homogenate was vortexed for 1 min (1800 rev/min) and the juice was extracted by separating the juice from debris using a teflon glass homogenizator. The aqueous garlic extract was used freshly or it was aliquoted 11 minutes after the cloves pulverization and stored at -70°C (64 mg of extract per ml). The frozen juice was used for several days. Na₂S was used as H₂S donor.

Measurement of NO release from GSNO

NO-donor GSNO (100 µmol/l) and garlic, onion or leek aqueous extracts were mixed in the buffer, incubated at 21 ± 1°C for 10 min. For measuring of NO oxidation product, nitrite (NO₂⁻), the Griess reagent was added. The samples were incubated for 10 min. Then, the absorption spectra at 540 nm were measured (µQuant, Bio-Tek Instrument Inc., USA) (Ondrias et al. 2008). Concentration of NO₂⁻ was calibrated by NaNO₂. Paired t-test was used to determine significance of an effect of the compounds. Data represent means ± SEM, *n* ≥ 3. Piperazine-N,N'-bis(2-ethanesulfonic acid) and TRIS were used to adjust buffer to 6.0 and 8.0 pH, respectively.

Measurement of rat aorta contractility

All procedures were approved by the State Veterinary and Food Administration of the Slovak Republic. Experiments were carried out as previously described (Ondrias et al. 2008, Bertova et al. 2010). Briefly, the rings of thoracic aorta were prepared from male Wistar rats and were mounted for recording of isometric tension changes in pneumoxid-oxygenated (95% O₂; CO₂; 37°C) Krebs-bicarbonate solution. The aortic rings were precontracted by 1 µmol/l noradrenaline (NA). The time dependent relaxation effect of GSNO (50 nmol/l) and GSNO in the presence of garlic extract (45 µg/ml) was normalized to the maximum relaxation peak (in %) obtained by GSNO (100%) or GSNO in the presence of garlic extract (100%). In these experiments, the final garlic extract concentration (45 µg/ml) was used, since this concentration did not have significant effect on the relaxation of aortic rings. Paired t-test was used to determine significance of an effect of the compounds. Data represent means ± SEM, *n* ≥ 3.

Bilayer lipid membrane measurements

Membrane vesicles containing crude rat heart mitochondrial, lysosomal and sarcoplasmic reticulum membranes were isolated as described previously (Malekova et al. 2009). Bilayer lipid membrane (BLM) was formed across an aperture (diameter ≤ 0.1 mm), separating the *cis* and *trans* chambers using a mixture of dioleoyl-glycero-phosphatidylcholine and dioleoyl-glycero-phosphoethanolamine at a molar ratio of 3 : 2 in *n*-decane (20 mg/ml). Composition of the *cis* and *trans* solutions was (in mmol/l) 0.1 CaCl₂, 0.3 EGTA, 1 MgCl₂, 10/5 HEPES/TRIS (pH 7.4), *cis/trans* KCl gradient was 250/50 mmol/l. The membrane vesicles were applied to the *cis* solution and the KCl gradient of 800/50 mmol/l *cis/trans* was used to fuse the vesicle into BLM. Single channel currents were measured by the bilayer clamp amplifier (BC-525C, Warner Instrument, Hamden, CT, USA). They were filtered at low pass filter of 1 kHz and were digitized at a sampling rate of 4 kHz using a DigiData 1200 digitizer (Axon Instruments, Foster City, CA, USA). Data were stored in a computer by means of pClamp5 software (Axon Instruments), which was also used for processing of the data (Malekova et al. 2009). The stored garlic extract (100 µl) was applied to the *cis* and *trans* solutions to obtain final concentration of 6.4 mg/ml.

Results

NO release by garlic, onion and leek extracts

Garlic, onion and leek extracts released NO from GSNO in a concentration- and pH-dependent manner (Fig. 1). Their

