

## Long-term effects of somatostatin 14 on the pituitary-ovarian axis in rats

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**Abstract.** The long-term effects of somatostatin 14 (SST-14) on the pituitary-ovarian axis were examined. Female Wistar rats received 20 µg/100 g b.w. doses subcutaneously twice daily for 5 consecutive days in the infantile (from 11<sup>th</sup> to 15<sup>th</sup> day) or peripubertal (from 33<sup>rd</sup> to 37<sup>th</sup> day) period of life. Females treated as infants were killed in the peripubertal (38<sup>th</sup> day) or adult period of life (80<sup>th</sup> day), and those treated during peripuberty as adults (80<sup>th</sup> day). Pituitary follicle-stimulating (FSH), luteinizing (LH) and somatotropic (GH) cells, and ovaries were analyzed by stereology and morphometry. Serum FSH and LH concentrations were determined by RIA. FSH and LH cell volumes were significantly decreased in pituitaries of peripubertal females treated with SST-14 as infants, and in adult females treated during peripuberty. GH cell volume was decreased in all treated rats. In the ovaries, enlargement of the non-growing pool of follicles was detected in adult females treated during peripuberty. SST-14 applied to infant rats did not lead to changes in initial follicular recruitment, but it disturbed follicle growth and development at later stages. It can be concluded that SST-14 exerted long-term inhibitory effects on the pituitary-ovarian axis and GH cells in rats.

**Key words:** Gonadotropic cells — Somatotropic cells — Ovary — Folliculogenesis — Stereology

**Abbreviations:** FSH, follicle-stimulating hormone; GH, growth hormone; LH, luteinizing hormone; SST, somatostatin; sstr, somatostatin receptor; CL, *corpus luteum*.

### Introduction

Functionality of the hypothalamo-pituitary growth hormone (HPGH) and hypothalamo-pituitary-gonadal (HPG) axes is closely related and age-dependant (Hull and Harvey 2001, 2002). Both axes mature at puberty, and, working in tandem, growth hormone (GH) and gonadotropins account for the somatic and physiological changes that are collectively recognized as puberty. The involvement of GH in activation of the hypothalamo-pituitary-ovarian axis is also of great importance (Hull and Harvey 2001, 2002). In early life, i.e. the infantile period, GH pulsatility is not fully established (Gabriel et al. 1992). On the other hand, blood concentrations of follicle-stimulating hormone (FSH) and luteinizing hormone (LH) reach maxima

around day 12 of age (Ojeda and Ramirez 1972; Dohler and Wuttke 1975; Taya and Sasamoto 1988). High concentrations of circulating gonadotropic hormones during this period are partly the result of very marked FSH and LH responsiveness to the positive influence of gonadotropin-releasing hormone (Ojeda et al. 1977; Taya and Sasamoto 1988), and low responsiveness to the negative influence of endogenous estrogen due to high levels of alpha-fetoprotein (Meijs-Roelofs and Kramer 1979). At the same time, the ovaries are under strong gonadotropic influence and almost twice as many follicles begin to move into more advanced developmental stages during the second postnatal week than at later ages (Ojeda and Urbanski 1994). Therefore, factors that lower GH secretion, such as treatment with somatostatin (SST) analogs, can affect the HPGH and HPG axes. Indeed it has been shown that SST slows down the growth and tempo of reproductive maturation in female rhesus monkeys (Wilson and Tanner 1994).

SST is widely distributed throughout the central nervous system and peripheral tissues in mammals and has

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broad, mostly inhibitory effects on endocrine and exocrine secretions, as well as anti-proliferative action (Patel 1999). SST produced in the periventricular hypothalamic nucleus is a major regulator of GH secretion and other pituitary hormones.

Regulation of the HPG axis by SST is still not well examined. In healthy volunteers, SST inhibited LH pulse amplitude but not frequency, without affecting FSH pulsatility (Samuels et al. 1992). It reduced the serum concentration of LH, but did not affect the release of FSH in women with polycystic ovary syndrome (Prelević et al. 1990) or in male rats (Starčević et al. 2002; Milošević et al. 2004). In adult female rats, SST-14 had an inhibitory influence on both types of gonadotropic cells (Nestorović et al. 2001, 2004a).

Earlier studies have shown the presence of endogenous SST in the ovaries of various species (Mori et al. 1984; McIntyre et al. 1992; Masini et al. 1999; Zhang et al. 2009; Gougeon et al. 2010). Moreover, expression of SST receptors (sstr) was demonstrated in human granulosa (Strauss et al. 2003; Adams et al. 2004) and granulosa lutein cells (Strauss et al. 2003), mouse granulosa cells and oocytes (Gougeon et al. 2010) and rat granulosa cells (Nakamura et al. 2013). An inhibitory effect of SST on ovarian steroidogenesis in granulosa and granulosa lutein cells has also been observed (Rajkumar et al. 1992; Andreani et al. 1995; Holst et al. 1995, 1997; Nakamura et al. 2013).

We have previously shown that SST-14 inhibits gonadotropic cells and the initial phase of folliculogenesis in the infantile (Nestorović et al. 2011) and peripubertal (Nestorović et al. 2008) periods of life. However, whether these effects are long-lasting or not is less well evaluated. Therefore, the aim of this study was to examine possible long-term effects of SST-14 applied at critical periods of life, on some morphological and functional features of pituitary gonadotropic cells, the immunohistochemical and morphometric characteristics of somatotropic cells and on ovarian follicular development in rats.

## Materials and Methods

Time-mated pregnant Wistar rats were housed individually and maintained in a controlled environment (12 h light/12 h dark;  $22 \pm 2^\circ\text{C}$ ), with food (Veterinarski Zavod "Subotica", Subotica, Serbia) and water *ad libitum*.

Two groups of infant females were injected subcutaneously (s.c.) twice daily (8 a.m. and 8 p.m.) with 20  $\mu\text{g}$  SST-14 (S9129, Sigma, St. Louis, Mo., USA) *per* 100 g body weight (b.w.) for 5 consecutive days between 11 and 15 days of age. The first group was sacrificed in the peripubertal period at 38 days old, and the second in adulthood at 80 days old. A third group of female rats was treated with SST-14 in the peripubertal period between 33 and 37 days old by the

same schedule and later killed at 80 days of age. Each treated group had a corresponding control group, which received the equivalent volume of saline s.c. by the same schedule. The dose regimen was based on that of Rebuffat et al. (1984) except that SST-14 was administered every 12 h instead of every 8 h (Nestorović et al. 2001). Blood was collected from individual animals and the sera were stored at  $-70^\circ\text{C}$  until FSH and LH determination. The experimental protocols were approved by the Local Animal Care Committee in conformity with the recommendations provided in the European Convention for Protection of Vertebrate Animals used for Experimental and Other Scientific Purposes (ETS no. 123. Appendix A).

After removal, the pituitaries and ovaries were weighed, fixed in Bouin's solution, dehydrated and embedded in paraffin wax. Pituitary sections (5  $\mu\text{m}$  thick) were stained immunohistochemically, while serial ovary sections (6  $\mu\text{m}$  thick) were stained with hematoxylin and eosin (H&E).

