Serum biomarkers analyzed by LC-MS/MS as predictors for short outcome of non-small cell lung cancer patients treated with chemoradiotherapy

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Received February 16, 2012 / Accepted May 18, 2012

Purpose: To find potential serum biomarkers that can predict clinical outcome upon treatment in non-small cell lung cancer (NSCLC) by analyzing differential proteins in serum with different sensitivity of chemoradiotherapy (CRT).

Materials and Methods: Sera were collected from 37 NSCLC patients before they were treated with concurrent cisplatin-based CRT. According to the outcome of CRT, the patients were divided into sensitive group and resistant one. The proteins in sera were separated by two-dimensional gel electrophoresis after the high abundance proteins were removed from sera. The significantly differentially expressed proteins between two groups were analyzed by liquid chromatography and tandem mass spectrometry (LC-MS/MS). Then, an additional 50 serum samples were used for ELISA analysis to validate the identified proteins that we got in the experiment.

Results: Proteins in sera of each group were successfully separated on 2D gels. There were significant discrepancies in serum protein expression between NSCLC patients with different CRT sensitivity. Among eight differently expressed proteins, six proteins were successfully identified with five of which higher expressed and one lower expressed in resistant group. The increased Alpha-1-antitrypsin (α1-AT) level in resistant group comparing to sensitive one were validated by ELISA analysis (p<0.05).

Conclusions: Proteomic approach may serve as a useful method in detecting the potential biomarkers for predicting the outcome of treatment in NSCLC patients. NSCLC patients with high α1-AT level in serum before CRT may have a worse treatment outcome.

Key words: chemoradiotherapy, non-small cell lung cancer, biomarkers, proteomics

Lung cancer is the leading cause of cancer death in the world, and 80-85% of lung cancer cases are classified as non-small cell lung cancer (NSCLC)[1]. Standard treatment for NSCLC involves platinum-based combination chemotherapy with concurrent radiotherapy which could improve survival for some patients. However, only a minority of patients respond to the chemoradiotherapy (CRT) at the cost of substantial toxicity for all treated patients[2]. There is an urgent need for methods to predict outcome in order to select patients who are likely to benefit from treatment. Oncologists have attempted to identify clinical, functional imaging and molecular markers that may help clinicians and researchers distinguish the subgroup of CRT resistant NSCLC patients. However, few factors have been widely accepted as useful predictive markers for an individual patient undergoing CRT.

Secreted proteins derived from cancer tissues in blood may be the potential serologic biomarkers of cancer. However, serum proteins are difficult to be analyzed as potential biomarkers because of their complexity and dynamic range, which spans ten orders of magnitude. Also, abundant serum proteins, such as albumin, IgG and transferrin mask many of the low abundant proteins that represent putative biomarkers. It is necessary to globally analyze the secreted proteins that would provide new clues to identify potential serologic biomarkers of lung cancer. Proteomics is a relatively new approach for analysis of complex biological specimens, including serum[3], urine[4] and tissue[5]. Mass spectrom-
etry (MS) has shown promise in biomarker discovery, it will potentially benefit the patients from such therapies.

We hypothesized that specific serum biomarkers could predict clinical outcome of patients who underwent CRT. Data analyses of serum peptide profiles between subgroups of patients with different CRT sensitivity were conducted. The goal was to identify potential serum biomarkers that can predict the outcome of NSCLC patients treated with standard CRT.

**Patients and methods**

**Patients and CRT.** Pre-treatment serum samples were collected from 37 patients with stage IIIB or IV NSCLC between September 2009 and March 2011 at the Shandong Cancer Hospital, Jinan, China. Studies were performed after obtaining patient consent and under protocols approved by the institutional review board. Patient characteristics were listed in Table 1. All patients were treated with cisplatin-based combination chemotherapy and concurrent radiotherapy. Radiation therapy was delivered up to a total dose of 62.4 to 68.0 Gy using intensity-modulated radiation therapy or conformal radiation therapy technique. Clinical endpoints used to establish these signatures were response to treatment according to the Response Evaluation Criteria in Solid Tumors (RECIST) criteria at 4 weeks after treatment. Patients with outcome of a complete response (CR) or a partial response (PR) were subsequently classified as sensitive group. However, those who got an outcome of stable disease (SD) or progressive disease (PD) were defined as resistant group.