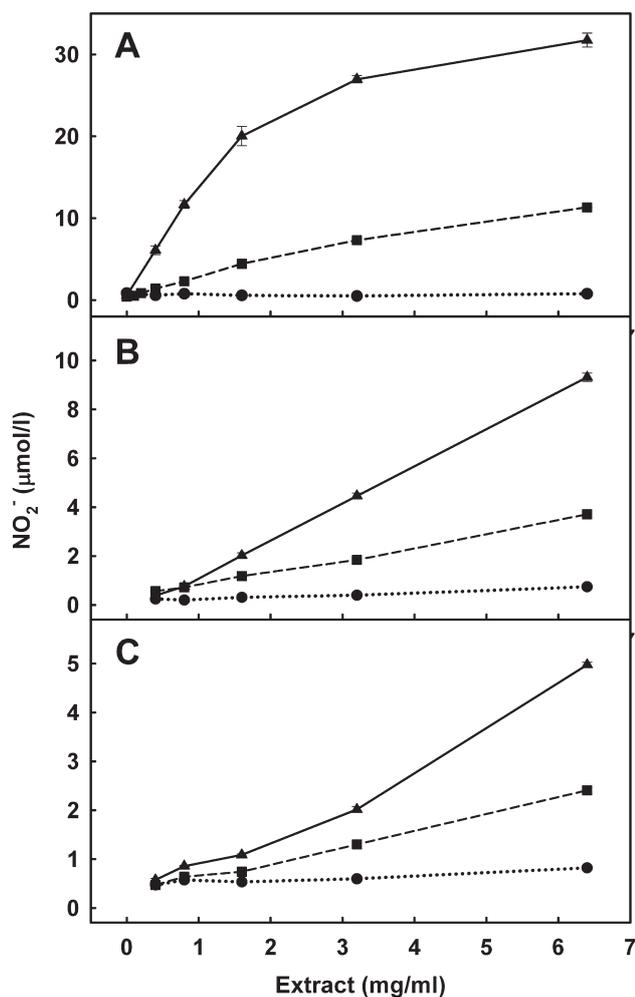


Figure 1. Concentration- and pH-dependent effect of the stored garlic (A), onion (B) and leek (C) extracts on NO release from 100 $\mu\text{mol/l}$ GSNO at pH 6.0 (circles, dotted line), pH 7.4 (squares, dashed line) and pH 8.0 (triangles, full line). NO concentration is indicated by NO_2^- formation according to Griess assay ($n = 3$).

releasing effect was negligible at pH 6.0, but increased at pH 7.4 and was pronounced at pH 8.0. Garlic extract released NO also from NO-donors: S-nitroso-cysteine and S-nitroso-N-acetyl-DL-penicillamine (data not shown). Since garlic extract was the most efficient to release NO, we used it for further studies.

Time dependence of NO release

The potency of the fresh garlic extract to release NO decreased in time. Whether the fresh garlic extract (14.5 mg/ml) after the cloves pulverization was incubated either in the open glass under air at $22 \pm 1^\circ\text{C}$ at the surface : volume ratio of $4.52 \text{ cm}^2 : 1.47 \text{ ml}$, or in closed (1.5 ml volume)