## Pituitary

### Immunohistochemistry

Series of seven sections cut through three tissue levels (dorsal, middle and ventral portions) of the *pars distalis* were used for immunostaining. After rehydration, the sections were stained immunohistochemically. Gonadotropic cells were visualized using the peroxidase-antiperoxidase (PAP) method, as previously described (Nestorović et al. 2001). Anti-rat  $\beta\text{FSH}$  (1:300 v/v) or anti-rat  $\beta\text{LH}$  polyclonal antibodies (1:500 v/v) (National Institute of Health, Bethesda, MD, USA) served as the primary antibodies. For detection of somatotropic cells, sections were incubated with polyclonal anti-human GH antibodies (1:300 v/v (DAKO A/S, Glostrup, Denmark)) (Nestorović et al. 2008).

### Stereological measurements

Immunohistochemically stained 5  $\mu\text{m}$  thick sections of the *pars distalis* were used for morphometric examination of FSH, LH and GH cells that possessed visible nuclei. Cell volumes ( $V_c$ ) and volume densities ( $V_V$ ) were estimated by light microscopy at 1000 $\times$  magnification using the M<sub>42</sub> multipurpose test system (Weibel 1979). Cell volumes were expressed in  $\mu\text{m}^3$ , while volume densities were given as percentages of total pituitary cells *per*  $\text{mm}^3$ . At the same time, the number of immunoreactive cells *per* unit area ( $\text{mm}^2$ ) in each section was analyzed.

## Ovary

Every second ovarian section (6  $\mu\text{m}$  thick) was examined by light microscopy. Follicle and *corpora lutea* (CL) num-

bers were determined by simple point counting. Healthy follicles were divided into two groups: small follicles measuring  $< 275 \mu\text{m}$  in the largest cross-section (LCS) and large follicles measuring  $\geq 275 \mu\text{m}$  LCS. Small follicles were further divided into six classes as defined by Gaytan et al. (1998) as follows: (1) primordial follicles containing an oocyte surrounded by a layer of 3–6 flattened pre-granulosa cells; (2) primary follicles consisting of an oocyte surrounded by a layer containing one enlarged cell or a whole layer of cuboidal pre-granulosa cells; (3) multilaminar class A follicles (Ma) with 1–2 layers of granulosa cells measuring up to  $75 \mu\text{m}$  in diameter; (4) multilaminar class B follicles (Mb) measuring from 76 to  $150 \mu\text{m}$ ; (5) multilaminar class C follicles (Mc) measuring from 151 to  $200 \mu\text{m}$ ; and (6) multilaminar class D follicles (Md) measuring from 201 to  $274 \mu\text{m}$ . Large follicles were divided into five classes as previously defined by Osman (1985): class I, from 276 to  $350 \mu\text{m}$  in diameter; class II, from 351 to  $400 \mu\text{m}$ ; class III, from 401 to  $450 \mu\text{m}$ ; class IV, from 451 to  $575 \mu\text{m}$ ; and class V, more than  $575 \mu\text{m}$  in diameter. Follicle diameters were measured (two diameters at right angles) in the LCS containing the oocyte nucleolus. Atretic follicles exhibited alterations in the granulosa layer and/or on the oocyte. Follicles that contained pycnotic granulosa cells in abundance and/or a degenerated oocyte were classified as atretic. Oocyte degeneration was apparent from the highly irregular shape and/or the presence of a pycnotic nucleus. The number of CL at different phases of development (newly formed, mature and regressive) was determined *per ovary*.

#### Determination of serum hormone concentrations

Serum concentrations of FSH and LH were measured with commercial radioimmunoassay (RIA) kits (Amersham Biosciences UK Ltd, Little Chalfont, Buckinghamshire, UK). Intra-assay and inter-assay coefficients of variation were 4.2 and 7.9% for FSH and 6.5 and 10.9% for LH, respectively.

#### Statistical analysis

All results were expressed as means for five animals *per group*  $\pm$  standard deviation (SD). The data were tested for normality of distribution by the Kolmogorov-Smirnov test. One-way analysis of variance followed by the Newman-Keuls test was used for comparisons of differences between the groups. A probability value of 5% or less was considered as statistically significant.

#### Results

##### Body and organ weights

SST 14 applied to infant rats did not lead to a decreased body weight in the peripubertal period. However, a small but significant reduction of this parameter was noted in adulthood. Body weight of adult females treated with SST-14 during peripuberty was significantly (17.1%) lower than that of the corresponding controls (Table 1).

Absolute pituitary and ovarian weights in peripubertal females were decreased by 40.5% and 35%, respectively, after SST-14 application as infants, but no reduction was noted in adulthood. Peripubertal SST-14 treatment led to a 19.7% lower pituitary weight in adult females ( $p < 0.05$ ), while ovarian weight was not affected, when compared to the corresponding control values.

Relative pituitary and ovarian weights were 38.5% and 30.7% lower, respectively, in peripubertal females treated with SST-14 as infants.

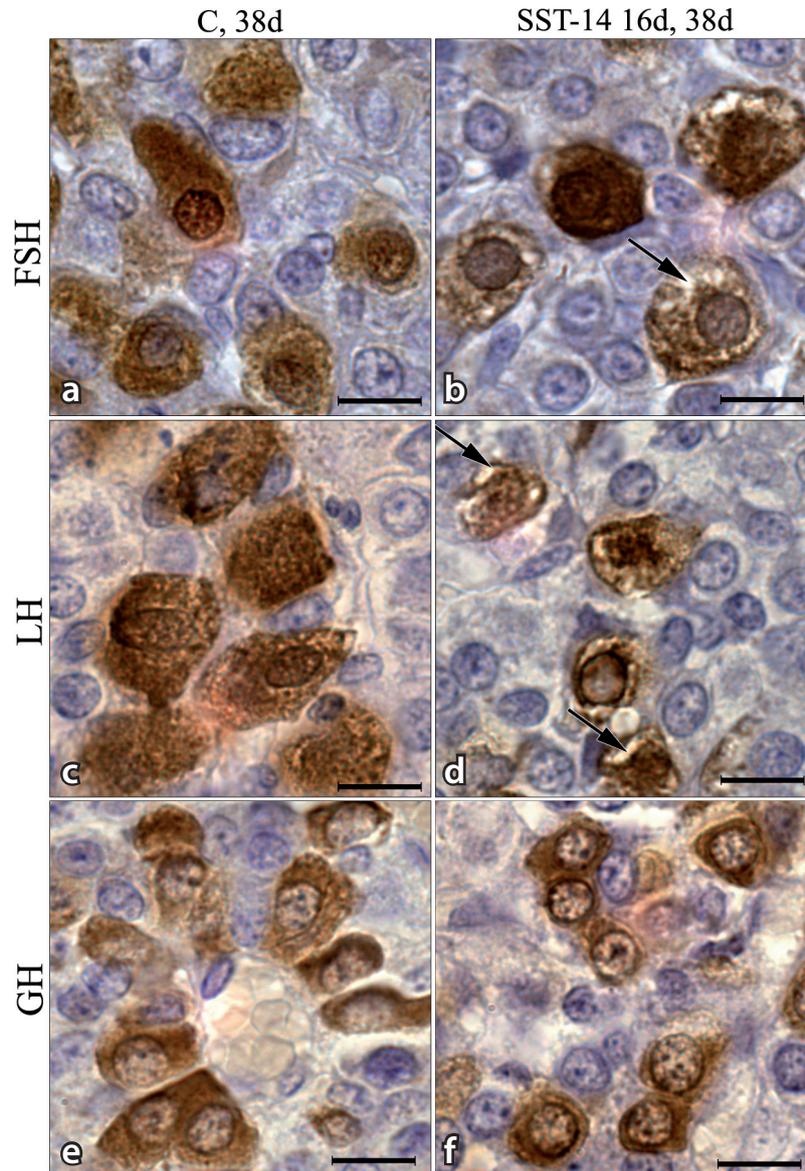
##### Pituitary

FSH (follicle-stimulating hormone) and LH (luteinizing hormone) cells in the pituitary of control peripubertal and adult females were numerous, positioned throughout the *pars distalis*, present in groups or occasionally alone, and often in close contact with blood capillaries. They were strongly