**Serum samples.** Serum samples were collected by venipuncture and 5 milliliters of whole blood were collected from NSCLC patients before CRT. Venous blood samples were taken in the morning’s fasting state, then centrifuged at 20°C for 10 min at 1,500g between half an hour to two hours. The sera were stored in plastic vials at -80°C until the consecutive analyses. Abundant serum proteins such as albumin, IgG, IgA, and transferrin were removed with Aurum Serum Protein Kit (Bio-Rad, USA).

**Two-dimensional electrophoresis (2-DE).** 350μl mixed serum samples of sensitive and resistant groups underwent 2-DE, and each experiment was repeated 3 times. Isoelectric focusing electrophoresis (IEF) was performed using IPGStrip (pH 3–10L) on IPGphor isoelectric focusing cell (Amersham pharmacia Biotech, Sweden). Second-dimension SDS-PAGE (Bio-Rad, USA) was conducted as described by manufacturer and Görg[6]. After electrophoresis, the protein spots were visualized by silver-based staining technique with the protein silver stain kit (Amersham pharmacia Biotech, Sweden).

**Image analysis.** The 2-DE images were captured using ImageScanner (Amersham Pharmacia Biotech, Sweden). Spot detection, quantification and alignment were performed with the ImageMaster 2D Elite 4.01 software (Amersham Pharmacia Biotech, Sweden). Intensity of each spot was quantified by calculation spot volume after normalization of the image using the total spot volume normalization method multiplied by the total area of all the spots. The reproducibility of spot position was calculated with Gorbett’s method[7]. Molecular mass weight and pI values were determined according to standard protein markers covering the range of 10–100 KDa and the immobilized pH gradient strips. Statistical analysis was carried out with SPSS for Windows 13.0 and Excel.

**LC-MS/MS analysis.** The differentially expressed protein patterns in the CBB staining were the same as that of 2-DE gel visualized by silver-based staining. The protein band containing the desired biomarker was cut from the gel and subjected to overnight tryptic digestion. The selected protein spots were identified by LC-MS/MS using a nanoiquid chromatography–electrospray ionization–tendem mass spectrometry (nano–LC–ESI–MS/MS) system consisting of an nanoAquity UPLC system (Waters, USA) and Synapt High Definition Mass Spectrometry (Waters, USA) equipped with a nano–ESI source. The peptide solutions were loaded to a C18 trap column (nanoACQUITY) (180μm×20mm×5μm (symmetry)). The flow rate was 15 μL/min. Then the desalted peptides were analyzed by C18 analytical column (nanoACQUITY) (75μm×250mm×1.7μm (symmetry)) at a flow rate of 200 nL/min. The solutions used as the mobile phases were (A) H2O and (B) ACN, and both contained 0.1% v/v formic acid. The flow rate was maintained at 200 nL/min. The gradient was started at 5% B, reached 40% B in 65 min, 85% B in the next 10

