CLINICAL RESEARCH

IL1 cluster gene polymorphisms in macedonian patients with chronic periodontitis

Atanasovska-Stojanovska A1, Popovska M1, Trajkov D2, Spiroski M2

Dental Clinical Center, Department of Oral Pathology and Periodontology, Faculty of Stomatology, University "Ss. Cyril and Methodius", Skopje, Republic of Macedonia. mspiroski@yahoo.com

Abstract: Several studies have investigated the genetic polymorphisms for cytokines as potential genetic markers for periodontitis. The aim of this study was to determine the prevalence of *IL1* cluster genes polymorphisms and their association with chronic periodontitis in the Macedonian population. The group of 114 unrelated Macedonian subjects with chronic periodontitis and 301 periodontitis-free Macedonian subjects were studied. DNA was isolated from peripheral blood leukocytes by phenol-chloroform extraction method. Cytokine genotyping was performed by PCR-SSP. The population genetics analysis package (PyPop) was used for analysis of the cytokine data for this report. Crude odds ratio (OR) was calculated as estimates of the relative risk with 95 % confidence interval (CI). Genotype frequency of *IL1B -511/C:T* was significantly higher in patients with periodontitis than in controls (OR=2.11, 95 % CI=1.35–3.32, p=0.001). *IL1* cluster gene haplotype frequencies of *TTTCT* and *TCTTT* were associated with higher risk for periodontitis (OR=5.06, 95 % CI=1.68–15.26, p<0.0014 and OR=8.35, 95 % CI=1.67–41.69, p<0.002, respectively). No significant association of *IL1* composite genotype (*IL1 -889A:IL1B +3962*) with periodontitis in Macedonians was found. The latter association was found to be significant in genotype (*IL1B -511/C:T*, haplotype *TTTCT*, and haplotype *TCTTT*, but without significant association in *IL1* composite genotype (*Tab. 5, Ref. 43*). Full Text in PDF *www.elis.sk*. Key words: periodontitis, SSP polymorphism, genetic, interleukine-1, Republic of Macedonia.

Periodontitis is a multifactor disease whereby both environmental and genetic factors contribute to its etiology and/or clinical severity. Cytokines are potent immunomodulating molecules that mediate the inflammation and immune response, and at the same time influence the cellular activation, differentiation, and function. There are many reports showing that a number of cytokine genes are polymorphic and that polymorphisms in gene regulatory regions correlate with the cytokine secretion (1, 2). As these polymorphisms are independently segregated, one individual may have a cytokine expression pattern quite different from that of another (3).

Until recently, the *IL1* ligand family consisted of four members: *IL1A, IL1B, IL1RA*, and *IL18*. Based on conservation of amino acid sequence, identification of gene structure and three dimensional structures, six additional members of this family have been described since. The entire new gene map of the region of chromosome 2 between the *IL1B* and *IL1RN* loci was proposed, suggesting that each of the new *IL1* family members arose from a common ancestral gene that later became duplicated (4, 5). The novel *IL1* family members have been described by several groups using their own nomenclature, thus resulting in a number of different names for the same molecule (6, 7).

The prototypic members of the *IL1* family gene cluster are the genes *IL1A* (MIM 147760), *IL1B* (MIM 147720), and *IL1RN*. *IL1A* and *IL1B* encode pro inflammatory cytokines involved in host defense against infection. The IL-1 receptor antagonist, encoded by the gene *IL1RN*, is an anti-inflammatory non-signaling molecule that competes for receptor binding with IL-1A and IL-1B (8, 9). However, hundreds of single nucleotide polymorphisms are known for each gene of the *IL1* cluster (10).

The protein encoded by *IL1R1* gene is a cytokine receptor that belongs to the interleukin 1 receptor family. This protein is a receptor for interleukin 1 alpha (IL-1A), interleukin 1 beta (IL-1B), and interleukin 1 receptor, type I (IL-1R1/IL-1RN). It is an important mediator involved in many cytokine-induced immune and inflammatory responses (11). Five cytokine polymorphisms connected with the *IL1* cluster gene on chromosome 2 were included in the cytokine polymorphism component: *IL1A – 889, IL1B - 511, IL1B + 3962, IL1R psti 1970*, and *IL1RN mspa 11100*. Cytokines are the key factors of mediating the inflammatory process during periodontal disease. Functional polymorphisms of the *IL1* genes have been proposed to be a risk factor for periodontitis. Periodontitis is an infective disease, where chronic inflammation develops under the influence of microorganisms forming the dental plaque.