**Table 1.** Body, pituitary and ovary weights of peripubertal (C, 38d) and adult (C, 80d) control females, peripubertal (SST-14 16d, 38d) and adult females treated with SST-14 in infantile (SST-14 16d, 80d) and peripubertal (SST-14 38d, 80d) period of life

Group	Body weight (g)	Absolute pituitary weight (mg)	Relative pituitary weight	Absolute ovarian weight (mg)	Relative ovarian weight
C, 38d	117.44 $\pm$ 7.71	5.60 $\pm$ 0.80	4.78 $\pm$ 0.74	26.17 $\pm$ 5.53	21.99 $\pm$ 2.74
SST-14 16d, 38d	113.33 $\pm$ 8.17	3.33 $\pm$ 0.82*	2.94 $\pm$ 0.67*	17.00 $\pm$ 2.45*	15.23 $\pm$ 2.69*
C, 80d	253.33 $\pm$ 8.16 <sup>#</sup>	11.83 $\pm$ 1.47 <sup>#</sup>	4.67 $\pm$ 0.56	38.00 $\pm$ 1.41 <sup>#</sup>	16.26 $\pm$ 0.43
SST-14 16d, 80d	235.00 $\pm$ 13.78*	11.58 $\pm$ 1.28	4.93 $\pm$ 0.45	38.08 $\pm$ 1.28	16.57 $\pm$ 0.83
SST-14 38d, 80d	210.00 $\pm$ 8.94*	9.5 $\pm$ 1.22*	4.54 $\pm$ 0.68	38.17 $\pm$ 1.94	17.07 $\pm$ 1.06

All values are given as mean  $\pm$  SD ( $n = 5$ ). <sup>#</sup>  $p < 0.05$  adult control *vs.* peripubertal control females; \*  $p < 0.05$  SST-14-treatment *vs.* corresponding control females.

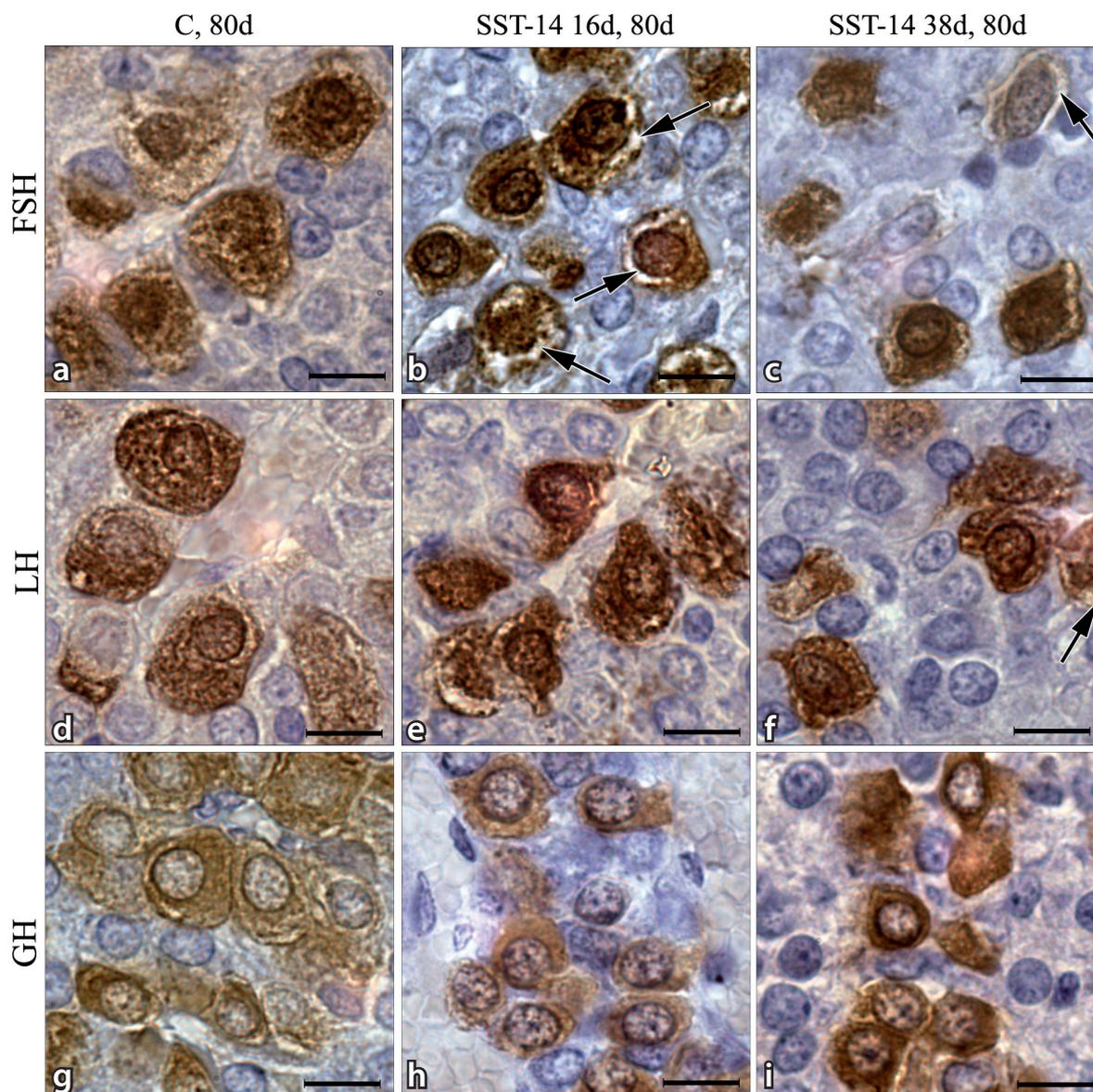


**Figure 1.** Representative micrographs of gonadotropes (a–d) and somatotropes (e, f) from peripubertal control (a, c, e) and peripubertal female rats treated with SST-14 as infants (b, d, f). In the pituitaries of SST-14-treated females, FSH (b) and LH (d) cells were smaller, changed in shape, with a non-uniformly stained cytoplasm (arrows). GH cells (f) were also smaller and altered in shape, bar 10  $\mu\text{m}$ . FSH, follicle-stimulating hormone; GH, growth hormone; LH, luteinizing hormone.

immunohistochemically stained, large, polygonal, oval or polyhedral in shape with prominent, often eccentrically located nuclei (Figs. 1a, 1c, 2a, 2d). In the pituitaries of all treated females FSH (Figs. 1b, 2b, 2c) and LH (Figs. 1d, 2e, 2f) cells appeared smaller, when compared to the controls, with altered shapes, i.e. they were elongated and more polygonal. Their cytoplasm did not stain uniformly. Immunolabeled granules were clustered in groups and separated by unstained regions. The intensity of cytoplasmic immunostaining varied from weak to very intense (Figs. 1b, 2c).

