CLINICAL STUDY

Prevalence of mutations in thiopurine S-methyltransferase gene among Slovak IBD patients

Desatova B², Hlavaty T¹, Balakova D³, Pav I², Celec P^{3,4}, Gregus M⁶, Zakuciova M⁷, Hlista M⁸, Horakova M⁹, Kadasi L^{3,5}, Huorka M¹, Batovsky M² (The Slovak IBD study group – SK IBD)

Department of Internal Medicine V, Division of Gastroenterology and Hepatology, University Hospital Bratislava Ruzinov, Slovakia. barboradesat@yahoo.com

Abstract: *Background:* Thiopurine S-methyltransferase (TPMT) plays an important role in the metabolism of thiopurines. It has been suggested that TPMT genetic polymorphisms lead to dose-related hematopoietic toxicity. Since there are major ethnic differences in the prevalence of particular TPMT variants, it is important for each country to study their own prevalence in order to estimate the role of TPMT variants-related thiopurines toxicity in population suffering from particular inflammatory bowel disease (IBD).

Aims: The aim of this study was to determine the frequency of the four most common allelic variants of TPMT gene in the population of Slovak IBD patients.

Methods: TPMT genetic polymorphisms (TPMT*2, TPMT*3A, TPMT*3B, TPMT*3C) were amplified using PCR and consequently genotyped with genetic analyzer. The allele frequencies of particular allelic variants were calculated and compared with other Caucasian populations reported so far.

Results: Three hundred and thirty IBD patients were included; 196/132/2 cases of Crohn's disease/ulcerative colitis/unclassified colitis; 180 (55 %) males. Ninety-three percent of patients were homozygous for wild-type TPMT variant. Heterozygous genotype of any of the studied polymorphisms was present in 6 % of patients while only one patient was homozygous for TPMT*3A allele (0.3 %). The most prevalent mutant allele was that of TPMT*3A (3.2 %). The distribution of most common allelic variants of TPMT gene among Slovak IBD patients was in accordance with previously reported prevalence in Caucasian populations.

Conclusion: This study shows the prevalence of TPMT genetic polymorphisms in population of Slovak IBD patients. As in other Caucasian populations, the most common mutant allelic variant is that of TPMT*3A while the prevalence of homozygosity is relatively low (*Tab. 3, Ref. 22*). Full Text in PDF *www.elis.sk.* Key words: thiopurine S-methyltransferase, thiopurines toxicity, genetic polymorphisms.

Thiopurines such as azathioprine (AZA) and 6-mercaptopurine (6-MP) are immunosuppressive drugs effective in the induction and maintenance of remission of inflammatory bowel disease (IBD) (20). These antimetabolites are involved in the metabolism of nucleic acids. They downregulate their production and have an impact on the activity of lymphocytes. They decrease the proliferation of lymphocytes involved in the inflammation and directly inhibit their cytotoxic activity (15, 16). Thiopurines are used in the treatment of cancer, various autoimmune and chronic inflammatory diseases such as inflammatory bowel diseases, autoimmune hepatitis, autoimmune myasthenia gravis, sclerosis multiplex, psoriasis, systemic lupus erythematosus, primary biliary cirrhosis and rheumatoid arthritis (19, 21). The substance of 6-mercaptopurine was first synthesized in 1951 (15). Azathioprine, a derivate of 6-mercaptopurine, was developed in 1957 and has a longer biological half-life. In 1969 it was also used in the treatment of Crohn's disease (15). Thiopurines are inactive pro-drugs that require multi-enzyme activation after their entry into organism. The final products of metabolism of thiopurines are 6-thioguanine nucleotides (6-TGN) which are in addition to the clinical benefit responsible also for side effects of thiopurines (6). 6-TGN act as purine antagonists and downregulate the synthesis of nucleic acids and proteins. They compete with endogenous guanozine triphosphate (GTP) that is an essential part of signaling pathways and the source of energy for cells. In addition they influence the growth and proliferation of T and B lymphocytes and inhibit the activated immune system in IBD patients (1, 12). After oral ingestion and absorption, 90 % of azathioprine is converted to 6-mercaptopurine by nonenzymatic reaction with contribution of glutathione or cystein (19). 6-mercaptopurine is then metabolized by three competitive enzymes, xantin-oxidase, thiopurine S methyltransferase and

¹Department of Internal Medicine V, Division of Gastroenterology and Hepatology, University Hospital Bratislava Ruzinov, Slovakia, ²Department of Gastroenterology, University Hospital Bratislava Petrzalka, Slovakia, ³Department of Molecular Biology, Comenius University, Bratislava, Slovakia, ⁴Institute of Molecular Biomedicine, Comenius University, Bratislava, Slovakia, ⁵Institute of Molecular Physiology and Genetics, Slovak Academy of Sciences, Bratislava, Slovakia, ⁶KM Gastroenterology Centre Nitra, Slovakia, ⁷Department of Internal Medicine I, Division of Gastroenterology and Hepatology, University Hospital Kosice, Slovakia, ⁸Department of Internal Medicine, Hospital Trencin, Slovakia, and ⁹Department of Internal Medicine II, Division of Gastroenterology and Hepatology, University Hospital Martin, Slovakia

