EXPERIMENTAL STUDY

The effect of diosmin on pancreatic injury induced by hepatic ischemia reperfusion in rats

Serin Kilicoglu S1, Tanrikulu Y2, Kismet K3, Devrim E4, Erel S3, Sen Tanrikulu C5, Aydogan A3, Tasova V3, Sabuncuoglu MZ3, Kilicoglu B3

Department of Histology and Embriology, Ufuk University Faculty of Medicine, Cankaya, Ankara, Turkey. kiliicoglu belts@yahoo.com

Abstract: Background: Hepatic ischemia-reperfusion causes histological injury to the pancreatic cells during transplantation, trauma and emergency surgery. We investigated the effects of diosmin, a phlebotrophic drug with antioxidant and antiinflammatory effects, on pancreatic injury in the experimental liver ischemia-reperfusion model. Materials and methods: Forty rats were divided into the four groups: sham (Group 1), control (Group 2), preoperative diosmin (50 mg/kg) treatment (Group 3) and preoperative 10-day diosmin (50 mg/kg) treatment group (Group 4). Ischemia-reperfusion model was carried out by clamping the hepatic pedicle for 60 min and then reperfusing the liver for 90 min. At the end of the procedures, blood and pancreas tissue samples were obtained for biochemical and histopathological assessment. Results: According to the results of liver function tests, amylase and the plasma and pancreatic tissue oxidative stress parameters, there was a significant difference between the control and other groups. Histopathologically; the specimens of the Group 2 showed specific morphological abnormalities. The groups 3 and 4 showed the pancreas histomorphology similar to the sham group. Pathological scores were significantly different between the Group 2 and other groups. Conclusions: Diosmin can be administered for a protection of destructive effects of hepatic ischemia–reperfusion injury on pancreas both emergent and elective hepatic surgical operations in which possible ischemic periods were expected (Fig. 1, Tab. 3, Ref. 33). Full Text in PDF www.elis.sk.

Key words: diosmin, liver, ischemia-reperfusion, pancreatic injury.

Hepatic ischemia-reperfusion injury is ordinarily at the time of the repair of extensive liver trauma, shock, tumour resection and liver transplantation (1). Prolonged organ ischemia is characterized by a lack of tissue oxygen. The several situations occur at the lack of tissue oxygen such as adenosine triphosphate (ATP) depletion, derangements in calcium homeostasis, conversion of cellular metabolism to anaerobic pathways, loss of cellular function and cell death. However, once the blood flow is re-established, reperfusion enhances the injury caused by the ischemic period, aggravating the damage caused at the cellular level. Ischemia-reperfusion injury in various organs was recognized as a complex phenomenon. The various factors play a role in the pathogenesis of ischemia reperfusion injury such as increased inflammatory response, reactive oxygen species, cytokine release and neutrophil activation (2, 3).

There are many techniques for liver inflow occlusion during hepatectomy; continuous or intermittent Pringle manoeuvre is used most frequently (4). The Pringle manoeuvre induces not only a stagnation of the portal venous blood, thereby increasing portal venous pressure, but also an ischemic liver damage. There is an increasing evidence that the changes in the portal circulation during hepatic ischemia reperfusion also causes damages in other organs, such as the intestine, lung, heart (5–7). In addition, an increased portal venous pressure may induce pancreatitis and contribute to mortality (8). Nevertheless, the way how the pancreas responds to congestion and acute elevation of portal venous pressure by portal vein occlusion is not well understood (9, 10).

Furthermore, the relationship between liver resection and hyperamylasemia are examined. Hyperamylasemia is a well-documented sequel of liver surgery, and has been demonstrated following other types of abdominal surgery and even extra-abdominal surgery (5, 6). In addition, hyperamylasemia occurs in the process of an acute pancreatitis and in the postoperative period of various types of surgery, including hepatic surgery (8, 11, 12).