Morphometric analysis showed that gonadotrop numbers *per unit area* ( $\text{mm}^2$ ) were significantly decreased in the pituitaries of adult females when compared to corresponding values for peripubertal controls. This resulted in a significantly lower volume density of LH cells, while that of FSH cells was somewhat lower, but the difference was not statistically significant (Table 2).

Treatment of infant female rats with SST-14 led to decreased FSH and LH cell volume during peripuberty, by 20.9% and 23%, respectively, in comparison to corre-



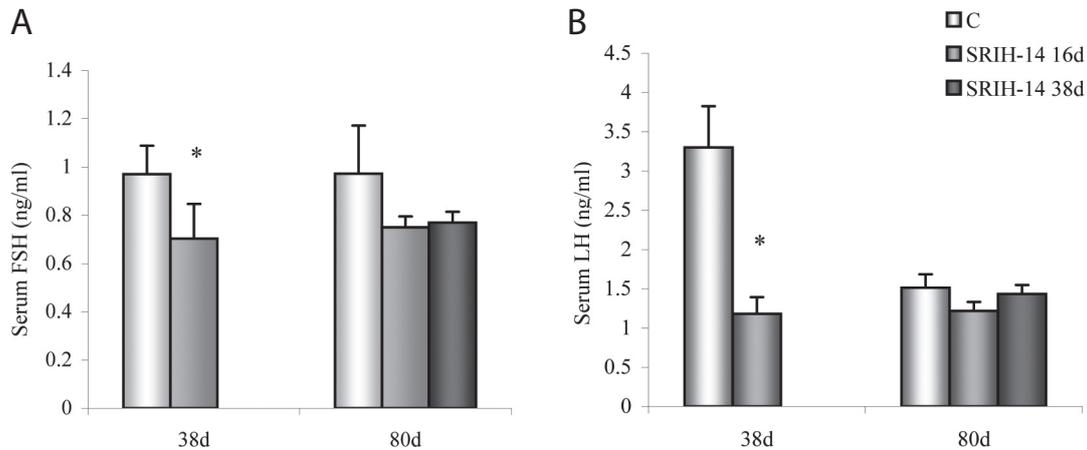
**Figure 2.** Representative micrographs of gonadotrops (a–f) and somatotrops (g–i) from adult control (a, d, g) and adult female rats treated with SST-14 in the infantile (b, e, h) and peripubertal periods of life (c, f, i). In the pituitaries of SST-14-treated females, FSH (b, c) and LH (e, f) cells had changed shapes, with non-uniformly stained cytoplasm (arrows). GH cells (f) were significantly smaller than in control rats, bar 10  $\mu\text{m}$ . FSH, follicle-stimulating hormone; GH, growth hormone; LH, luteinizing hormone.

spending controls (Table 2). When examined as adults, the volume of both types of gonadotropic cells did not differ from the values for adult control rats. The volume density of FSH and LH cells and their number *per unit area* did not change between the peripubertal and adult period of life (Table 2).

When applied during peripuberty, SST-14 decreased the volume and volume density of FSH cells by 21.9% and 20.8%, respectively, in comparison to aged-matched adult controls (Table 2). Similarly, the volume and volume density of LH

cells were 26.7% and 23.1% lower, respectively. Numbers of both types of gonadotrops *per unit area* were not changed by SST-14 treatment, regardless of the age when it was applied (Table 2).

GH cells were abundant in the pituitaries of control peripubertal and adult females. They were positioned throughout the *pars distalis*, mostly in groups in close contact with blood capillaries. Their cytoplasm was immunohistochemically stained strongly and homogeneously and their shape ranged from ovoid to pyramidal (Figs. 1e, 2g). In



**Figure 3.** Serum concentrations of FSH (A) and LH (B) in peripubertal (38d) and adult (80d) control (C) females, and those treated in infantile (SST-14 16d) and peripubertal (SST-14 38d) periods of life. All values are given as means  $\pm$  SD ( $n = 5$ ); \*  $p < 0.05$  SST-14-treatment vs. corresponding control.

the pituitaries of all SST-14 treated females, GH cells were smaller in size than in the corresponding controls (Figs. 1f, 2h, 2i). When GH cells in the pituitaries of peripubertal and adult control rats were compared, it was evident that they were larger in adults (Figs. 1e, 2g). Namely, stereological analysis showed a significant rise of 41.6% in GH cell volume between peripuberty and adulthood.

Infantile SST-14 treatment diminished GH cell volume at peripuberty by 33.7% and in adulthood by 27.4%, when compared to the corresponding controls. The volume density of GH cells was also lower after infantile SST-14 treatment, but the decrease (by 27.6%) was statistically significant only when cells were examined in the adult period of life. SST-14 applied during peripuberty decreased GH cell volume and

**Table 2.** Cell volume ( $V_c$ ;  $\mu\text{m}^3$ ), volume density ( $V_V$ ; %) and number of FSH-, LH- and GH-immunolabelled cells per unit area ( $\text{mm}^2$ ) in the pituitaries of control peripubertal (C, 38d) and adult females (C, 80d) and females treated with SST-14. Pituitary cells of females treated with SST-14 in infantile period of life were analyzed in peripubertal (SST-14 16d, 38d) and adult (SST-14 16d, 80d) period of life. Pituitary cells of females treated with SST-14 in peripubertal period of life were analyzed in adulthood (SST-14 38d, 80d)

Cells	Groups	$V_c$ ( $\mu\text{m}^3$ )	$V_V$ (%)	No/ $\text{mm}^2$
FSH	C, 38d	1305.15 $\pm$ 120.71	13.12 $\pm$ 0.86	572 $\pm$ 46
	SST-14 16d, 38d	1031.80 $\pm$ 105.19*	11.17 $\pm$ 1.50	602 $\pm$ 40
	C, 80d	1171.47 $\pm$ 129.63	10.77 $\pm$ 0.79	251 $\pm$ 5 <sup>#</sup>
	SST-14 16d, 80d	951.46 $\pm$ 72.46	8.95 $\pm$ 1.07	266 $\pm$ 18
LH	C, 38d	1319.54 $\pm$ 67.42	14.01 $\pm$ 1.63	593 $\pm$ 23
	SST-14 16d, 38d	1016.30 $\pm$ 47.75*	11.59 $\pm$ 0.76	583 $\pm$ 71
	C, 80d	1297.38 $\pm$ 197.54	10.12 $\pm$ 1.58 <sup>#</sup>	300 $\pm$ 10 <sup>#</sup>
	SST-14 16d, 80d	1072.28 $\pm$ 169.06	10.38 $\pm$ 0.92	283 $\pm$ 61
GH	C, 38d	649.43 $\pm$ 68.03	30.40 $\pm$ 2.13	2531 $\pm$ 268
	SST-14 16d, 38d	430.73 $\pm$ 17.45*	26.60 $\pm$ 4.18	3000 $\pm$ 464
	C, 80d	919.59 $\pm$ 137.46 <sup>#</sup>	32.62 $\pm$ 0.969	2084 $\pm$ 298
	SST-14 16d, 80d	668.07 $\pm$ 15.23*	23.61 $\pm$ 0.94*	1925 $\pm$ 89
	SST-14 38d, 80d	674.84 $\pm$ 59.68*	26.56 $\pm$ 1.79*	2181 $\pm$ 163

All values are given as mean  $\pm$  SD ( $n = 5$ ); <sup>#</sup>  $p < 0.05$  adult control vs. peripubertal control; \*  $p < 0.05$  SST-14-treatment vs. corresponding control.