¹Dental Clinical Center, Department of Oral Pathology and Periodontology, Faculty of Stomatology, University "Ss. Cyril and Methodius", Skopje, Republic of Macedonia, and ²Institute of Immunobiology and Human Genetics, Faculty of Medicine, University "Ss. Cyril and Methodius", Skopje, Republic of Macedonia

Address for correspondence: M. Spiroski, MD, PhD, Institute of Immunobiology and Human Genetics, Faculty of Medicine, University "Ss. Cyril and Methodius", 1109 Skopje, PO Box 60, Republic of Macedonia. Phone: +389.2.3110556, Fax: +389.2.3110558

Acknowledgement: This research is part of the project "Molecular analysis of cytokine gene polymorphisms in the Republic of Macedonia" supported by the Ministry of Education and Science from Republic of Macedonia (Project No. 13-874/3-05). We declare we have no conflict of interest.

The interaction between the host inflammatory response and the bacterial virulent factors destructs the attachment and increase the osteoclastic resorption of the alveolar bone, which as a final effect results in teeth loss (12). It is well documented that their biological activities in vivo are sufficient to produce local inflammation and destruction of the connective tissue and bone (13). IL-1 acts as a signal in many different systems of the body and it strongly affects the progression and severity of periodontitis in adult population. The connection between IL-1 and periodontitis was extensively described by Offenbacher (14). Plates of individual basis of polymorphism (SNPs) of IL1 locus, their functional consequences and their connection with receptiveness and severity of various inflammatory diseases are described in literature (15). The presence of the rare allele T for this polymorphism is connected with several medical conditions as juvenile rheumatoid arthritis (JPA), systematic lupus erythematosus (SLE) ulcerative colitis, myasthenia gravis, alopecia and some forms of diabetes (16).

The aim of this study was to determine the prevalence of *IL1* cluster genes polymorphisms and their association with chronic periodontitis in the Macedonian population.

Material and methods

Healthy subjects were recruited from the patient pool at the Clinic for Oral Diseases and Periodontology, University Clinical Centre of Stomatology, Skopje, Macedonia (17–19). All subjects were over 20 years of age, had at least 20 teeth present, were from Macedonian ethnic background to the third generation, and were unrelated residents from different geographical regions of the Republic of Macedonia. Subjects were excluded if they had any systemic disease, were pregnant, current smokers, or taking medications known to affect the host immunity (steroids, immunosuppressant, etc.).

The periodontitis group consisted of 114 subjects, age 38.97 ± 10.12 years, previously diagnosed with moderate or severe chronic periodontal disease according to established criteria while the healthy control group consisted of 301 subjects, age 35.20 ± 9.90 years, displaying no sites with a probing depth over 3 mm, no clinical attachment loss (CAL) over 2 mm, gingival Loe-Silnes Index (GI) of 1, and bleeding on probing (BOP) $6.8\pm6.4\%$ (20–23). The study was approved by the Committee of the Ministry of Education and Science from the Republic of Macedonia, as well as by the Ethical Committee of the Medical Faculty in Skopje and was part of the Cytokine Polymorphisms and Expression, 15th International Histocompatibility and Immunogenetics Workshop (IHIWS) (24).

Testing Polymorphism on IL1 cluster gene

Genomic DNA was isolated by the phenol-chloroform method from peripheral leukocytes and samples stored in the Macedonian bank for human DNA as previously described (25, 26). *IL1* gene polymorphisms were determined using the PCR-SSP (Heidelberg kit, Cytokine genotyping Tray, *Invitrogen*, GmbH, Karlsruhe, Germany) at the Institute for Immunobiology and Human Genetics at the Faculty of Medicine in Skopje (27). Fourteen cytokine genes with 22 single nucleotide polymorphisms (SNP) were typed: *ILlalpha-889, IL1beta-511, IL1beta+3962, IL1R pst1 1970, IL1RA mspa 11100, IL4Ralpha+1902, IL12-1188, IFN*gamautr5644, *TGFbeta1 cdn10, TGFbeta1 cdn25, TNFalpha-308, TNFalpha -238, IL2-330, IL2+166, IL4-1098, IL4-590, IL4-33, IL6-174, IL6 565, IL10-1082, IL10-819,* and *IL10-592.*