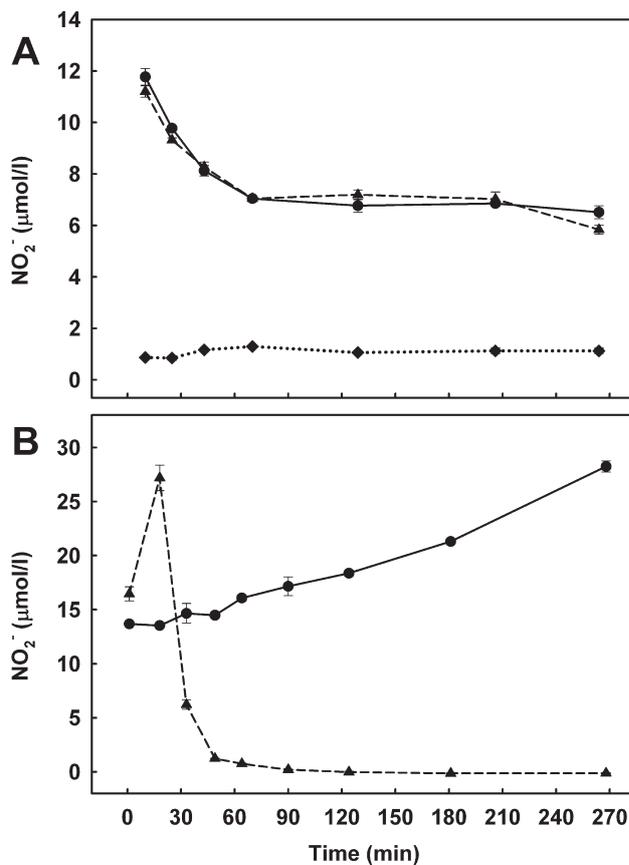


Figure 2. A. Effect of incubation time of the fresh garlic extract (3.7 mg/ml, final concentration) on NO release from 100 $\mu\text{mol/l}$ GSNO. The stock solution of the fresh garlic extract (14.5 mg/ml) was incubated in a closed (1.5 ml volume) ependorf (circles, full line) or under air at $22 \pm 1^\circ\text{C}$ at the surface : volume ratio of $4.52 \text{ cm}^2 : 1.47 \text{ ml}$ (triangles, dashed line). 100 $\mu\text{mol/l}$ GSNO was used as a control (diamond, dotted line). NO concentration is indicated by NO_2^- formation according to Griess assay ($n = 3$). **B.** Effect of incubation time of Na_2S (400 $\mu\text{mol/l}$, final concentration) on NO release from 100 $\mu\text{mol/l}$ GSNO. The fresh Na_2S stock solution (5 mmol/l) was incubated in closed (1.5 ml volume) ependorf (circles, full line) or under air at $22 \pm 1^\circ\text{C}$ at the surface : volume ratio of $4.52 \text{ cm}^2 : 1.47 \text{ ml}$ (triangles, dashed line). NO concentration is indicated by NO_2^- formation according to Griess assay ($n = 3$).

ependorf, the efficiency to release NO decreased similarly in both samples during time to 64% after 70 minutes of the incubation. Later, the efficiency was constant for at least 270 minutes (Fig. 2A). To evaluate whether H_2S as a possible component of the garlic extract was responsible for the NO release, the H_2S donor Na_2S was incubated at the same conditions as garlic extract and its potency to release NO was measured. The time dependency of NO release by garlic extract and by H_2S donor Na_2S was different. When

Na₂S was incubated in the open glass under air its potency to release NO increased at ~18 min of the incubation and later gradually decreased to zero during ~60 min. However, when the Na₂S solution (1.5 ml) was incubated in a full closed ependorf, the Na₂S efficiency to release NO gradually increased with time (Fig. 2B).

Modulation of NO release by thiols

The low molecular thiols (oxidised glutathione (GSSG), reduced glutathione (GSH), N-acetylcysteine (NAC), Cys) significantly modulated garlic extract-induced NO release from GSNO. Their potency depended on their chemical structure and concentrations of the thiols. At 100 µmol/l thiols concentration, the order of the potency to increase the NO release induced by garlic extract was the following: Cys >> GSH = NAC > GSSG. At higher concentration (500 µmol/l), the order of the potency was Cys >> GSSG, whereas GSH had no effect and NAC decreased the NO release with comparison to the NO release induced by garlic extract alone (Fig. 3).

Potentiation of GSNO-induced relaxation of aortic rings by garlic extract

To test biological significance of garlic extract-induced NO release, we studied its effect on the NA-precontracted

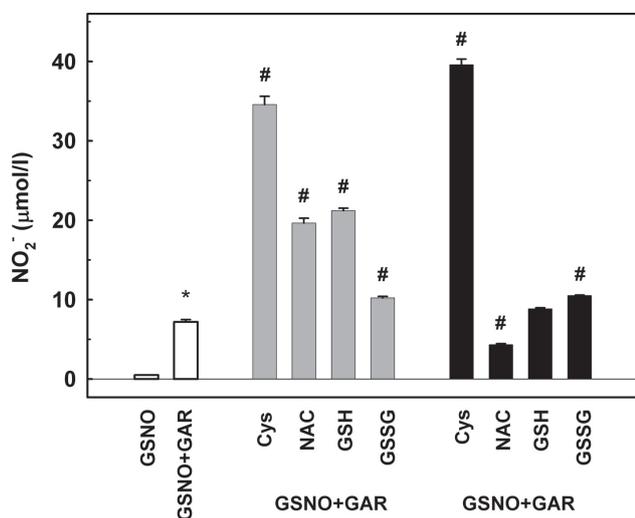


Figure 3. Comparison of the effects of thiols on the garlic extract induced NO release from GSNO. Effect of the stored garlic extract (GAR, 3.7 mg/ml) on NO release from 100 µmol/l GSNO alone (white) and in the presence of Cys, NAC, GSH and GSSG at 100 µmol/l (gray) and 500 µmol/l (black) concentrations. NO concentration is indicated by NO₂⁻ formation according to Griess assay ($n = 3$, \pm SEM). * $p < 0.05$ versus GSNO; # $p < 0.05$ versus GSNO+GAR).

aortic rings. GSNO (50 nmol/l) or the stored garlic extract (45 µg/ml) had small relaxation effects ($22.4 \pm 2.8\%$, $n = 32$ and $5.4 \pm 5.0\%$, $n = 8$, respectively), but the presence of the garlic extract influenced the relaxation induced by GSNO. The relaxation effect of GSNO in the presence of garlic extract was significantly prolonged in time with the comparison to the effect of GSNO alone (Fig. 4).