**Table 3.** The summarized number of healthy and atretic small growing and antral follicles and the healthy to atretic follicle number ratio, in the ovaries of control peripubertal (C, 38d) and adult (C, 80d) and SST-14 treated rats. Ovaries of females treated with SST-14 in infantile period of life were analyzed in peripubertal (SST-14 16d, 38d) and adult (SST-14 16d, 80d) period of life. Ovaries of females treated with SST-14 in peripubertal period of life were analyzed in adulthood (SST-14 38d, 80d)

Groups	Number of follicles		Ratio	Number of follicles		Ratio
	Healthy small growing follicles ( $\Sigma$ )	Atretic small follicles ( $\Sigma$ )		Healthy antral follicles ( $\Sigma$ )	Atretic antral follicles ( $\Sigma$ )	
C, 38d	647 $\pm$ 91	29 $\pm$ 3	22 : 1	30 $\pm$ 2	65 $\pm$ 10	1 : 2
SST-14 16d, 38d	350 $\pm$ 65*	39 $\pm$ 9	9 : 1	17 $\pm$ 5*	44 $\pm$ 9	1 : 3
C, 80d	335 $\pm$ 53 <sup>#</sup>	16 $\pm$ 5 <sup>#</sup>	21 : 1	24 $\pm$ 4	33 $\pm$ 8 <sup>#</sup>	1 : 1
SST-14 16d, 80d	271 $\pm$ 30	22 $\pm$ 4	12 : 1	16 $\pm$ 4	34 $\pm$ 4	1 : 2
SST-14 38d, 80d	389 $\pm$ 52	20 $\pm$ 4	19 : 1	26 $\pm$ 8	38 $\pm$ 15	1 : 1

All values are given as mean  $\pm$  SD ( $n = 5$ ); <sup>#</sup>  $p < 0.05$  adult control vs. peripubertal control; \*  $p < 0.05$  SST-14-treatment vs. corresponding control.

volume density, by 26.6% and 18.6%, respectively, but the number of GH cells *per* unit area of pituitary was not changed after SST-14 treatment regardless of the time of application or analysis (Table 2).

#### Serum FSH and LH concentrations

Significant declines of serum FSH and LH concentrations were observed in peripubertal rats after infantile SST-14 treatment. Mean FSH concentration was 27.5% lower and mean LH 64.2% lower than the corresponding values for control females (Fig. 3). In adult females no differences in gonadotropic hormone concentrations were observed.

#### Ovary

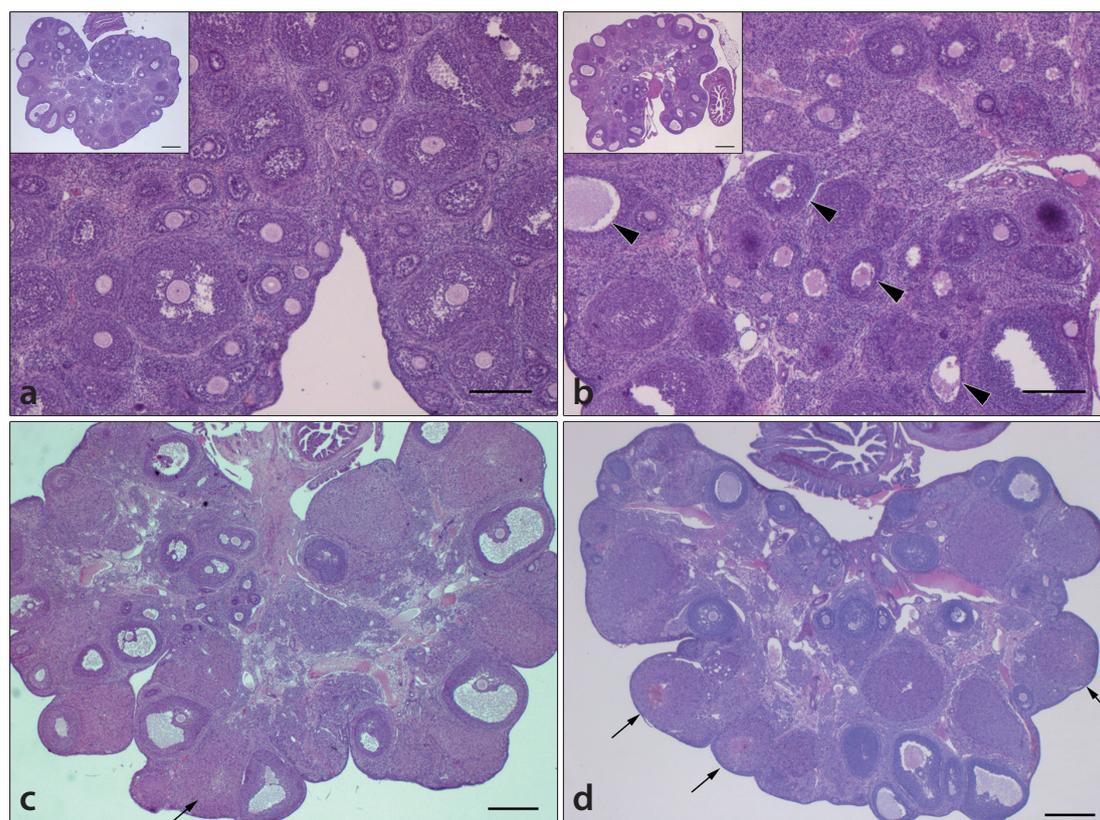
Numerous healthy follicles at all stages of folliculogenesis, as well as rare atretic follicles at different stages of degeneration, were present in the ovaries of peripubertal and adult controls (Fig. 4 a,c). Since the peripubertal control and treated rats had not reached puberty, CL were not present in their ovaries (Fig. 4 a,b). In the ovaries of adult females, healthy newly formed and mature follicles and regressive CL were present (Fig. 4 c,d). Quantification of follicles at different stages of development showed that the number of primordial and healthy multilayered (secondary, preantral) follicles in the ovaries of control adult females was lower than in peripubertal control rats (Fig. 5a). Additionally, atretic small and antral follicle numbers were decreased by 43.4% and 48.7%, respectively (Table 3).

The ovaries of peripubertal females treated with SST-14 as infants were well organized and follicles at all stages of development and degeneration were present. However, light microscopic examination of the ovaries revealed many fewer healthy small follicles than in controls and prevalence of atretic follicles (Fig. 4b). Indeed, the morphometric analysis

showed that the number of healthy small, multilayered follicles was diminished. Significant decreases were evident in several individual classes: Ma by 48.0%, Mb by 62.0%, Mc by 58.9% and Md by 51.3% (Fig. 5a). The sum of healthy small follicles was 45.9% lower than for the corresponding control (Table 3). This resulted in declines within individual classes of healthy large, antral, follicles (Fig. 5b) and a significant 43.8% decrease in the sum of antral follicle numbers (Table 3). Fewer healthy follicles pushed the healthy to atretic follicle ratio in favor of atretic for both small and large follicles (Table 3).