Briefly, the PCR-SSP typing Heidelberg kit consists of 48 PCR primer mixes in aliquot amounts within 96-well PCR trays (two typings per tray). Master mix, which was supplied along with the reagents and consisted of MgCl2, buffer, dNTP's, and glycerol was mixed with 1.2–3.0 μ g DNA and 20 U Taq polymerase (GE Healthcare) and dispensed in 48 wells. The amplified products were separated by 2 % agarose gel electrophoresis, stained with 0.5 μ g/mL ethidium bromide and visualized by ultraviolet exposure.

Statistical analysis

The population genetics analysis package, PyPop, developed by the Biostatistics Core for the Workshop was used for analysis of the IL1 data for this report (28, 29). Allele frequencies and expected Hardy Weinberg proportions (HWP) for each IL1 allele were determined (30). The exact test for genotype frequency deviation from HWP was calculated using the Arlequin implementation accessed via PyPop (31, 32). The alleles that did not fit the HWP were evaluated to determine whether there was an excess of homozygotes or heterozygotes, or if any particular genotypes were significantly different from expected frequencies by the chi square test. The Ewens-Watterson homozygozity test of neutrality (EWN) with Slatkin's exact p-values (SEPV) was used to indicate any deviations from the hypothesis of neutral selection for each locus (33). Pearson's p-values, crude Odds Ratio (OR) and Wald 95 % confidence interval (CI) were calculated for associations analysis between IL1 (alleles, genotypes, haplotypes, and diplotypes) and periodontal disease with Simple Interactive Statistical Analysis (SISA): Two by two table analysis (http:// www.quantitativeskills.com/sisa/). Values for p less than 0.05 were taken as significant.

Results

The clinical and demographic data of our Macedonian study populations are shown in Table 1. As predicted, the values of the clinical parameters GI, CAL, and BOP were higher in the periodontitis group than those in healthy controls.

Tab. 1. Demographic and	periodontal	findings in	1 Macedonian	popu-
lation.				

	Chronic periodontitis (n=114)	Control group (n=301)	р
Age (years) Mean±SD	38.97±10.12	35.20±9.90	NS
Female	41.90 %	54.60 %	NS
Male	58.10 %	46.40 %	NS
Loe-Silnes Index (GI)	2.38±0.67	0.5±0	< 0.001
BOP %	82.52±8.14	6.8±6.4	< 0.001
CAL	5.18±0.716	1.8±0.3	< 0.001

SD - standard deviation; n, number of participants; NS - non significant; BOP - bleeding on probing; CAL - clinical attachment loss.

380-385

Tab. 2. Association of *IL1* gene cluster allele frequency with periodontitis in Macedonian patients.

IL1 Gene Cluster	Allele	PARO		CONTROL		OB	Wald 95%	Pearson's
	Allele	Ν	F	Ν	F	OR	CI	p-value
IL1A -889	C T	176 46	0.793 0.207	482 110	0.814 0.186	0.87	0.59-1.28	0.49
IL1B -511	C T	144 84	0.632 0.368	404 198	0.671 0.329	0.84	0.61-1.15	0.28
IL1B +3962	C T	165 59	0.737 0.263	439 163	0.729 0.271	1.03	0.73-1.46	0.83
IL1R pstl1970	C T	154 72	0.681 0.319	399 203	0.663 0.337	1.08	0.78-1.51	0.61
IL1RN mspa111100	$T \\ C$	168 60	0.737 0.263	420 182	0.698 0.302	1.21	0.86-1.70	0.26

PARO - patients with periodontitis; N - number of alleles; F - frequency of alleles; OR - Odds Ratio; CI - confidence interval.

Tab. 3. Association of IL1 gene cluster genotype frequency with periodontitis in Macedonian patients.