Table 1. Clinicopathological features of 37 patients with NSCLC

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Number of cases (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age(y)</td>
<td>56-67.5</td>
</tr>
<tr>
<td>Sex</td>
<td></td>
</tr>
<tr>
<td>male</td>
<td>24 (64.8)</td>
</tr>
<tr>
<td>female</td>
<td>13 (35.2)</td>
</tr>
<tr>
<td>Stage</td>
<td></td>
</tr>
<tr>
<td>IIIA</td>
<td>9 (24.3)</td>
</tr>
<tr>
<td>IIIB</td>
<td>23 (62.2)</td>
</tr>
<tr>
<td>IV</td>
<td>5 (13.5)</td>
</tr>
<tr>
<td>Pathologic type</td>
<td></td>
</tr>
<tr>
<td>Adenocarcinoma</td>
<td>14 (37.8)</td>
</tr>
<tr>
<td>Squamous cell carcinoma</td>
<td>20 (54.1)</td>
</tr>
<tr>
<td>other</td>
<td>3 (8.1)</td>
</tr>
<tr>
<td>Chemotherapy</td>
<td></td>
</tr>
<tr>
<td>cisplatin/gemcitabine</td>
<td>13(35.2)</td>
</tr>
<tr>
<td>cisplatin/docetaxel</td>
<td>9(24.3)</td>
</tr>
<tr>
<td>cisplatin/vinorelbine</td>
<td>9(24.3)</td>
</tr>
<tr>
<td>cisplatin/pemetrexed</td>
<td>6(16.2)</td>
</tr>
<tr>
<td>RECIST</td>
<td></td>
</tr>
<tr>
<td>complete response</td>
<td>6 (16.2)</td>
</tr>
<tr>
<td>partial response</td>
<td>18 (48.7)</td>
</tr>
<tr>
<td>stable disease</td>
<td>11 (29.7)</td>
</tr>
<tr>
<td>progressive disease</td>
<td>2 (5.4)</td>
</tr>
</tbody>
</table>
min, and 100% A in the final 10 min. Data-dependent acquisition mode (m/z 50–2000) was enabled, and each survey MS scan was followed by five MS/MS scans with the 30s dynamic exclusion option on. The nanospray parameters were 2500 V for capillary voltage, 35 V for cone voltage, 90°C for source temperature, and 15 psi collision gas back pressure.

Bioinformatics analysis. All mass spectra were used to search the Swiss-Prot database with the Mascot search engine (www.matrixscience.com). The searching parameter was set up as follows: the taxonomy was selected as homo sapiens; the peptide mass tolerance was 1.0 Da; the MS/MS tolerance was 0.5 Da; the missed cleavage sites were allowed up to 1; the fixed modifications were selected as carbamidomethyl (cysteine); the variable modification was selected as oxidation (methionine) or none. The data on 2-DE images of intergroup and intragroup were analyzed by SPSS (version 16.0) using χ² tests and p-values less than 0.05 were considered statistically significant.

ELISA Analysis. To validate the effect of Alpha-1-antitrypsin (α1-AT) and Apolipoprotein A-1 (ApoA-1) in sera from NSCLC patients before CRT, an additional 50 serum samples were applied for ELISA analysis, 37 from sensitive group with an age range of 38-73 years (median, 56 years), 13 from resistant group with an age range of 29-78 years (median, 51 years). We strictly detected the level of α1-AT and ApoA-1 in sera by ELISA with Human α1-AT /ApoA-1 ELISA Kit according to the manufacturer’s instructions (Rapidbio, USA). Read the optical concentration (O.D.) at 450nm using a microtiter plate reader within 30 minutes. All samples were assayed in triplicate and the results were the mean values of the reading. For each serum sample, O.D. value of the fusion protein. The cutoff value of reactivity was defined as mean O.D. of sample plus 3 folds of standard deviations (SD) of normal sera.

Results

Tumor Response. At 4 weeks after chemotherapy and radiotherapy, those who achieved a CR/PR or a PD/SD were deemed as sensitive group or resistant group respectively on CT assessment by RECIST criteria. Twenty-four patients (6 patients attained CR and 18 had PR) were assessable for response and the overall response rate was 64.9%. In contrast, 11 patients achieved SD and only 2 patients attained PD.

Analysis of the 2-DE image. In order to measure the reproducibility, 2-DE for the sensitive group and resistant group was repeated three times, respectively. The image analyses showed that these 2-DE maps were reproducible. Protein spots in sensitive and resistant group were 825±64 and 845±48, respectively. Comparison of 2-DE images between sensitive and resistant group revealed significant difference (χ²=1.483, P=0.0032). A total of 78 well-resolved and matched spots among three tumor-gels were chosen randomly to calculate the deviation of the spot position. (Fig. 1)

Analysis of the differential protein-spots. The differential protein-spots between sensitive and resistant group were detected by 2-DE gel image analyses software. Eight differential protein-spots (seven spots higher expressed and one spot lower expressed in resistant group, Fig. 2) were detected by methods of computer software combined with manually operation.