Follicles and CL at all stages of development were seen in the ovaries of control adult females and those treated with SST-14 as infants (not shown). However, many fewer healthy antral follicles were observed than in control ovaries. Morphometric analysis showed fewer healthy antral follicles, but the difference was significant only for class I follicles (Fig. 5b). Although the number of healthy or atretic follicles was not significantly changed, the healthy to atretic follicle ratio favored atretic for both small and large follicles (Table 3). The number of newly formed CL was similar to that for the corresponding controls, while the number of mature CL was 31.5% lower and regressive CL 52.9% higher (Fig. 6).

Numerous follicles at different stages of development and atresia, as well as CL with slight prevalence of CL in regression were present on H&E-stained sections of ovaries from adult females that had received SST-14 during peripuberty (Fig. 4d). Detailed histological and morphometrical analysis showed that the number of primordial i.e. non-growing, quiescent follicles, was significantly higher (by 81.0%) when compared to the corresponding control value. Healthy or atretic small and large follicle numbers were not changed (Figs. 5a, b; Table 3). However, the numbers of young and mature CL were decreased by 50.2% and 32.2%, respectively, while that of regressive CL was increased by 97.6%, when compared to corresponding control values (Fig. 6).



**Figure 4.** Representative micrographs of ovaries of a peripubertal control female (a); a peripubertal female treated with SST-14 as an infant (b); an adult control female (c) and an adult female treated with SST-14 during peripuberty (d). Ovaries of control females are characterized by the presence of numerous healthy follicles in various stages of development, while in the ovaries of SST-14 treated females numerous degenerated follicles in various stages of atresia (arrowheads) and regressive CL (arrows) are present (b, d). a, b, bar 40  $\mu\text{m}$ ; c, d, bar 400  $\mu\text{m}$ . Respective whole-ovarian section is shown in the inset, bar 400  $\mu\text{m}$ .

## Discussion

Short-term inhibitory effects of SST-14 on rat pituitaries and ovaries in critical periods of life have been documented (Nestorović et al. 2008, 2011). To elucidate whether SST-14 has long-term inhibitory effects on the pituitary-ovarian axis and somatotrophic cells, female rats were treated with SST-14 in the critical infantile or peripubertal period, and the effects were examined at later ages. Our results demonstrate that inhibition by SST-14, although less severe than immediately after application, can be detected in the pituitary and ovaries in later periods of life.

The biological effects of SST are mediated through five distinct receptor subtypes ( $\text{sstr}_{1-5}$ ). All five receptor mRNAs are expressed in the rat pituitary. This is developmentally determined in a subtype-specific manner, with the expression of  $\text{sstr}_2$  mRNA and protein rising from birth and reaching a maximum in the adult (Reed et al. 1999). The abundance of other  $\text{sstr}$  mRNAs does not change with age (Reed et al. 1999). Therefore, in our experimental conditions, SST-14 could

directly inhibit pituitary cells, since the  $\text{sstrs}$  are expressed at the ages when SST-14 was applied, and this hormone has the same affinity for all types of  $\text{sstrs}$  (Patel 1999).

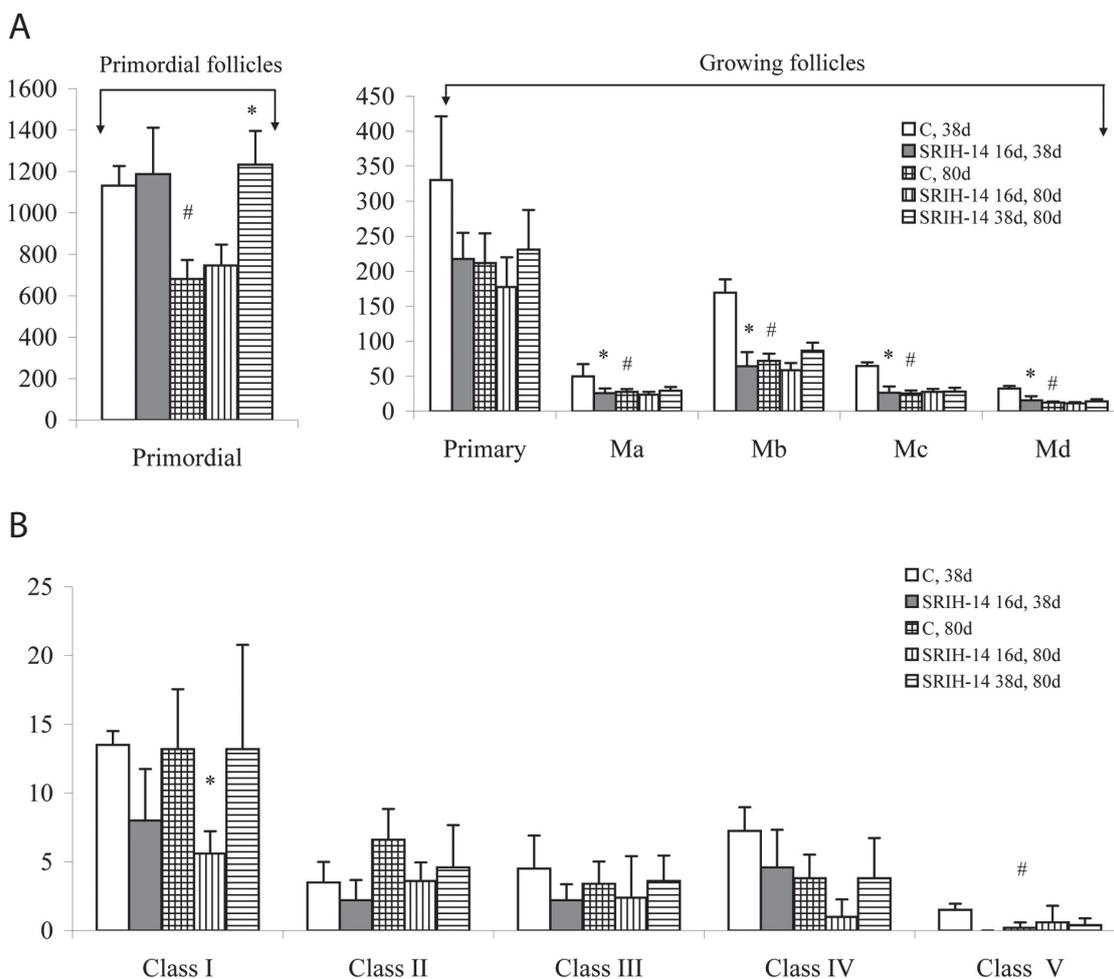
However, since SST-14 was applied systemically and not centrally, an inhibitory effect on gonadotropic and GH cells, mediated through inhibition of specific hypothalamic-releasing factors cannot be excluded. The somatostatin analog, octreotide, was shown to inhibit activation of gonadotropin-releasing hormone (GnRH)-containing cells in the hypothalamus of ovariectomized adult rats (Van Vugt et al. 2004) and expression of  $\text{sstr}_{2-4}$  mRNA was detected in mouse GnRH neurons (Todman et al. 2005). SST inhibits secretion of growth hormone-releasing hormone (GHRH) (Giustina and Veldhuis 1998) and expression of  $\text{sstrs}$  was demonstrated in the rat arcuate nucleus even during the neonatal period of life (Thoss et al. 1995).