IL1 Gene Cluster	PARO		CONTROL		OD	Wald	Pearson's	
	Genotype -	Ν	F	Ν	F	OR	95% CI	p-value
	C:C	70	63.1	204	68.9	0.77	0.49-1.22	0.26
IL1A -889	C:T	36	32.4	74	25.0	1.44	0.89-2.32	0.13
	T:T	5	4.5	18	6.1	0.73	0.26-2.01	0.54
	C:C	42	36.8	143	47.5	0.64	0.41-1.00	0.051
IL1B -511	C:T	60	52.6	118	39.2	2.11	1.35-3.32	0.001*
	T:T	12	10.5	40	13.3	0.77	0.39-1.52	0.45
	C:C	64	57.1	174	57.8	0.97	0.63-1.51	0.90
IL1B +3962	C:T	37	33.0	91	30.2	1.14	0.72-1.81	0.58
	T:T	11	9.8	36	12.0	0.80	0.39-1.64	0.54
	C:C	54	47.8	133	44.2	1.16	0.75-1.78	0.51
IL1R pstl1970	C:T	46	40.7	133	44.2	0.87	0.56-1.35	0.52
illin powity to	T:T	13	11.5	35	11.6	0.99	0.50-1.94	0.97
	C:C	6	5.3	30	10.0	0.50	0.20-1.24	0.13
IL1RA mspa11100	C:T	48	42.1	122	40.5	1.07	0.69-1.65	0.77
1	T:T	60	52.6	149	49.5	1.13	0.74-1.75	0.57

PARO - patients with periodontitis; N - number of genotypes; F - frequency of genotypes; OR - Odds Ratio; CI - confidence interval; * - statistically significant difference.

The association analysis of *IL1* gene cluster allele frequency with periodontitis in Macedonian patients is given in Table 2 but the differences between patients with periodontitis and control subjects were not significant for any allele.

The association analysis of *IL1* gene cluster genotype frequency with periodontitis in Macedonian patients have shown that the genotype *IL1B -511/C:T* was significantly higher in patients with periodontitis than that in controls (OR 2.11, 95 % CI 1.35–3.32, p=0.001). The rest of *IL1* gene cluster genotypes were non-signficantly associated with periodontitis (Tab. 3).

Association of *IL1* cluster haplotype frequency estimated for loci: *IL1A -889: IL1B -511: IL1B +3962: IL1R psti1079: IL1RN mspa11100* with periodontitis in Macedonian patients have shown that haplotypes *TTTCT* and *TCTTT* were associated with higher risk for periodontitis (OR=5.06, 95 % CI=1.68–15.26, p<0.0014 and OR=8.35, 95 % CI=1.67–41.69, p<0.002, respectively). The rest of *IL1* cluster haplotypes were not significantly associated with periodontitis (Tab. 4).

No significant association of *IL1* composite genotype (*IL1* -889A:*IL1B* +3962) with periodontitis in Macedonians was found (Tab. 5).

Discussion

In this report we presented the association of *IL1* cluster gene (alleles, genotypes, haplotypes and composite genotype) with peridontitis in Macedonian patients. We found a significant association of genotype *IL1B -511/C:T*, haplotype *TTTCT*, and haplotype *TCTTT* with periodontitis. For the rest of *IL1* cluster gene alleles, genotypes, haplotypes and composite genotype we did not find a significant association with periodontitis in Macedonian patients.

Pro-inflammatory cytokine interleukin-1 has a very important role in periodontal tissue destruction by stimulating the bone destruction and participating in the production of proteases and arachidonic acid, i.e. in activities directly connected to periodontitis. The level of IL-1 beta is increased in the gingival tissue of patients with periodontitis (34). The association between a composite genotype of allele 2 of *IL1B* +3953 gene and allele 2 of *IL1A* -889 gene in severe form of adult periodontitis was published (35). Our results of composite *IL1* genotype in Macedonian patients with periodontitis are in agreement with several published papers in which no significant association between composite genotype of *IL1* with periodontitis was found (36–38).