Diosmin is an antioxidant, phlebotrophic and vascular protective flavonoid, which increases venous tone, improves lymphatic drainage and reduces capillary hyperpermeability. It also inhibits the activation, migration and adhesion of leukocytes, which leads to a reduction in the release of inflammatory mediators and...
thereby a reduction in capillary hyperpermeability. The antioxidant activity of diosmin has been shown in many studies (13–15).

In the present study, we investigated the effects of diosmin on pancreatic injury in the experimental liver ischemia-reperfusion model.

Materials and methods

Animals
Forty Wistar-Albino female rats, weighing 250 ± 30 g, were housed under a constant temperature (21 ± 2 °C) individually in wire cages with 12 h light-dark cycle, fed water and rodent chow ad libitum. Twelve hours before anaesthesia, animals were deprived of food, but had free access to water 2 h before anaesthesia. No enteral or parenteral antibiotics were administered. The procedures in this experimental study were performed in accordance with the National Guidelines for The Use and Care of Laboratory Animals and approved by the Animal Ethics Committee of Ankara Research and Training Hospital.

Study groups and operative procedure
Rats were randomly divided into the four groups each including 10 animals: SHAM group, Control group (ischemia-reperfusion) and treatment groups (peroperative group and preoperative group). Animals were anaesthetized by intramuscular injection of 80 mg/kg ketamine hydrochloride (Ketalar; Parke- Davis, Istanbul, Turkey) and 20 mg/kg xylasine (Rompun, Bayer, Istanbul, Turkey). After the abdomen was shaved and disinfected, a midline incision was made and rats underwent either sham surgery or ischemia-reperfusion. Ischemia was carried out by clamping for 60 min with a microvascular “bulldog” clamp of hepatic pedicle. After the ischemic period, liver was reperfused by opening the clamp, and reperfusion was achieved for 90 min. In the peroperative treatment group, diosmin was given to the rats, just after ischemia induction at a dose of 50 mg/kg in gavage form with nasogastric tube; therefore the peak plasma levels of diosmin were achieved before reperfusion was initiated. In the preoperative treatment group, 50 mg/kg/day diosmin (Vendios; Bilim, Istanbul, Turkey) was given to the rats one dose daily for ten days before the operation, in gavage form with nasogastric tube (7 Gauge feeding tube) that was inserted daily and taken off after drug administration.

At the end of the procedures, blood and pancreas tissue samples were obtained for biochemical and histopathological assessment.

Biochemical analyses
Plasma alanine aminotransferase (ALT), aspartate aminotransferase (AST) and amylase levels were measured for evaluating the liver functions by using Olympus Au 640 autoanalyzer. To assess oxidative injury, malondialdehyde (MDA) levels, glutathione peroxidase (GSH-Px) xanthine oxidase (XO) enzyme activities were determined in the blood samples and malondialdehyde (MDA) levels and glutathione peroxidase (GSH-Px) enzyme activities were determined in the pancreas samples.

Evaluation of oxidative stress
After sacrifice of the animals, pancreas samples were removed and kept on an ice bath until homogenization. The sample of pancreas was first washed with distilled water, the tissues were homogenized in (20 % w/v, approximately 1 g in 5 ml for each) physiological saline, then they were centrifuged 4000 x g for 15 min and upper clear supernatants were used in the assays. All the procedures were performed at +4 °C throughout the experiments. Protein level of the clear supernatants was studied by Lowry’s method (16), and then they were adjusted to equal concentrations before the other analyses. All the results were expressed as unit/mg protein for those of pancreas tissues. Malondialdehyde (MDA) level (nmol/mg) and glutathione peroxidase (GSH-Px) enzyme activity (mU/mg) were measured in the supernatants.

MDA level was measured by thiobarbituric acid reactive substances method. GSH-Px activity was measured by following changes in NADPH absorbance at 340 nm.