Address for correspondence: B. Desatova MD, PhD, Department of Gastroenterology, University hospital Bratislava, Antolska 11, SK-851 07 Bratislava, Slovakia. Phone: +421.911540280

hypoxantin-guanin phosphoribozyltransferase (5, 19). The drug response and also side effects vary among individuals. Age, gender, disease activity, co-morbidity but also inherited signs like single nucleotide polymorphisms (SNPs) are responsible for these differences. SNPs are simple nucleotide variations in DNA localized anywhere in the genome, and they are the most common cause of inter-individual differences in the reaction of organism to the drug.

Thiopurine S-methyltransferase (TPMT) is a cytosolic enzyme that plays an important role in the metabolism of thiopurines. It catalyses the thiopurines S-methylation (4, 17, 22). TPMT enzyme activity is inherited as an autosomal codominant sign and is under the control of genetic polymorphisms that have been extensively studied in the past (21). It has been suggested that TPMT genetic polymorphisms are associated with reduced TPMT enzyme activity that can lead to dose-related hematopoietic toxicity in patients treated with thiopurines. TPMT enzyme is encoded by an approximately 34 kb gene located on chromosome 6 (6p22.3) and contains 10 exons and 9 introns (3, 22). To date, at least 24 mutant alleles are known and have been reported in association with reduced TPMT enzyme activity (3). The most common mutant alleles such as TPMT*3A, TPMT*2, TPMT*3C and TPMT*3B are detected in 80-95 % of the Caucasian population (22). The molecular defect in TPMT*3A allele is caused by two nucleotide transition mutations $(G460 \rightarrow A \text{ and } A719 \rightarrow G)$. The defect in TPMT*2 contains the mutation G238 \rightarrow C. In TPMT*3C allele, it is the mutation A719 \rightarrow G and in TPMT*3B, G460 \rightarrow A. Patients homozygous for wild type TPMT genotype have the TPMT enzyme activity normal or high and are good metabolizers of thiopurines. Patients with both mutant alleles, homozygotes for mutant TPMT genotype have very low TPMT enzyme activity. They are poor metabolizers of thiopurines, which leads to high levels of 6-TGN in organism as well as increases the risk of myelotoxicity. Patients with heterozygous genotype have intermediate enzyme activity and intermediate drug level in organism. In patients with high TPMT enzyme activity, the risk of hepatotoxicity is high because of huge production of methylated mercaptopurine metabolites damaging the liver (1, 12). Ethnic differences between distribution of TPMT mutant alleles have been described (11, 14, 22). The activity in Caucasian population has a trimodal distribution with poor, intermediate and high methylators. Since there are major ethnic differences in the prevalence of particular TPMT variants, it is important for each country to study their own prevalence in order to estimate the role of TPMT variants-related thiopurines toxicity in the particular IBD population.

Material and methods

A total of 330 IBD patients treated in Slovakia between years 2007 and 2009 were included. The only inclusion criterion was the diagnosis of IBD (Crohn's disease, ulcerative colitis or unclassified colitis) established by endoscopic, histological and radiological findings. The mode of therapy was not taken into consideration. The most common TPMT variant alleles TPMT*2, TPMT*3A, TPMT*3B, TPMT*3C were determined. The allele frequencies of particular allelic variants were calculated and compared with other Caucasian populations reported so far. For descriptive statistics and

frequencies SPSS system version 15.0 was used. Genomic DNA was isolated from 1 milliliter of venous blood using the Puregene Blood Core kit (Qiagen, Hilden, Germany). Three different PCR were used for preamplification of fragments containing polymorphisms 460G>A, 719A>G and 238G>C, respectively. Amplifications were performed in a 10 µl reaction volume containing 50 ng of genomic DNA, 1x concentrated 5PRIME HotMasterMix (5PRIME, Hamburg, Germany) and 2.5 pmol of primers. After amplifications, PCR products were used as a template for sequencing the reactions with BigDye Terminator v 3.1 Cycle Sequencing Kit (Applied Biosystems, Foster City, USA). Electrophoresis was carried out with ABI PRISM® 3100 Genetic Analyzer (Applied Biosystems, Foster City, USA) and obtained sequences were compared with the reference sequences using BLAST sequence alignment software.