IL1 Gene Cluster Haplotype	PARO		CON	TROL	OR	Wald	Pearson's
	Ν	F	Ν	F		95% CI	p-value
CCCCT	46.9	0.215	113.3	0.191	0.91	0.63-1.33	0.64
CTCCT	36.9	0.169	75.0	0.127	1.12	0.73-1.71	0.60
CCCCC	17.7	0.081	50.1	0.085	0.98	0.55-1.71	0.93
CCCTC	14.8	0.068	22.5	0.038	1.83	0.93-3.57	0.07
CCCTT	14.3	0.065	58.3	0.098	0.63	0.34-1.16	0.13
CTCTT	11.0	0.050	48.5	0.082	0.60	0.31-1.18	0.14
CCTCT	6.5	0.030	28.3	0.048	0.67	0.29-1.55	0.35
TCTCT	5.5	0.025	27.7	0.047	0.47	0.18-1.24	0.12
TCTCC	7.4	0.038	22.0	0.037	0.86	0.36-2.04	0.73
CTCCC	0	0	19.4	0.033	&	&	&
TCCCT	4.0	0.018	17.9	0.030	0.59	0.19-1.78	0.35
TCTTC	0	0	15.8	0.027	&	&	&
CCTCC	7.9	0.036	14.3	0.024	1.57	0.65-3.80	0.31
CTCTC	5.2	0.024	12.1	0.020	1.13	0.39-3.26	0.81
CTTCT	3.8	0.017	11.7	0.020	0.90	0.29-2.83	0.86
CCTTC	0	0	10.1	0.017	&	&	&
CCTTT	5.8	0.026	9.1	0.015	1.83	0.64-5.21	0.25
TCCTC	1.2	0.005	6.5	0.011	0.45	0.05-3.76	0.45
CTTTT	0	0	5.8	0.010	&	&	&
TTTCT	8.5	0.039	4.8	0.008	5.06	1.68-15.26	0.0014*
TTCTT	1.9	0.008	4.6	0.008	1.10	0.21-5.64	0.92
TTCCT	3.9	0.018	3.9	0.007	2.75	0.68-11.08	0.14
TTTCC	0	0	2.6	0.004	&	&	&
CTTTC	2.3	0.011	2.5	0.004	2.73	0.38-19.51	0.29
TTTTC	0	0	2.1	0.004	&	&	&
TCTTT	6.1	0.028	2.1	0.003	8.35	1.67-41.69	0.002*
CTTCC	0	0	2.1	0.002	&	&	&
TTTTT	4.0	0.018	0	0	&	&	&
TTCTC	1.5	0.005	0	0	&	&	&

Tab. 4. Association of *IL1* cluster haplotype frequency estimated for loci: *IL1A*-889: *IL1B*-511: *IL1B*+3962: *IL1R* psti1079: *IL1RN* mspa11100 with periodontitis in Macedonian patients.

Haplotype frequencies were estimated from unphased data using the expectation-maximization (EM) algorithm (29, 32) reported by PyPop. PARO – patients with periodontitis; N – number of haplotypes; F – frequency of haplotypes; OR – Odds Ratio; CI – confidence interval; * statistically significant difference. &, cannot be calculated because expected value was <5, χ^2 test.

Tab. 5. Association of con	mposite genotype (IL)	l -889A:IL1B +3962) wi	th periodontitis in N	Aacedonian patients.

IL1 Polymorphism	Composite PARO		RO	CONTROL			Wald	Pearson's
	genotype*	Ν	F	Ν	F	OR	95% CI	p-value
IL1A -889:	CC	147	0.672	398	0.644	1.02	0.73-1.43	0.89
IL1B +3962	СТ	27	0.120	83	0.140	0.87	0.55-1.39	0.56
	TT	31	0.146	78	0.130	1.10	0.70-1.72	0.68
	TC	12	0.054	73	0.056	0.99	0.50-1.96	0.98

*Comman et al 1997 (35). PARO - patients with periodontitis; N - number of composite genotypes; F - frequency of composite genotypes; OR - Odds Ratio; CI - confidence interval.

A most recent study has shown that genotype 2/2 of *IL1RN* for the whole Brazilian population and allele *T* of *IL1B* (*C*-511*T*) in a subgroup of Afro-Americans and mulattos were suggested to be putative risk indicators for chronic periodontitis (39).