Garlic extract inhibited chloride channels

Previously we reported that H₂S donor NaHS, which has ability to release NO from GSNO, inhibited activity of intracellular chloride channels (Malekova et al. 2009). To test a hypothesis that the ability of a compound to release NO from NO-donors and to block chloride channels is related and whether intracellular chloride channels are involved in garlic biological effects, we tested the property of garlic extract to modulate activity of the chloride channels. Garlic extract (6.4 mg/ml) perturbed the single channel behaviour (4 out of 5 experiments) 6 minutes after addition and closed the channels (3 out of 5 experiments) 10 minutes after addition (Fig. 5).

Discussion

Considering that garlic extract released NO from compounds with different chemical structures: GSNO, S-nitroso-cysteine and S-nitroso-N-acetyl-DL-penicillamine, we may assume that garlic extract may be able to release NO from other

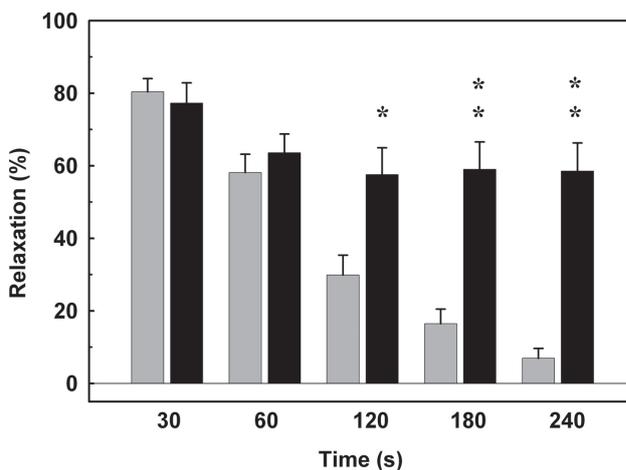


Figure 4. Relaxation effect of 50 nmol/l GSNO (gray) and GSNO in the presence of the garlic extract (45 µg/ml) (black) on the NA (1 µmol/l) precontracted aortic rings measured at different times after GSNO and the extract application. The percent of relaxation is normalized to the maximum relaxation peak obtained by GSNO (100%) or GSNO in the presence of the garlic extract (100%). * $p < 0.01$, ** $p < 0.001$.

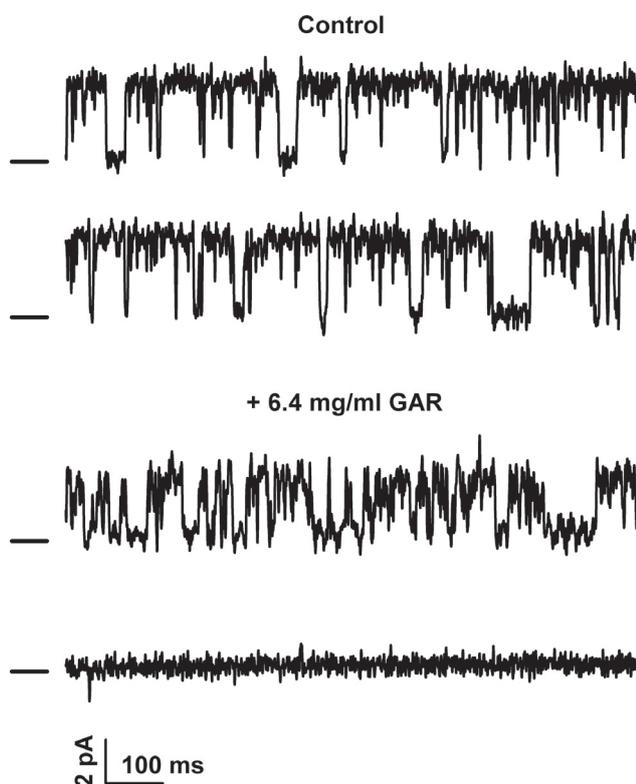


Figure 5. Representative traces of the effect of garlic extract on the single chloride channel current. The application of 6.4 mg/ml garlic extract perturbed single channel current 6 minutes after application and closed the channel 10 minutes after application. The voltage is 0 mV. The lines on the left mark the closed state of the channels.

NO-donors, too. The different NO-releasing potency of the extracts (garlic > onion > leek) (Fig. 1) roughly correlates with their beneficial biological effects, which are most pronounced in garlic extract (Singh and Singh 2008; Iciek et al. 2009; Ginter and Simko 2010; Rahimi et al. 2010). The pronounced pH dependency of garlic, onion and leek extracts ability to release NO (see Fig. 1), is similar to pH-dependency of the ability of H₂S donor NaHS to release NO from GSNO (Ondrias et al. 2008). Based on this analogy we may speculate that pK_a values of garlic extract compounds responsible for the release are about pH 7, and that an active moiety of the compounds may be R-SH group.