Results

One hundred and ninety six patients with Crohn's disease, 132 with ulcerative colitis and 2 with unclassified colitis were included in this study. The mean age of included patients was 37 years in range of 17-75 years while 180 (55 %) patients were males and 150 (45%) were females. Ninety three percent of patients were homozygous for wild type TPMT variant (TPMT*1/*1). Heterozygous genotype of any of the studied polymorphisms was present in 6 % of patients, only one patient was homozygous for TPMT*3A allele (0.3 %). The most prevalent mutant allele was that of TPMT*3A (3.2%). The frequency of mutant alleles TPMT*3C and TPMT*2 was 0.2 %. In this studied group, no TPMT*3B mutant allele was detected. The distribution of most common allelic variants of TPMT gene among Slovak IBD patients were in accordance with previously reported high prevalence of TPMT*3A variant and lower frequencies of TPMT*3C and TPMT*2 in Caucasian populations (Tabs 1-3).

Tab. 1. Demographic characteristic of Slovak IBD population.

Demographic Characteristic	Number of patients
Number of patients	330
Male/Female	180/150
Age (mean + range) in years	37 (17-75)
Crohn's disease/Ulcerative colitis/Unclassified colitis	196/132/2

Tab. 2. Prevalence of wild type homozygotes, mutant heterozygotes and mutant homozygotes among Slovak IBD patients.

	Number of patients	%
Number of patients	330	100
Wild type homozygotes (TPMT *1/*1)	308	93
Mutant heterozygotes		
TPMT *2/*1	1	0.3
TPMT *3A/*1	19	5.8
TPMT *3B/*1	0	0
TPMT *3C/*1	1	0.3
Mutant homozygotes	1	0.3
TPMT *2/*2	0	0
TPMT *3A/*3A	1	0.3
TPMT *3B/*3B	0	0
TPMT *3C/*3C	0	0

330-332

Tab. 3. Allelic variants of TPMT gene and their frequencies among Slovak IBD patients.

Allelic variants	Number of alleles	%
Number of alleles	660	100
TPMT *1	637	96.5
TPMT *2	1	0.2
TPMT *3A	21	3.2
TPMT *3B	0	0
TPMT *3C	1	0.2

Discussion

TMPT genotyping is a way of determining a group of IBD patients treated with thiopurines with high risk of serious side effects such as bone marrow toxicity (18). Patients heterozygous or homozygous for mutant TPMT allele (low TPMT activity) are at higher risk of developing severe hematopoietic toxicity while treated with standard dose of thiopurines (13). In our cohort, 93 % of patients were homozygous for wild type TPMT, 6 % had heterozygous genotype of any of the studied polymorphisms, and only one patient had homozygous genotype for mutant variant TPMT*3A. There are big ethnic differences and also an ethnic heterogeneity in distribution of TPMT mutant variants. It is important for each country to determine their own prevalence by reason of estimating the role of TMPT mutant alleles-related thiopurines toxicity in each particular IBD population. This is a TPMT genotype study that shows the distribution of TPMT mutant variants among Slovak IBD patients. The allele frequencies of TPMT mutant allelic variants are comparable with those in other Caucasian and Latin-American population. The most common allelic variant among Slovak IBD patients was TPMT*3A (3.2 %), while TPMT*2 and TPMT*3C were present only rarely (0.2 %). In Chinese, Japanese, Indian (9), Korean (11) and African populations, TMPT*3C is the most prevalent mutant allele (7, 10) while in Latin-American population it was not detected at all (8). On the other hand the TPMT*2 allele is present in 9.4 % of mutant alleles in British population, while being very rare in Slovak IBD patients and not present in any of Ghanaian subjects (2). The TMPT*3B was not detected in Slovak IBD population at all. The mutant allele TPMT*7 has been recently found in the European Caucasian population, TPMT*8 in African Americans, and TPMT*6 in the Korean population (10). They seem to be rare in the Caucasian population, but they have not been investigated in our cohort.

In conclusion, we have determined that the distribution of mutant TPMT variants in Slovak IBD patients are in accordance with previously reported distribution in other Caucasian populations.

References

1. Al Hadithy AF, De Boer NKH, Derijks LJJ, Escher JC, Mulder CJ, Brouwers JR. Thiopurines in inflammatory bowel disease: pharmacogenetics, therapeutic drug monitoring and clinical recommendations. Dig Liver Dis 2005; 37 (4): 282–297.

2. Ameyaw MM, Collie-Duguid ESR, Powrie RH, Ofori-Adjei D, McLeod HL. Thiopurine methyltransferase alleles in British and Ghanaian populations. Hum Mol Genet 1999; 8 (2): 367–370.

3. Cao Q, Zhu Q, Shang Y, Gao M, Si J. Thiopurine Methyltransferase Gene Polymorphisms in Chinese Patients with Inflammatory Bowel Disease. Digestion 2009; 79 (1): 58–63.