However, a case-control association study on 415 northern European Caucasian patients with aggressive periodontitis and 874 healthy controls was conducted to examine 10 single-nucleotide polymorphisms in the genes of the *IL1* cluster for association with *IL1A*, *IL1B*, *CKAP2L* (cytoskeleton-associated protein 2-like), and *IL1RN* (IL-1 receptor antagonist). The results do not support any association between variants in the *IL1* gene cluster and aggressive periodontitis (40).

The association between *IL1* genes and aggressive periodontitis was investigated using 70 markers spanning the 1.1-Mb region where the *IL1* gene family is mapped. The case-control study included 95 patients and 121 control individuals and explored both the linkage disequilibrium (LD) and the haplotype structure. No association between aggressive periodontitis and *IL1A*, *IL1B*, and *IL1RN* genes was found in either single-point or haplotype analyses, and the study failed to support the existence of a causative variant for generalized aggressive periodontitis within the 2q13-14 region in an Italian Caucasian population (41).

The relationship between specific *IL1* genotypes and level of IL-1 β in the gingival crevicular fluid is unclear. Similarly, the ability of the genetic susceptibility test to forecast which patients will develop increased bleeding on probing, periodontitis, or loss of teeth or dental implants is ambiguous. Additional prospective clinical trials are needed to determine the risk of developing peri-

380-385

odontitis or peri-implantitis when allele 2 at IL1A + 4845 and IL1B + 3954 loci is present (42).

Recently we published significant associations (after the Bonferroni adjustment) between subjects with periodontitis and the following: (1) cytokine alleles IL4-1098 and IL4-33; (2) cytokine genotypes IL4-1098/G:T; IL4-1098/T:T, and IL4-33/T:T, (3) cytokine haplotypes IL4/GCC, IL4/TCC, and IL4/TTC; and (4) cytokine haplotype zygotes IL4/TTC: TCC, IL4/TCT:TTT, and IL4/GCC:TTC. Cytokine polymorphism on IL4 gene appears to be associated with susceptibility to chronic periodontitis in Macedonians (43).

Periodontitis is a complex genetic trait which includes a lot of associated candidate genes. Our results with only one gene cluster (IL1) can be only part of the complex investigation of candidate genes for periodontitis in Macedonians. The numbers of patients and controls of our study is small. In association studies, there are possibilities that some positive results might be spurious and some negative findings might be a consequence of low statistical power. It could be due to their small sample size or methodological shortcomings such as selection of an inappropriate control group. Further studies are merited to assess these associations in greater detail (including any gene-gene and gene-environment interactions) and to determine any implications with regard to potential therapies.

In summary, we can conclude that a significant association of genotype *IL1B -511/C:T*, haplotype *TTTCT*, and haplotype *TCTTT* with periodontitis in Macedonian patients was found, however without a significant association of *IL1* composite genotype. Previous reports of the association between *IL1* composite genotype and periodontitis might reflect the subpopulation effects and have to be interpreted with care.

References

1. Kilpinen S, Huhtala H, Hurme M. The combination of the interleukinlalpha (IL-lalpha-889) genotype and the interleukin-10 (IL-10 ATA) haplotype is associated with increased interleukin-10 (IL-10) plasma levels in healthy individuals. Eur Cytokine Netw 2002; 13 (1): 66–71.

2. Warle MC, Farhan A, Metselaar HJ, Hop WC, Perrey C, Zondervan PE et al. Are cytokine gene polymorphisms related to in vitro cytokine production profiles? Liver Transpl 2003; 9 (2): 170–181.

3. Hoffmann SC, Stanley EM, Cox ED, DiMercurio BS, Koziol DE, Harlan DM et al. Ethnicity greatly influences cytokine gene polymorphism distribution. Am J Transplant 2002; 2 (6): 560–567.

4. Dunn E, Sims JE, Nicklin MJH, O'Neill LAJ. Annotating genes with potential roles in the immune system: six new members of the IL-1 family. Trends Immunol 2001; 22: 533–536.

5. Lin H, Ho AS, Haley-Vicente D, Zhang J, Bernal-Fussell J, Pace AM et al. Cloning and characterization of IL-1HY2, a novel interleukin-1 family member. J Biol Chem 2001; 276 (23): 20597–20602.

