

## HISTOPATHOLOGICAL STUDY

## Iron-rich complexes in human spleen in hereditary spherocytosis

Biro C<sup>1</sup>, Busikova P<sup>2</sup>, Fujerikova G<sup>3</sup>, El-Hassoun O<sup>3</sup>, Kopaniova A<sup>4</sup>, Caplovicova M<sup>5</sup>, Galfiova P<sup>6</sup>, Sisovsky V<sup>3</sup>, Kopani M<sup>3</sup>, Jakubovsky J<sup>3</sup>

Histopatologia Inc., Bratislava, Slovakia. martin.kopani@fmed.uniba.sk

**Abstract:** *Objective:* Tissue iron plays an important role in the development of certain diseases. Although it is one of biogenic elements, its excess induces the reactive oxygen species (ROS) formation. The aim of the present work is to examine the protection against free or loosely bound iron from the view of morphology and chemical composition of iron-rich complexes in human spleen tissues with hereditary spherocytosis (HS) by scanning and transmission electron microscope with energy-dispersive microanalysis (EDX).

*Results:* The examination of human spleen tissues by scanning and transmission electron microscope showed covering of iron-rich particles. EDX revealed many iron-rich complexes of multi-element composition in HS samples with sulphur and phosphorus as the major elements. Detection was negative in the reference samples.

*Conclusion:* The covering of iron-rich particles can be explained by elimination and isolation of ferritin/iron complexes from surrounding environment to prevent the ROS formation. Sulphur, phosphorus and their compounds are probably the most significant elements that influence the ROS formation (Fig. 5, Ref. 16). Full Text in PDF [www.elis.sk](http://www.elis.sk).  
Key words: iron, spleen, EDX microanalysis, electron microscopy, reactive oxygen species.

Tissue iron plays an important role in the etiology of several diseases. Iron in brain plays a decisive role in the process of aging and neurodegenerative diseases (1). It is one of the biogenic elements and its excess induces oxidative stress in tissues. Our knowledge about the iron presence comes mainly from measurements by nuclear magnetic resonance (NMR, 2). A disadvantage of this method, however, is that it provides only qualitative information about iron in the analyzed tissue. Compared to NMR methods, histochemical methods have a higher specificity and a higher specificity level. However, they do not provide the information about physicochemical properties of iron and its compounds in the examined samples. Another weak point of the histochemical methods is the inability to present iron compounds if their dimensions or content are below the discrimination power of the light microscope.

Iron is present in the human body predominantly in the form of ferritin (3). This protein forms spherical formations of the size about 12 nm. Under physiological conditions, in the spleen old or damaged red blood cells (RBC) are destroyed (4) and subsequently engulfed by its lysosomes of macrophages (5). Hereditary spherocytosis is a spherocytic malformation of the red blood cells. The shape abnormalities cause a defect in spectrin-a RBC membrane protein (6). The change in shape of RBC affects their sequestration

in the spleen. They are mistaken for old or damaged RBC and are phagocytosed and destroyed. Iron accumulation in cords can result in their fibrosis (7). Iron is a prominent inductor of ROS. Their overproduction is believed to play a role in the pathogenesis of many diseases in general (8).

The aim of the present work was to examine the protection against free or loosely bound iron from the view of morphology and chemical composition of iron-rich particles in human spleen tissues with hereditary spherocytosis (HS) by the scanning and transmission electron microscope with energy-dispersive microanalysis (EDX).

## Methods

### Samples

Five samples of human tissues with clinicopathological diagnosis of hereditary spherocytosis were studied by scanning and transmission electron microscope with EDX. As the reference samples, the samples of human spleen after a crash accident were used.

### Scanning electron microscope (SEM) and EDX microanalysis

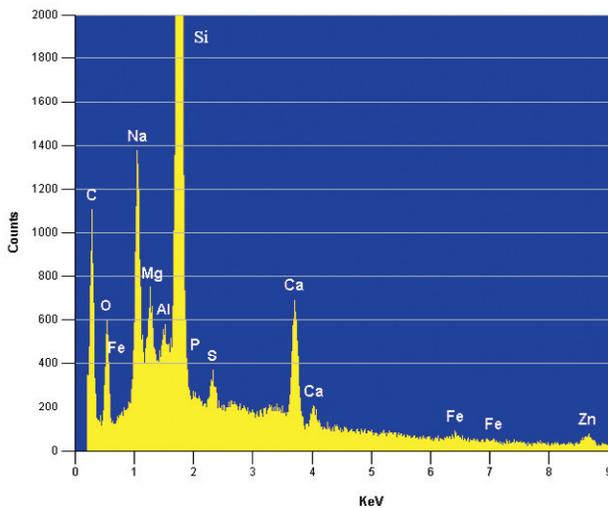
The samples of human tissues were embedded in paraffin blocks, cut by microtome to 5 µm thin sections and mounted on glass support. Unstained and uncovered samples were analyzed by the scanning electron microscope on the instrument JXA 840 A (JEOL, Japan) with the accelerating voltage of 20 kV. Simultaneous EDX analysis was performed with KEVEX 3205-1200 (Kevex, Valencia, Ca). The time period of spectrum collection was 200 s with the energy range 0.160 to 9 keV.