Identification of specific protein spots. The resulting protein was determined by comprehensively considering the corresponding experimental pI, Mr, the number of matched peptides, and expression level (Fig. 3). Among eight differently expressed protein spots, six differentially expressed proteins were successfully identified, five of which higher expressed and one lower expressed in resistant group (Table 2).

Serum levels of α1-AT and APOP-1. As shown in Fig. 4, the concentration of ApoA-1 before CRT in sensitive and resistant group were (852.81±371.31) ng/ml and (692.25±299.26) ng/ml, respectively (p>0.05). However, the concentration of α1-AT before CRT was (282.94±143.88) ng/ml in sensitive group and (400.36±152.26) ng/ml in resistant group (p=0.016).

Discussion

This study was prompted by the desire to discover serum biomarker/protein in samples from NSCLC patients that can predict the outcome of CRT. Biomarker discovery has primarily been focused on diagnostic biomarkers, while less attention has been given to predictive biomarkers of therapy.
Figure 1. Two dimensional electrophoretic map of sensitive and resistant group

Figure 2. Differential expression of protein spots over expressed and lower expressed in resistant group
response. One limitation of nearly all anti-cancer therapies is that most exhibits clinical activity in only a subset of patients. The successful development of any drug should be linked to predictors of its efficacy, as these biomarkers would considerably increase the likelihood that an individual patient will benefit. Given the financial and societal costs of treating unselected patients with CRT, the predictive biomarker could be tremendously valuable, thus it is imperative to endeavor to identify biomarkers predictive of treatment benefit. Large number of proteins present in the most important body fluid and serum are useful biomarkers of irregular or pathological conditions (i.e., tumors, modified immune responses, or vascular disease), as these proteins are often differentially expressed in abnormal and normal conditions. Proteomic analyses of serum proteins have been discussed for more than 10 years because serum biomarkers can be analyzed relatively noninvasively, and it continues to be a topic of intensive investigations. There are several advantages to the serum MS profiling described here, especially compared with more labor intensive and technically challenging assays, such as immunohistochemistry, fluorescence in situ hybridization, and PCR, due to the unavailability of high-quality tumor specimens and intratumoral heterogeneity[8,9]. LC-MS/MS has shown promise in biomarker discovery and proved to be a useful method for detection of low abundance proteins[10], potentially allowing the selection of patients who may benefit from such therapies. Early studies using MS to develop proteomic profiles from patient serum, coupled with advanced data mining algorithms had triggered hopes that this technology could be used for cancer diagnosis, prognosis, and disease/treatment monitoring. In a study by Stuart Salmon et al., a predictive MALDI-TOF-MS-based proteomic algorithm for “good” or “poor” clinical outcome upon epidermal growth factor tyrosine kinase inhibitor (EGFR-TKI) therapy was established[11]. However, in that study none of the peptides corresponding to the classifying m/z features were identified.

This study was exploratory based on the hypothesis that serum protein may reliably predict the outcome of patients treated with cisplatin-based CRT. To avoid systematic or procedural bias in our study, the positions on the plates were randomized, and the sample spectrometry
was replicated a total of three times and on several days. Six differential proteins were identified by LC-MS/MS coupled to bioinformatics pattern discovery to predict treatment outcome, including five proteins having higher expression and the other one lower expression in patients with resistant tumor. Some of the identified proteins with differential expression are structural proteins (TMC5B and GFAP), and others are regulatory proteins of cell apoptosis (POTE E) and signal transcription (TXLNG)[12-15]. We do believe that informative molecules originating from tumor cells or their microenvironment may indeed be present in biological fluids and that their identification may lead to the discovery of potential new biomarkers. But, it is highly unlikely that a small and localized tumor will be able to modify the serum proteomic pattern to a degree that can be recognized by the proteomic technologies based on MS. The identification of these molecules will likely require ultrasensitive techniques, capable of measuring concentration ranges 10^{-12} -10^{-13} mol/liter or lower (far lower than those achieved by current MS).