6. Smith DE, Renshaw BR, Ketchem RR, Kubin M, Garka KE, Sims JE. Four new members expand the interleukin-1 superfamily. J Biol Chem 2000; 275 (2): 1169–1175.

7. Sims JE, Nicklin MJ, Bazan JF, Barton JL, Busfield SJ, Ford JE et al. A new nomenclature for IL-1-family genes. Trends Immunol 2001; 22 (10): 536–537.

8. Dinarello CA. Biologic basis for interleukin-1 in disease. Blood 1996; 87: 2095–2147.

9. Arend WP, Malyak M, Guthridge CJ, Gabay C. Interleukin-1 receptor antagonist: role in biology. Annu Rev Immunol 1998; 16: 27–55.

10. NCBI dbSNP Build 128. dbSNP development team October 23, 2007. http://www.ncbi.nlm.nih.gov/SNP/. National Center for Biotechnology Information (NCBI), National Library of Medicine, National Institutes of Health, Bethesda, MD, USA.

11. Dale M, Nicklin MJ. Interleukin-1 receptor cluster: gene organization of IL1R2, IL1R1, IL1RL2 (IL-1Rrp2), IL1RL1 (T1/ST2), and IL18R1 (IL-1Rrp) on human chromosome 2q. Genomics 1999; 57: 177–179.

12. Newman M. Genetic, environmental, and behavioral influences on periodontal infections. Compend Contin Educ Dent 1998; 19 (1): 25–31.

13. Christgau M, Aslanidis C, Felden A, Hiller KA, Schmitz G, Schmalz G. Influence of interleukin-1 gene polymorphism on periodontal regeneration in intrabony defects. J Periodontal Res 2003; 38: 20–27.

14. Offenbacher S. Periodontal diseases: pathogenesis. Ann Periodontol 1996; 1: 821–878.

15. Engebretson SP, Grbic JT, Singer R, Lamster IB. GCF IL-1beta profiles in periodontal disease. J Clin Periodontol 2002; 29: 48–53.

16. D'Aiuto F, Parkar M, Brett PM, Ready D, Tonetti MS. Gene polymorphisms in pro-inflammatory cytokines are associated with systemic inflammation in pateints with severe periodontal infections. Cytokine 2004; 28: 29–34.

17. Trajkov D, Arsov T, Petlichkovski A, Strezova A, Efinska-Mladenovska O, Spiroski M. Cytokine gene polymorphisms in population of ethnic Macedonians. Croat Med J 2005; 46: 685–692.

18. Trajkov D, Atanasovska-Stojanovska A, Petlichkovski A, Strezova A, Gogusev J, Hristomanova S et al. IL-1 gene cluster polymorphisms in the Macedonian population. Mac J Med Sci 2008; 1: 21–28.

19. Trajkov D, Arsov T, Petlichkovski A, Strezova A, Efinska-Mladenovska O, Gogusev J et al. Distribution of the 22 cytokine gene polymorphisms in healthy Macedonian population. Bratisl Lek Listy 2009; 110 (1): 7–17.

20. Armitage GC. Development of a classification system for periodontal diseases and conditions. Ann Periodontol 1999; 4 (1): 1–6.

21. Burt B. Research, Science and Therapy Committee of the American Academy of Periodontology. Position paper: epidemiology of periodontal diseases. J Periodontol 2005; 76 (8): 1406–1419.

22. Loe H, Silness J. Periodontal disease in pregnancy. I. Prevalence and severity. Acta Odontol Scand 1963; 21: 533–551.

23. Ramfjord SP. The periodontal disease index (PDI). J Periodontol 1967; 38: 602–610.

24. Atanasovska-Stojanovska A, Trajkov D, Popovska M, Spiroski M. Analysis of transforming growth factor-beta1 gene polymorphisms in Macedonian patients with chronic periodontitis. Mac J Med Sci 2009; 2: 30–35.

25. Towner P. Purification of DNA. 47–54. In: Brown TA (Ed). Essential Molecular Biology Oxford: Oxford University Press, 1995.