It was reported that human red blood cells (RBC) convert garlic-derived organic polysulfides into H₂S and that H₂S production depended on reduced thiols (Benavides et al. 2007). Further, H₂S was found to release NO from NO-donors (Ondrias et al. 2008; Teng et al. 2008; Bertova et al. 2010). Therefore we tested whether H₂S as a possible component of garlic extract is responsible for the NO release. The time dependence of NO release by fresh garlic extract was significantly different from time dependence

of the NO release caused by the H₂S donor Na₂S (Fig. 2). This may indicate that the fresh garlic extract did not contain H₂S responsible for the NO release. Such explanation is supported by the study of Benavides et al. (2007), who did not detect H₂S in garlic extract. However, we cannot exclude a production of H₂S as an intermediate reaction product during chemical reactions in fresh garlic extract. The time dependence of fresh garlic extract ability to release NO (Fig. 2) may indicate that the fresh garlic extract contained at least two different compounds (groups), which can release NO from NO-donors. The concentration of one of them decreased during the first 70 min of incubation due to a chemical interaction(s) and/or a partial evaporation. The second compound(s) was stable for at least 270 min after the garlic pulverization.

The NO-releasing effect of garlic extract was significantly potentiated by low molecular thiols (Fig. 3) indicating that the thiols reacted with those components of the extract, which released NO. This suggestion is supported by the observation of Benavides et al. (2007), who reported that cellular thiols GSH, Cys and NAC converted garlic-derived organic polysulfides into H₂S (in the order of efficiency GSH > Cys > NAC), which is known to release NO from NO-donors (Ondrias et al. 2008). We may hypothesise that the produced H₂S contribute to the NO release. However, the order of thiols efficiency to produce H₂S (Benavides et al. 2007) was different from the order of their efficiency to increase NO release (Fig. 3). Therefore additional unknown compound(s) and/or reactions may be involved in the increase of the NO release.

Garlic extracts are known to relax precontracted aortic rings (Aqel et al. 1991; Ashraf et al. 2004; Benavides et al. 2007). In our study garlic extract prolonged GSNO-induced relaxation of aortic rings (Fig. 4). This result may indicate a possibility that some compound(s) of garlic extract reacted with GSNO and released NO and/or interacted with the pathway of the NO-induced relaxation. Thus suggestion is supported by recent reports of an interaction of a garlic extract with NO pathways during a smooth muscle relaxation. Two groups demonstrated that garlic exerts its therapeutic effect (e.g. vascular relaxation and reduction of blood pressure) by increasing NO production (Kim-Park and Ku 2000; Morihara et al. 2002).

Garlic extract, H₂S or chloride channels have been reported to be involved in blood pressure regulation, muscle tone and apoptosis (Aqel et al. 1991; Puljak and Kilic 2006; Su et al. 2006; Lowicka and Beltowski 2007; Singh and Singh 2008; Ginter and Simko 2010). H₂S was reported to inhibit the activity of chloride channels (Malekova et al. 2009). Therefore we tested whether intracellular chloride channels could be involved in garlic biological effects. We demonstrated modulation of electrical properties of intracellular chloride channel by garlic extract (Fig. 5). Therefore we

suggest that a garlic extract-chloride channel interaction may play a role in garlic biological effects.

Garlic extract has a complex composition (Rose et al. 2005; Jacob et al. 2008; Jacob and Anwar 2008; Iciek et al. 2009). Garlic contains at least 100 sulphur-containing compounds relevant for applications (Singh and Singh 2008). Thiosulphinates released from crushed garlic are reactive molecules and undergo a number of transformations, depending on the temperature, pH and solvent conditions (Rose et al. 2005; Jacob et al. 2008; Jacob and Anwar 2008; Iciek et al. 2009). Since plasma or intracellular concentrations of Cys or GSH are comparable to those used in our study (Davis et al. 2006), we may suppose, Cys-garlic and GSH-garlic interactions *in vivo* may be physiologically relevant. Still, garlic, onion and leek extracts may contain other unknown compounds, which can release NO from NO-donors alone or in the presence of thiols. We suggest that NO-releasing properties of garlic, onion and leek compounds may be involved in the NO-signalling pathway and through this pathway they may exert some of their biological effects.

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