The dominant effect of SST on pituitary function is inhibition of hormone secretion, mostly as a result of acute inhibition of hormone exocytosis (Ben Shlomo and Melmed, 2010). However, in our study we observed a long-term

effect of SST-14 on gonadotropic and somatotropic cells after multiple treatments of infant and peripubertal rats. By adulthood, gonadotropic cells showed signs of recovery in terms of blood concentrations of both FSH and LH but gonadotropic cell morphology still showed signs of inhibition. Thus, morphometric parameters did not reach adult control values after peripubertal SST-14 application. Higher sensitivity of gonadotropic cells in the peripubertal period could be expected, as *sstr*<sub>2</sub> expression in the pituitary rises with age (Reed et al. 1999) and the dominant types of *sstr*s in LH cells are *sstr*<sub>2</sub> and *sstr*<sub>1</sub>. Low but similar amounts of all *sstr*s are expressed in FSH cells (O'Carroll and Krempels 1995).

SST-14 has a long-term inhibitory effect on GH cells, as judged by their morphology and morphometric parameters. Thus, GH cell volume did not recover till adulthood after SST-14 application to both infant and peripubertal rats. In

the pituitaries of control animals GH cell volume rose by 41.6% between peripuberty and adulthood. This increase in cell volume was not achieved by GH cells exposed to SST-14 in the infantile period of life. These results confirm those from an earlier study where neonatally applied SST-14 markedly decreased all morphometric parameters of GH cells and these changes were sustained until adulthood (Nestorović et al. 2006). Therefore, besides the inhibitory effect of SST on pituitary hormone secretion, from our results it can be concluded that SST has an antigrowth influence on gonadotropic and somatotropic cells. Ben-Shlomo and Melmed (2010) suggested that this effect is probably mediated through induction of cell apoptosis or induction of cell senescence, rather than inhibition of mitogenesis. Indeed, pycnotic gonadotropic cells were observed after acute central octreotide administration (Nestorović et al. 2004b).



**Figure 5.** Average number of healthy small follicles (A) and healthy large follicles (B) per ovary of control peripubertal (C, 38d) and adult (C, 80d) animals, peripubertal females treated with SST-14 as infants (SST-14 16d, 38d) and adult females treated with SST-14 in the infantile (SST-14 16d, 80d) and peripubertal (SST-14 38d, 80d) periods of life. All values are given as means  $\pm$  SD ( $n = 5$ ); #  $p < 0.05$  adult control vs. peripubertal control; \*  $p < 0.05$  SST-14-treatment vs. corresponding control. For more details see Materials and Methods.

The decreased absolute pituitary weight of peripubertal and adult females that had received SST-14 as infants and during peripuberty, respectively, could be the result of this antigrowth effect of SST. However, relative pituitary weight was decreased only in peripubertal females treated as infants, since SST-14 did not affect body weight. On the other hand, peripubertal application of SST-14 led to reduction of both pituitary and body weight of adult females.

In the ovary, increase of primordial follicle number was noted in adult females that had received multiple SST-14 treatment during peripuberty, while in peripubertal and adult females given SST as infants, normalization of primordial to primary follicle transition occurred. This suggests that the period of life when SST-14 is administered determines the effect of this hormone on the non-growing pool of follicles. This is in accordance with our previous findings of enlargement of the non-growing pool of follicles after application of SST-14 chronically to immature, or in multiple doses to infantile and peripubertal females (Nestorović et al. 2004a, 2008, 2011), but not if it was applied in multiple doses to adult rats (Nestorović et al. 2001).

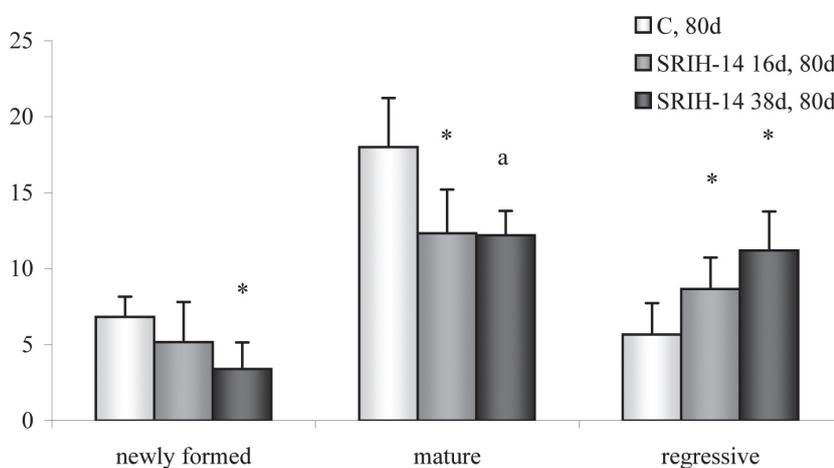
The mechanisms that regulate the initial process of folliculogenesis still remain unclear, but it is certain that the pool of primordial follicles is under inhibitory control, since removal of ovarian tissue and its fragmentation causes the primordial follicles to move *en masse* towards the stage of late primary follicles (Wandji et al. 1997). Somatostatin probably enhanced the inhibitory influence on initial follicle recruitment, since *in vitro* treatment of mouse ovaries with an antagonist caused an opposite effect (Gougeon et al. 2010).

Although treatment with SST-14 in the infantile period of life did not have long-term consequences on primordial to primary follicle transition, an impaired process of folliculogenesis can be seen at later stages, as the numbers of preantral and small antral follicles were reduced in both

peripubertal and adult periods. This resulted in a decrease of absolute and relative ovary weight in peripubertal animals. In the ovaries of peripubertal and adult females that received SST-14 as infants, the healthy to atretic follicle ratio changed in favor of atretic follicles in both the small and large follicle group. This implies that SST-14 enhanced atretic processes, which contrasts with the short-term effect of SST-14 applied in the infantile or peripubertal period of life (Nestorović et al. 2008, 2011). In adult females that received SST-14 during peripuberty, enhancement of atretic processes was not noted, which could suggest that the age when SST-14 is administered determines its effect on follicular atresia.

Disturbed folliculogenesis was also seen at the level of CL. The overall number was not significantly changed, but regressive CL had increased at the expense of healthy ones. Thus, the ratio of healthy newly formed and mature CL to regressive ones was disturbed in adult females treated with SST-14 during peripuberty in the same manner as in those treated as infants. An inhibitory effect of SST on ovarian steroidogenesis in granulosa and granulosa lutein cells has been described previously (Rajkumar et al. 1992; Andreani et al. 1995; Holst et al. 1995, 1997). Recently Nakamura et al. (2013) showed that treatment of rat granulosa cells *in vitro* with SST analogs at pharmacologic doses decreased FSH-induced estradiol and progesterone production with reduction of mRNA for key steroidogenic enzymes. Therefore, since survival of CL depends on progesterone (Goyeneche et al. 2003), fewer healthy CL and more CL in regression in the ovaries of SST-14 treated females could be due to decreased progesterone production.