4. Egan LJ, Derijks LJJ, Hommes DW. Pharmacogenomics in inflammatory bowel disease. Clin Gastroenterol Hepatol 2006; 4 (1): 21–28.

5. Gisbert JP, Nino P, Rodrigo L, Cara C, Guijarro LG. Thiopurine Methyltransferase (TPMT) Activity and Adverse Effects od Azathioprine in Inflammatory Bowel Disease: Long-Term Follow-Up Study of 394 Patients. Am J Gastroenterol 2006; 101 (12): 2769–2776.

6. Gurwitz D, Rodríguez-Antona C, Payne K et al. Improving pharmacovigilance in Europe: TPMT genotyping and phenotyping in the UK and Spain. Eur J Hum Genet 2009; 17 (8): 991–998.

7. Hon YY, Fessing MY, Pui ChH, Relling MV, Krynetski EY, Evans WE. Polymophism of the thiopurines S-methyltransferase gene in African-Americans. Hum Mol Genet 1999; 8 (2): 371–376.

8. Isaza C, Henao J, López AM, Cacabelos R. Allelic variants of the thiopurine methyltransferase (TPMT) gene in the Colombian population. Methods Find Exp Clin Pharmacol 2003; 25 (6): 423.

9. Kham SKY, Soh ChK, Liu TCh, Chan YH, Ariffin H, Tan PL, Yeoh AE. Thiopurine S-methyltransferase activity in three major Asian populations: a population based study in Singapore. Eur J Clin Pharmacol 2008; 64 (4): 373–379.

10. Kubota T, Chiba K. Frequencies of thiopurne S-methyltransferase mutant alleles (TPMT*2, *3A, *3B and *3C) in 151 healthy Japanese subjects and the inheritance of TPMT*3C in the family of a propositus. Br J Clin Pharmacol 2001; 51 (5): 475–477.

11. Lennard L. TPMT in the treatment of Crohn's disease with azathioprine. Gut 2002; 51 (2): 143–146.

12. Lennard L. The clinical Pharmacology of 6-mercaptopurine. Eur J Clin Pharmacol 1992; 43: 329–339.

13. Leung M, Piatkov I, Rochester C, Boyages SC, Leong RWL. Normal thiopurine methyltransferase phenotype testing in a Crohn disease patient with azathioprine induced myelosuppression. Intern Med J 2009; 39 (2): 121–126.

14. Otterness DM, Szumlanski CL, Wood TC, Weinshilboum RM. Human Thiopurine Methyltransferase Pharmacogenetics. Kindred with a Terminal Exon Splices Junction Mutation That Results in Loss of Activity. J Clin Invest 1998; 101 (5): 1036–1044.

15. Pearson DC, May GR, Fick GH, Sutherland LR. Azathioprine and 6-Mercaptopurine in Crohn Disease. A Meta-Analysis. Ann Intern Med 1995; 123 (2): 132–142.

16. Regueiro M, Mardini H. Determination of Thiopurine Methyltransferase Genotype or Phenotype Optimizes Initial Dosing of Azathioprine for the Treatment of Crohn's disease. J Clin Gastroenterol 2002; 35: 240–244.

17. Takatsu N, Matsui T, Murakami Y et al. Adverse reactions to azathioprine cannot be predicted by thiopurine S-methyltransferase genotype in Japanese patients with inflammatory bowel disease. J Gastroenterol Hepatol 2009; 24 (7): 1258–1264.

18. Teml A, Schaeffeler E, Schwab M. Pretreatment determination of TPMT – state of the art in clinical practice. Eur J Clin Pharmacol 2009; 65 (3): 219–221.

19. Tiede I, Fritz G, Strand S, Poppe D, Dvorsky R, Strand D, Lehr HA, Wirtz S, Becker C, Atreva R, Mudter J, Hildner K, Bartsch B, Holtmann M, Blumberg R, Walczak H, Iven H, Galle PR, Ahmadian MR, Neurath MF. CD28-dependent Rac1 activation is the molecular target of azathioprine in primary human CD4+ T lymphocytes. J Clin Invest 2003; 111 (8): 1133–1145.

20. Winter JW, Gaffney D, Shapiro D et al. Assessment of thiopurine methyltransferase enzyme activity is superior to genotype in predicting myelosuppression following azathioprine therapy in patients with inflammatory bowel disease. Aliment Pharmacol Ther 2007; 25 (9): 1069–1077.

21. Yates ChR, Krynetski EY, Loennechen T. Molecular diagnosis of thiopurine S-methyltrasnferase deficiency: genetic basis for azathioprine and mercaptopurine intolerance. Ann Intern Med 1997; 126 (8): 608–614.

22. Zhou S. Clinical pharmacogenomics of Thiopurine S-methyltransferase. Curr Clin Pharmacol 2006; 1 (1): 119–128.

Received May 23, 2011. Accepted January 27, 2013.