An alternative hypothesis for the observed differences in proteomic patterns in serum between different treatment sensitive patients may be the detection of high-abundance molecules that are not produced by the tumor cells but rather represent epiphenomena of tumor presence or treatment reaction. For example, it has been postulated by Diamondis et al. that at least some of the detected molecules represent acute-phase reactants that are released into the circulation by the liver and other organs[16]. It has been shown as early as 30–40 years ago that such molecules are not specific for the presence of any cancer, and for this reason they have not been used in clinical practice for cancer diagnosis or monitoring, although their concentrations may be elevated in serum of treatment sensitive/resistant patients.

Based on the second hypothesis, we elaborate on two potential candidate biomarkers, ApoA-1 and α1-AT. Given the flood of new targets identified as potential biomarkers, it would greatly strengthen the study to show subsequent validation of these two, especially as they are commercially available ELISA tests for them. ApoA-1 is the major protein component of high-concentration lipoprotein (HDL) and a key element of the reverse cholesterol transport pathway, a process that protects against the development of atherosclerosis[17]. Previous studies have reported the diagnostic value of ApoA-1 and proApolipoprotein A1 (the precursor of ApoA-1) in tumor diagnosis. Decreased levels of ApoA-1 were found in a variety of cancers[18-23], but pro-ApoA-1 increased in breast cancer[24]. Marchi et al. found that pro-ApoA-1 significantly increased (p<0.05) in lung cancer patients with brain metastases[25]. The identified proteins that we got have been validated in this experiment. Although the concentration of ApoA-1 in resistant group was lower than that in sensitive group, no significant difference between the two groups was found. α1-AT is one of the most important extra-cellular serine protease inhibitors. The major physiological role of α1-AT in the lung is to bind and inhibit elastase released from leucocytes in the lower respiratory tract, thereby preventing the destruction of lung tissue[26,27]. Elevation of α1-AT serum levels has been observed in the course of a large number of malignant diseases. Mean serum levels of α1-AT were found to be elevated in NSCLC patients and significantly correlated with the stage of cancer[28]. It was previously reported that the serum concentration of α1-AT increased with tumor growth and could be utilized following tumor resection as an indicator of relapse[29,30]. The prognostic significance of α1-AT expression in lung adenocarcinoma has been evaluated using immunohistochemistry; strongly α1-AT positive cases had a worse prognosis than weak-to-moderately α1-AT-positive or α1-AT-negative cases, suggesting that increased α1-AT expression in lung adenocarcinoma patients may be a prognostic indicator[31]. The level of α1-AT in resistant group was higher than that in sensitive group in our experiments (p<0.05). So, our study confirmed that NSCLC patients with higher α1-AT level in serum before CRT may have a worse treatment outcome. In other words, serum α1-AT is a predictive biomarker for outcome of NSCLC patients treated with CRT.

The study still has some limitations. We do not have a blind test set of patients for verifying. Additionally, due to the lack of reproducibility, procedural bias, and virtually no independent confirmation, protein identification and detection of potential disease biomarkers in human serum are still big challenges for MS and bioinformatics analysis[32]. For example, the reproducibility of 2-DE profiles was very central to the key point. As variation in experimental procedures is a common source of variability in 2-DE patterns, 2-DE gel patterns were not easily reproducible between laboratories, making validation difficult. MS is a largely qualitative technique. Relationship between peak height and molecule abundance is not linear and could be very complex. Discriminating peaks identified by different investigators for the same disease are different. (Diamandis, EP. 2003)

In summary, the results presented here provide clues in identifying new serologic predictive biomarkers, demonstrate whether NSCLC patients could benefit from CRT or not, and might bear further investigation. Serum α1-AT was proved by our experiments as a predictive biomarker for outcome of NSCLC patients treated with CRT. Further rigorous testing in a prospective clinical trial setting can determine whether any candidate biomarker will have clinical utility regardless of its identification.

Acknowledgements: The authors are grateful to the National Center of Biomedical Analysis, China and professor Yazhou Cui of Shandong Academy of Medical Sciences for the technical guidance.

References

[1] MANEGOLD C, THATCHER N: Survival improvement in thoracic cancer: progress from the last decade and be-