### Transmission electron microscope (TEM)

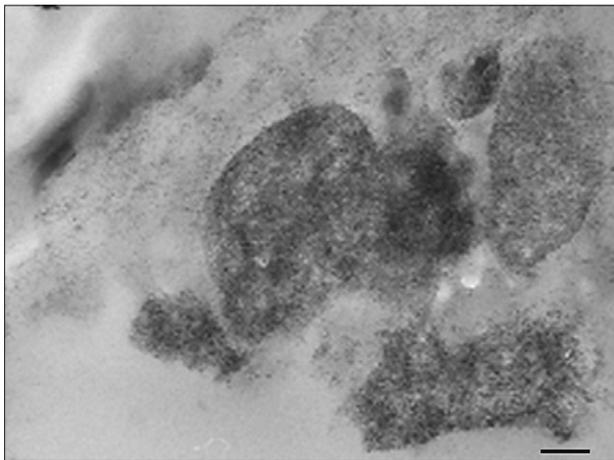
The samples for the transmission electron microscopy study were fixed in 3% solution of glutar(di)aldehyde (SERVA, Hei-

<sup>1</sup>Histopatologia Inc., Bratislava, <sup>2</sup>Histopathology Inc., Bratislava, <sup>3</sup>Comenius University, Faculty of Medicine, Department of Pathology, Bratislava, <sup>4</sup>2nd Department of Neurology, Comenius University, Faculty of Medicine, Bratislava, <sup>5</sup>Department of Geology of Mineral Deposits, Faculty of Natural Science, Comenius University, Bratislava, and <sup>6</sup>Department of Histology and Embryology, Comenius University, School of Medicine, Bratislava, Slovakia

**Address for correspondence:** C. Biro, MD, Histopatologia Inc., Antolska 11, SK-851 07 Bratislava 5, Slovakia.



**Fig. 1.** Human spleen, reference sample, EDX microanalysis. Spectrum showed the element composition with various concentration of C, O, Na, Mg, Al, Si, S, Ca, Fe and Zn. The presence of silicon and sodium in the sample is caused by a glass support. Carbon-coated, energy range 0.160–9 keV, collection time 200 s.

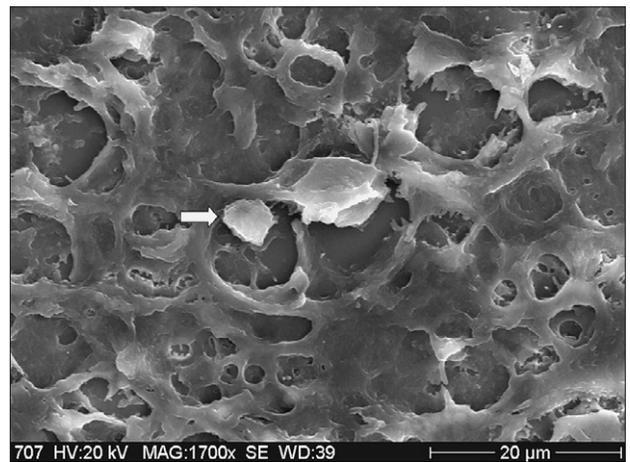


**Fig. 2.** Human spleen, reference sample. TEM micrograph reveal a cluster of round shape particles of ferritin. EDX of this cluster revealed O and Fe. Scale bar = 100 nm.