Histological evaluation
For the light microscopic analyses, the samples obtained from the pancreas were fixed in 10% neutral buffered formaline solution for 2 days. Tissues were washed in flowing water and were dehydrated with rising concentrations of ethanol (50 %, 75 %, 96 %, 100 %). After dehydration, specimens were put into xylene to obtain transparency and were then infiltrated with and embedded in paraffin. Embedded tissues were cut into sections of 5 μm thickness by Leica RM 2125 RT and stained with hematoxylin and eosin. Histopathologic examinations were performed by two histopathologists, blinded to the study design. Photographs were taken by Olympus BX-51 microscope with Olympus DP-71 photo attachment. The histological grading of edema was made using a scale ranging from 0 to 3 (0 = no edema, 1 = interlobular edema, 2 = interlobular and moderate intralobular edema, and 3 = interlobular edema and severe intralobular edema). Leukocytic infiltration was also graded from 0 to 3 (0 = absent, 1 = scarce perivascular infiltration, 2 = moderate perivascular and scarce diffuse infiltration, 3 = abundant diffuse infiltration). Grading of vacuolization was based on the appropriate percentage of acinar cells involved: 0 = absent, 1 = less than 25 %, 2 = 25–50 % and 3 = more than 50 % of acinar cells. Haemorrhage was graded as: 0 = no haemorrhage, 1 = 1–2 haemorrhagic foci per slide, 2 = 3–5 haemorrhagic foci per slide, 3 = more than 5 haemorrhagic foci per slide. Necrosis was graded as: 0 = no necrosis, 1 = less than 15 % of pancreatic cells involved, 2 = 15–35 % of cells involved, 3 = more than 35 % of cells involved.

Statistical analysis
Data analysis was performed using the SPSS 15.0 package program. Data were presented as the mean ± standard deviation. Any possible differences among the groups were evaluated by the One-Way ANOVA or Kruskal Wallis variance analysis, where appropriate. When the p-value from the variance analysis was statistically significant, the Mann–Whitney U multiple comparison test was used to know which group differs from the others. Furthermore, the Student’s t-test variance analysis was used for
Tab. 1. Liver function tests.

<table>
<thead>
<tr>
<th>Groups</th>
<th>AST (U/L)</th>
<th>ALT (U/L)</th>
<th>Amylase (U/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>SHAM Group</td>
<td>97.60±18.15*</td>
<td>36.60 ± 3.13*</td>
<td>1879.00 ± 184.18**</td>
</tr>
<tr>
<td>Control Group</td>
<td>465.60 ± 43.49</td>
<td>201.20 ± 49.49</td>
<td>2119.80 ± 156.00</td>
</tr>
<tr>
<td>Peroperative Treatment Group</td>
<td>318.60 ± 108.32*</td>
<td>159.40 ± 83.59*</td>
<td>1830.50 ± 353.07*</td>
</tr>
<tr>
<td>Preoperative Treatment Group</td>
<td>282.60 ± 38.02*</td>
<td>78.40 ± 16.04*</td>
<td>1547.50 ± 195.01*</td>
</tr>
</tbody>
</table>

Abbreviations: ALT – alanine aminotransferase, AST – aspartate aminotransferase, LDH – lactate dehydrogenase, (*) Amylase results were diluted 5 times, * p < 0.001 and ** p < 0.001 vs control, # p < 0.05 vs control, µ p = 0.001 and vs control, (κ) p < 0.05 and vs perop.treatment group, (ν) p < 0.05 vs SHAM group

Tab. 2. Plasma oxidative stress activities of the groups.