26. Spiroski M, Arsov T, Petlichkovski A, Strezova A, Trajkov D, Efinska-Mladenovska O et al. Case Study: Macedonian Human DNA Bank (hDNAMKD) as a source for public health Genetics. 33–44. In: Georgieva L, Burazeri G (Eds). Health Determinants in the Scope of New Public Health. Sofia: Hans Jacobs Company, 2005. **27. Tseng LH, Chen PJ, Lin MT, Singleton K, Martin EG, Yen AH et al.** Simultaneous genotyping of single nucleotide polymorphisms in the IL-1 gene complex by multiplex polymerase chain reaction-restriction fragment length polymorphism. J Immunol Methods 2002; 267: 151–156.

28. Lancaster A, Nelson MP, Meyer D, Thomson G, Single RM. Py-Pop: a software framework for population genomics: analyzing large-scale multi-locus genotype data. Pac Symp Biocomp 2003; 8: 514–525.

29. Lancaster AK, Single RM, Solberg OD, Nelson MP, Thomson G. PyPop update – a software pipeline for large-scale multilocus population genomics. Tissue Antigens 2007; 69 (Suppl 1): 192–197.

30. Single RM, Meyer D, Mack SJ, Lancaster A, Erlich HA, Thomson G. 14th International HLA and Immunogenetics Workshop: report of progress in methodology, data collection, and analyses. Tissue Antigens 2007; 69 (Suppl 1): 185–187.

31. Guo SW, Thompson EA. Performing the exact test of Hardy Weinberg proportion for multiple alleles. Biometrics 1992; 48: 361–372.

32. Schneider S, Roessli D, Excoffier L. Arlequin version 2.000: a software for population genetics data analysis. 2000; Geneva (Switzerland): Genetics and Biometry Laboratory, University of Geneva.

33. Watterson GA. The homozygozity test of neutrality. Genetics 1978; 88: 405–417.

34. Honig J, Rordorf-Adam C, Siegmund C, Wiedemann W, Erard F. Increased interleukin-1 beta (IL-1 beta) concentration in gingival tissue from periodontitis patients. J Periodontal Res 1989; 24: 362–367.

35. Kornman KS, Crane A, Wang HY, di Giovine FS, Newman MG, Pirk FW et al. The interleukin-1 genotype as a severity factor in adult periodontal disease. J Clin Periodontol 1997; 24 (1): 72–77. **36. Hodge PJ, Riggio MP, Kinane DF.** Failure to detect an association with IL1 genotypes in European Caucasians with generalised early onset periodontitis. J Clin Periodontol 2001; 28 (5): 430–436.

37. Sakellari D, Koukoudetsos S, Arsenakis M, Konstantinidis A. Prevalence of IL-1A and IL-1B polymorphisms in a Greek population. J Clin Periodontol 2003; 30: 35–41.

38. Gonzales JR, Mitchel J, Rodriguez EL, Herrmann JM, Bodeker RH, Meyle J. Comparison of interleukin-1 genotypes in two populations with aggressive periodontitis. Eur J of Oral Sci 2003; 111: 395–399.

39. Trevilatto PC, de Souza Pardo AP, Scarel-Caminaga RM, de Brito RB Jr, Alvim-Pereira F, Alvim-Pereira CC et al. Association of IL1 gene polymorphisms with chronic periodontitis in Brazilians. Arch Oral Biol 2011; 56 (1): 54–62.

40. Fiebig A, Jepsen S, Loos BG, Scholz C, Schäfer C, Rühling A et al. Polymorphisms in the interleukin-1 (IL1) gene cluster are not associated with aggressive periodontitis in a large Caucasian population. Genomics 2008; 92 (5): 309–315.

41. Scapoli C, Borzani I, Guarnelli ME, Mamolini E, Annunziata M, Guida L et al. IL-1 gene cluster is not linked to aggressive periodontitis. J Dent Res 2010; 89 (5): 457–461.

42. Greenstein G, Hart TC. Clinical utility of a genetic susceptibility test for severe chronic periodontitis: a critical evaluation. J Am Dent Assoc 2002; 133 (4): 452–459.

43. Atanasovska-Stojanovska A, Trajkov D, Nares S, Angelov N, Spiroski M. IL4 gene polymorphisms and their relation to periodontal disease in a Macedonian population. Hum Immunol 2011; 72 (5): 446–450.

Received July 18, 2011. Accepted January 23, 2013.