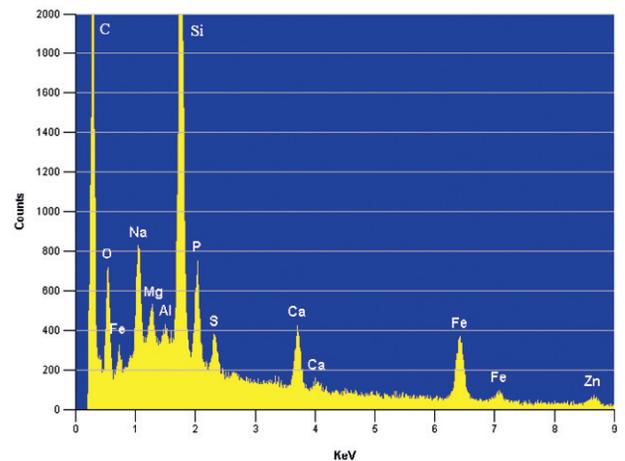
delberg, Germany) for two hours and buffered by phosphate (pH 7.2–7.4). After dehydrating the tissue by alcohol, samples were embedded into Durcupan ACM (Fluka AG, Busch, Switzerland) as recommended by the manufacturer and cut by ultramicrotome (C. Reichert, Wien, Austria). The thickness of samples was 200 nm. Non-contrasted ultrathin sections were mounted on the nickel grids and studied by the transmission electron microscope CM 100 (Philips, Eindhoven, Netherlands) with an acceleration voltage of 120 kV. To find the iron-rich complexes, chemical analysis by EDX was applied.

## Results

Reference samples contained no iron-rich complexes. EDX microanalysis revealed a multi-element composition. Carbon,



**Fig. 3.** Human spleen, hereditary spherocytosis. Histological section. The sharp-edged iron-rich complex with an irregular shape (arrow). Scanning electron microscopy. Scale bar: 20 μm.



**Fig. 4.** Human spleen, hereditary spherocytosis, EDX microanalysis. Spectrum showed a composition of iron-rich complex in human spleen consisting of C, O, Na, Mg, Al, Si, P, S, Ca and Zn. The presence of silicon and sodium in the sample is caused by a glass support. Carbon-coated, energy range 0.160–9 keV, collection time 200 s.

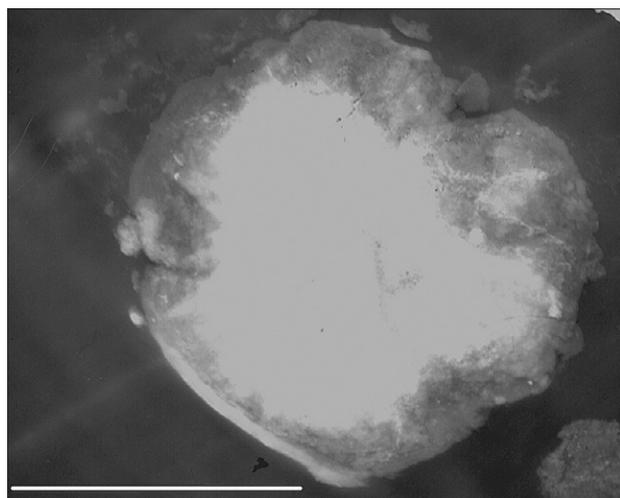
oxygen, sodium, magnesium, aluminium, silicon, sulphur, calcium and zinc were identified (Fig. 1).

Transmission electron micrographs of reference samples revealed ferritin particles, sometimes of their clusters (Fig. 2). EDX of ferritin clusters revealed O and Fe.

Sharp-edged complexes with a high concentration of iron (sometimes in the shape of a block) with dimensions of about 5–7 μm were observed by scanning electron microscopy in samples with HS (Fig. 3).

EDX microanalysis revealed a multi-element composition of these complexes. In addition to iron, carbon, oxygen, sodium, magnesium, aluminium, silicon, phosphorus, sulphur, calcium and zinc was identified (Fig. 4).

The transmission electron micrographs of HS samples revealed bumpy, solid particles of platy, well-defined crystal faces (Fig. 5). EDX of this crystal revealed besides O and Fe also Si, Ti, Cu, Cr,



**Fig. 5. Human spleen, hereditary spherocytosis. TEM micrograph revealed a bumpy, irregular particle. Sometimes, rounded shape particles were seen. The shape of smaller particles was more regular. EDX of this particle revealed besides O and Fe also Si, Ti, Cu, Cr, Pb. Scale bar = 3  $\mu$ m.**

Pb. Sometimes there were irregular, rounded shape particles seen. The shape of the smaller particles was more regular.

## Discussion

Under physiological conditions, ferritin forms spherical formations of the size about 12 nm. The core of this protein consists of iron and oxygen. Ferritin is the iron storage protein found in cells of human spleen. Its physiological function causes the presence of iron in spleen. Zinc is one of the essential dietary factors and physiological element. A decline of zinc level can contribute to metabolic diseases, high blood pressure or diminished immune response (Tubek, 2007, Maggini et al, 2007). Vayenas et al. (1998) examined changes in the concentration of zinc, manganese, and copper in the liver, spleen, and brain of rats after an iron overload by atomic absorption spectrophotometry. They found an increased amount of iron, zinc and manganese in the liver and spleen. They explained the increased amount of these elements by an increased uptake from the cell.