<table>
<thead>
<tr>
<th>Groups</th>
<th>MDA (unit/mL)</th>
<th>GSH-Px (unit/mL)</th>
<th>XO (unit/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>SHAM Group</td>
<td>1.72 ± 0.27*</td>
<td>452.06 ± 8.67*</td>
<td>0.51 ± 0.02*</td>
</tr>
<tr>
<td>Control Group</td>
<td>3.60 ± 0.88</td>
<td>307.02 ± 6.48</td>
<td>0.65 ± 0.04</td>
</tr>
<tr>
<td>Peroperative Treatment Group</td>
<td>1.93 ± 0.12*</td>
<td>377.96 ± 6.51*</td>
<td>0.59 ± 0.04*</td>
</tr>
<tr>
<td>Preoperative Treatment Group</td>
<td>1.86 ± 0.43*</td>
<td>424.06 ± 6.93*</td>
<td>0.56 ± 0.01*</td>
</tr>
</tbody>
</table>

Abbreviations: MDA – malondialdehyde, GSH-Px – glutathione peroxidase enzyme activity, XO – xanthine oxidase enzyme activity, * p < 0.05 vs control, µ p = 0.001 vs control, (κ) p < 0.05 vs perop. treatment group

Tab. 3. Tissue oxidative stress activities of the groups.

<table>
<thead>
<tr>
<th>Groups</th>
<th>MDA (nmol/mg)</th>
<th>GSH-Px (mlL/mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>SHAM Group</td>
<td>0.31±0.07</td>
<td>36.33±17.11</td>
</tr>
<tr>
<td>Control Group</td>
<td>0.47±0.13</td>
<td>24.39±5.68</td>
</tr>
<tr>
<td>Peroperative Treatment Group</td>
<td>0.32±0.14*</td>
<td>38.21±8.89**</td>
</tr>
<tr>
<td>Preoperative Treatment Group</td>
<td>0.31±0.07c</td>
<td>33.46±9.00c</td>
</tr>
</tbody>
</table>

Abbreviations: MDA – malondialdehyde, GSH-Px – glutathione peroxidase enzyme activity, * p < 0.05 and ** p = 0.001 vs control, µ p < 0.05 vs control, (κ) p < 0.05 vs perop. treatment group

the evaluation of the histopathological results. All results were accepted as statistically significant when p<0.05.

Results

All rats were sacrificed on postoperative day 10. No rat died during the experimental period.

Biochemical analyses

The results of liver function tests of the groups are summarized in the Table 1. There was a significant difference between the SHAM group and the Control group according to the levels of AST, ALT and amylase (p < 0.001 for AST and ALT, p = 0.001 for amylase). There was a significant difference between the Control group and the Peroperative treatment group according to the levels of AST, ALT and amylase (p < 0.05 for all). There was a significant difference between the Control group and the Preoperative treatment group according to the levels of AST, ALT and amylase (p < 0.001 for all). There was a significant difference between the Sham group and the Peroperative treatment group according to the levels of AST and ALT (p < 0.001). There was a significant difference between the Sham group and the Peroperative treatment group according to the levels of AST and ALT (p < 0.001). There was no significant difference between the Sham group and the treatment groups according to the MDA, GSH-Px and XO levels (p > 0.05 for all).

The pancreas tissue levels of MDA and GSH-Px are summarized in the Table 3. There was not a statistically significant difference between the SHAM group and the Control group according to MDA and GSH-Px levels (p > 0.05 for all). There was a statistically significant difference between the Control group and the Peroperative treatment group according to MDA and GSH-Px levels (p < 0.05 for MDA, p = 0.001 for GSH-Px). There was not a statistically significant difference between the Control group and the Peroperative treatment group according to MDA and XO levels (p > 0.05). GSH-Px levels were higher in the Preoperative treatment group than in the Peroperative treatment group and the difference was significant (p < 0.001). There was not a significant difference between the Sham group and the treatments groups according to the MDA, GSH-Px and XO levels (p > 0.05 for all).