Hereditary spherocytosis affects sequestration of RBC and their accumulation in the spleen. Numerous macrophages can be found and the sinus lining cells are hypertrophied. In certain circumstances, a lot of ferritin is seen both intra and extracytoplasmically (7). Free or loosely bound iron is a substantial inducer of ROS via the Fenton reaction (12). The ferric iron Fe (III) is permanently reduced to ferrous iron Fe (II) and vice versa to form ROS. They are able to damage lipids, DNA and proteins (13). There exists an effect against ROS formation by elimination and isolation of ferritin/iron complexes from surrounding environment by their covering. This protection effect can be seen from the size and EDX microanalysis of iron-rich complexes. SEM revealed around 10  $\mu$ m iron-rich complexes, but the size of iron-containing particles detected by TEM is around 3  $\mu$ m. The rest of complexes are organic compounds that prevent ROS formation. Compounds containing the thiol group exhibit a high radical-trapping activity

(14). Iron overload leads to oxidative stress and subsequently to the regulation of antioxidant defensive mechanisms involving thiol metabolism that may play a major role in the resistance to iron-induced oxidative damage (15). By comparison of the results from EDX microanalysis of reference and HS samples, the presence of phosphorus in the HS sample can be seen. Phosphorus and its compounds significantly affect the morphology and redox activity of iron what can markedly influence the ROS formation. In addition, trace amounts of other elements significantly affect the structure, chemical composition and stoichiometry of iron oxides (16).

## References

1. Zecca L, Youdim MB, Riederer P et al. Iron, brain ageing and neurodegenerative disorders. *Nat Rev Neurosci* 2004; 5: 863–873.
2. Weir MP, Gibson JF, Peters TJ. Effect of dexamethasone on glutamine-synthetase and glial fibrillary acidic protein in normal and transformed astrocytes. *Biochem J* 1984; 223: 31–38.
3. Quintana C, Cowley JM, Marhic C. Electron nanodiffraction and high-resolution electron microscopy studies of the structure and composition of physiological and pathological ferritin. *J Struct Biol* 2004; 147: 166–178.
4. Hromec A, Jakubovsky J. The role of the spleen in erythrocyte destruction. *Cesk Fysiol* 1983; 32: 438–442.
5. Isom HC, McDevitt EI, Moon MS. Elevated hepatic iron: a confounding factor in chronic hepatitis C. *Biochim Biophys Acta* 2009; 1790: 650–662.
6. Boyd AS. Hereditary spherocytosis. *Am Fam Physician* 1989; 39: 167–172.
7. Gregor P, Hromec A, Jakubovsky J et al. The spleen in hereditary spherocytosis. *Cesk Patol* 1996; 32: 7–11.
8. McIntyre JA, Wagenknecht DR, Ramsey CJ. Redox-reactive antiphospholipid antibody differences between serum from Alzheimer's patients and age-matched controls. *Autoimmun* 2009; 42: 646–652.
9. Tubek S. Role of zinc in regulation of arterial blood pressure and in the etiopathogenesis of arterial hypertension. *Biol Trace Elem Res* 2007; 117: 39–51.
10. Maggini S, Wintergerst ES, Beveridge S et al. Selected vitamins and trace elements support immune function by strengthening epithelial barriers and cellular and humoral immune responses. *Br J Nutr* 2007; 98: S29–S35.
11. Vayenas DV, Repanti M, Vassilopoulos A et al. Influence of iron overload on manganese, zinc, and copper concentration in rat tissues in vivo: study of liver, spleen, and brain. *Intern J Clin Lab Res* 1998; 28: 183–186.
12. Valko M, Morris H, Cronin MT. Metals, toxicity and oxidative stress. *Curr Med Chem* 2005; 12: 1161–1208.
13. Reif DW. Ferritin as a source of iron for oxidative damage. *Free Radic Biol Med* 1992; 12: 417–427.
14. Giri SN, Leonard S, Shi X et al. Effects of pirfenidone on the generation of reactive oxygen species in vitro. *J Environ Pathol Toxicol Oncol* 1999; 18: 169–177.
15. Brown KE, Dennery PA, Ridnour LA et al. Effect of iron overload and dietary fat on indices of oxidative stress and hepatic fibrogenesis in rats. *Liver International* 2003; 23: 232–242.
16. Borch T, Masue Y, Kukkadapu RK et al. Phosphate imposed limitations on biological reduction and alteration of ferrihydrite. *Environ Sci Technol* 2007; 41: 166–172.

Received November 10, 2010.

Accepted December 13, 2011.