The pancreas tissue levels of MDA and GSH-Px are summarized in the Table 3. There was not a statistically significant difference between the SHAM group and the Control group according to MDA and GSH-Px levels (p > 0.05 for all). There was a statistically significant difference between the Control group and the Peroperative treatment group according to MDA and GSH-Px levels (p < 0.05 for MDA, p = 0.001 for GSH-Px). There was not a statistically significant difference between the Control group and the Peroperative treatment group according to MDA and XO levels (p > 0.05). GSH-Px levels were higher in the Peroperative treatment group than in the Control group and the difference was significant (p < 0.05). There was a statistically significant difference between the Peroperative treatment group and the Preoperative treatment group according to MDA levels (p < 0.05). GSH-Px levels were higher in the Peroperative treatment group than in the Preoperative treatment group and the difference was not significant (p = 0.596).

Oxidative stress

The plasma levels of MDA, GSH-Px and XO are summarized in the Table 2. There was a significant difference between the Control group and other groups according to the MDA, GSH-Px and XO levels (p < 0.05 for all). There was not a significant difference between the Peroperative treatment group and the Preoperative treatment group according to the MDA and XO levels (p > 0.05), GSH-Px levels were higher in the Preoperative treatment group than in the Peroperative treatment group and the difference was significant (p < 0.001). There was not a significant difference between the Sham group and the Peroperative treatment group according to MDA and GSH-Px levels (p > 0.05). There was a statistically significant difference between the Peroperative treatment group and the Preoperative treatment group according to MDA levels (p < 0.05). GSH-Px levels were higher in the Peroperative treatment group than in the Preoperative treatment group and the difference was not significant (p = 0.596).

Histopathological results

The pancreas of the sham operated animals showed no tissue alteration on light microscopy in rats (Fig. 1A). The Control group (ischemia-reperfusion) showed interlobular edema and there was a moderate intralobular edema in four rats. We observed 1–2 hemorrhagic foci per slide (Fig. 1B1). The leukocytic infiltration was
Ischemia-reperfusion injury is now thought to be the main cause. In the early stages of hepatic reperfusion, endothelial cells swell, vessels constrict, leukocytes become entrapped, and platelets aggregate within sinusoids, resulting in microcirculatory failure. There have also been reports of sinusoids failing to refill and inflammatory cytokine production increasing during the reperfusion period (18, 19).

Hepatic ischemia-reperfusion injury involves interaction between different cell types and a variety of cellular and molecular mechanisms including Kupffer cell activation, formation of reactive oxygen species, release of cytokines and chemokines, and neutrophile recruitment (3).

There is an increasing evidence that changes in the portal circulation during hepatic ischemia reperfusion also causes damages in other organs, such as the intestine, lung, heart (5–7). Furthermore, there are published studies that specifically examine the association between pancreatitis and liver resection. In addition, there are studies that have examined the relationship between liver resection and hyperamylasemia, a biochemical marker of pancreatitis. Hyperamylasemia is a well-documented sequel of liver surgery, and has been demonstrated following other types of abdominal surgery and even extra-abdominal surgery (5, 6). Tsuzuki et al (12) observed hyperamylasemia in 70 (39.5 %) of 177 patients who had undergone hepatectomy for hepatocellular carcinoma or metastatic lesions, but they could not elucidate the mechanism responsible. Miyagawa et al (8) found that significant hyperamylasemia following liver resection was associated with prolonged occlusion of the hepatoduodenal ligament, the Pringle manoeuvre.

Many factors, such as an increased portal venous pressure, ischemic liver damage, and reduced functional liver volume may affect amylase metabolism after hepatectomy. It has been reported that the portal venous stasis associated with liver disease may predispose patients to develop pancreatitis (8, 20). An occlusion of the portal vein results in venous congestion in the pancreas and hyperamylasemia and pancreatitis have been induced in an experimental setting by complete occlusion of the pancreatic veins. A prolonged portal congestion also appears to be another considerable problem to be dealt with. Despite some recent attempts to understand it, the contribution of acute portal congestion to hepatic ischemia-reperfusion injury has not yet been fully clarified (21).

In the present study, we evaluated the biochemical analyses by measuring the plasma AST, ALT and amylase. Diosmin treatment was significantly ameliorates these parameters both in preoperative and peroperatively treated groups.
Oxidative stress most likely plays a major role in the early development of acute pancreatitis and several experimental animal models show a beneficial effect of anti-oxidative drugs. Therapy with antioxidants (such as nitric oxide, tetrandrine, L-arginine, allopurinol) administered intravenously has been investigated in a prospective double-blind placebo controlled randomized trial on patients with predicted severe AP but no effect on mortality could be demonstrated. The prophylactic effect on the incidence of post-ERCP pancreatitis was tested in two randomized prospective randomized trials with 256 and 106 patients, respectively. N-acetylcysteine (NAC) was administered before and after ERCP and both studies concluded that NAC was without any preventive effect (22, 23).

Despite advances in experimental studies on the pancreas, the mechanism involved in pancreatic changes resulting in an acute increase of portal pressure secondary to portal venous clamping has not been fully clarified. Aydede et al (24) pointed out that portal bed congestion can trigger an inflammatory response in the pancreas, with a directly proportional relationship between congestion and the changes.

Diosmin is a naturally occurring flavonoid glycoside that can be isolated from various plant sources or derived from the flavonoid hesperidin. Diosmin is considered to be a vascular-protecting agent used to treat chronic venous insufficiency, hemorrhoids, lymphedema, and varicose veins. As a flavonoid, diosmin also exhibits anti-inflammatory, free-radical scavenging, and antimutagenic properties. Diosmin acts by improving venous tone, increasing lymphatic drainage, protecting capillary bed microcirculation, inhibiting inflammatory reactions, and reducing capillary permeability. Micronised purified flavonoid fraction inhibits the activation, migration and adhesion of leukocytes at the capillary level. This leads to a reduction in the release of inflammatory mediators such as oxygen free radicals, prostaglandins and thromboxane, resulting in a decrease in capillary hyperpermeability (25).

Diosmin-hesperidin reported to have radical scavenging properties in the previous reports (26–28). Bouklouki and el al (29) has found that diosmin-hesperidin prevented the reperfusion injury in hamster cheek pouch ischaemia/reperfusion injury model. It was reported that ischaemia/reperfusion induced an increase in the microvascular permeability and the leukocyte adhesion was inhibited by diosmin-hesperidin, which is consistent with an improvement in blood flow through the microvascular network. This protective effect might include a decrease in endothelial cell swelling with a consequent decrease in flow resistance. Another possibility for the observed effects of diosmin-hesperidin is its antioxidant effect (29–32). In another study, Pehlivan et al (33) found that diosmin-hesperidin was effective in preventing intestinal reperfusion injury after an oral administration.

In the present study, we investigated the effect of diosmin on pancreas in the hepatic ischemia-reperfusion injury. There was a significant difference between the control and diosmin groups according to the results of amylase and liver function tests.

The groups treated with diosmin (the Peroperative group and the Preoperative group) showed no histological alteration to the ordinary pancreas tissue. There was no histological evidence of leukocyte infiltration, necrosis and/or vacuolization. We observed no perivascular infiltration or edema in the interlobular/intralobular region of the parenchyma.

In the present study, we evaluated the oxidative stress by measuring the plasma levels of MDA, GSH-Px, XO and pancreas tissue levels of MDA and GSH-Px. Diosmin treatment was also significantly decreased the oxidative stress both in preoperative and peroperatively treated groups.

**Conclusion**

The present study was the first study about the effect of diosmin on pancreas in the hepatic ischemia–reperfusion injury. When we compared the preoperatively and peroperatively treated groups, better results were achieved in the Preoperatively treated group; but the difference was not statistically significant. According to these results, we concluded that diosmin could be administered for the protection of destructive effects of hepatic ischemia–reperfusion injury on pancreas in both emergent and elective hepatic surgical operations, in which possible ischemic periods were expected.

**References**

7. Tanrikulu Y, Kismet K, Serin Kilicoglu S et al. The effect of diosmin on pancreatic...


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