The disturbance of ovarian folliculogenesis seen in this study could have been the consequence of a direct effect of SST on the ovary. The presence of sstrs in granulosa cells of mouse and rat ovaries has been documented (Gougeon et al. 2010; Nakamura et al. 2013). However, SST might affect ovarian folliculogenesis indirectly *via* pituitary gonadotropin and GH modulation, especially at later stages. Both gonadotropin hormones are



**Figure 6.** Average number of healthy newly formed and mature corpora lutea (CL) and regressive CL per ovary of control adult females (C, 80d), adult females treated with SST-14 as infants (SST-14 16d, 38d) and adult females treated with SST-14 during peripuberty (SST-14 16d, 80d). All values are given as means  $\pm$  SD ( $n = 5$ ); \*  $p < 0.05$  SST-14-treatment vs. control.

necessary for maintenance and control of ovarian functions including folliculo- and steroidogenesis (Richards, 1980). Additionally, there is evidence that GH is also required for ovarian follicular growth (Hull and Harvey 2001, 2002; Bachelot et al. 2002). The long-lasting inhibitory effects of SST-14 on pituitary gonadotropic and GH cells shown here could negatively affect ovarian folliculogenesis. Thus, SST-14 was applied to female rats in critical periods for development and maturation of the HPG and HPGH axes. The route of SST-14 application, i.e. peripheral administration, enabled estimation of effects of SST-14 on the pituitary-ovarian axis as a whole.

To conclude, application of SST-14 to infant or peripubertal females exerted long-term inhibitory effects on pituitary gonadotropic and somatotrophic cells. In the ovary SST-treatment disturbed the process of folliculogenesis creating an endocrine/paracrine environment that was less suitable for follicular growth and development.

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## References

- Adams I. P., Staunton B., Gilson J., Adams R. L., Robinson J., Atkin S. L. (2004): The expression of somatostatin receptors in human granulosa cells. *British Endocrine Societies Joint Meeting 2004*, Endocrine Abstracts 7, 185
- Andreani C. L., Lazzarin N., Pierro E., Lanzone A., Mancuso S. (1995): Somatostatin action on rat ovarian steroidogenesis. *Hum. Reprod.* **10**, 1968–1973
- Bachelot A., Monget P., Imbert-Bolloré P., Coshigano K., Kopchick J. J., Kelly P. A., Binart N. (2002): Growth hormone is required for ovarian follicular growth. *Endocrinology* **143**, 4104–4112 <http://dx.doi.org/10.1210/en.2002-220087>
- Ben-Shlomo A., Melmed S. (2010): Pituitary somatostatin receptor signaling. *Trends Endocrinol. Metab.* **21**, 123–133 <http://dx.doi.org/10.1016/j.tem.2009.12.003>
- Dohler K. D., Wuttke W. (1975): Changes with age in levels of serum gonadotropins, prolactin and gonadal steroids in pre-pubertal male and female rats. *Endocrinology* **97**, 898–907 <http://dx.doi.org/10.1210/endo-97-4-898>
- Gabriel S. M., Roncancio J. R., Ruiz N. S. (1992): Growth hormone pulsatility and the endocrine milieu during sexual maturation in male and female rats. *Neuroendocrinology* **56**, 619–628 <http://dx.doi.org/10.1159/000126284>
- Gaytan F., Morales C., Bellido C., Aguilar E., Sanchez-Criado J. E. (1998): Ovarian follicle macrophages: is follicular atresia in the immature rat a macrophage-mediated event? *Biol. Reprod.* **58**, 52–59 <http://dx.doi.org/10.1095/biolreprod58.1.52>
- Giustina A., Veldhuis J. D. (1998): Pathophysiology of the neuroregulation of growth hormone secretion in experimental animals and the human. *Endocr. Rev.* **19**, 717–797
- Gougeon A., Delangle A., Arouche N., Stridsberg M., Gottelandm J. P., Loumaye E. (2010): Kit ligand and the somatostatin receptor antagonist, BIM-23627, stimulate in vitro resting follicle growth in the neonatal mouse ovary. *Endocrinology* **151**, 1299–1309 <http://dx.doi.org/10.1210/en.2009-0762>
- Goyeneche, A. A., Deis, R. P., Gibori, G., Telleria, C. M. (2003): Progesterone promotes survival of the rat corpus luteum in the absence of cognate receptor. *Biol. Reprod.* **68**, 151–158 <http://dx.doi.org/10.1095/biolreprod.102.007898>
- Holst N., Jacobsen M. B., Haug E., Tanbo T., Abyholm T. (1995): Somatostatin in physiological concentrations inhibits basal and enhances luteinizing hormone-stimulated progesterone release from human granulosa-luteal cells. *Hum. Reprod.* **10**, 1363–1366 <http://dx.doi.org/10.1093/HUMREP/10.6.1363>
- Holst N., Kierulf K. H., Seppala M., Koistinen R., Jacobsen M. B. (1997): Regulation of insulin-like growth factor-binding protein-1 and progesterone secretion from human granulosa-luteal cells: effects of octreotide and insulin. *Fertil. Steril.* **68**, 478–482 [http://dx.doi.org/10.1016/S0015-0282\(97\)00248-3](http://dx.doi.org/10.1016/S0015-0282(97)00248-3)
- Hull K. L., Harvey S. (2001): Growth hormone: roles in female reproduction. *J. Endocrinol.* **168**, 1–23 <http://dx.doi.org/10.1677/joe.0.1680001>
- Hull K. L., Harvey S. (2002): GH as a co-gonadotropin: the relevance of correlative changes in GH secretion and reproductive state. *J. Endocrinol.* **172**, 1–19 <http://dx.doi.org/10.1677/joe.0.1720001>
- Masini M. A., Sturla M., Uva B. (1999): Somatostatin in the ovary of an African lungfish (Protopterus annectens): an in situ hybridisation, immunohistochemical, and autoradiographical study. *Gen. Comp. Endocrinol.* **114**, 287–292 <http://dx.doi.org/10.1006/gcen.1999.7257>
- McIntyre H. D., Marechal D. J., Deby G. P., Mathieu A. G. Hezee-Hagelstein M. T., Franchimont P. P. (1992): Immunoreactive somatostatin in the rat ovary. *Acta Endocrinol. (Copenh.)* **126**, 553–558
- Meijs-Roelofs H. M., Kramer P. (1979): Maturation of the inhibitory feedback action of oestrogen on follicle-stimulating hormone secretion in the immature female rat: a role for alpha-foetoprotein. *J. Endocrinol.* **81**, 199–208 <http://dx.doi.org/10.1677/joe.0.0810199>
- Milošević V., Nestorović N., Filipović B., Velković S., Starčević V. (2004): Centrally applied somatostatin influences morphology of pituitary FSH cells but not FSH release. *Gen. Physiol. Biophys.* **23**, 375–380
- Mori T., Saito H., Ohno Y., Irahara M., Hosoi E., Saito S. (1984) Evidence for existence of somatostatin-like immunoreactivity with molecular heterogeneity in porcine ovaries. *Acta Endocrinol. (Copenh.)* **106**, 254–259
- Nakamura E., Otsuka F., Inagaki K., Tsukamoto N., Ogura-Ochi K., Miyoshi T., Toma K., Takeda M., Makino H. (2013): Involvement of bone morphogenetic protein activity in somatostatin actions on ovarian steroidogenesis. *J. Steroid. Biochem. Mol. Biol.* **134**, 67–74 <http://dx.doi.org/10.1016/j.jsbmb.2012.10.018>
- Nestorović N., Lovren M., Sekulić M., Filipović B., Milošević V. (2001): Effects of multiple somatostatin treatment on rat gonadotrophic cells and ovaries. *Histochem. J.* **33**, 695–702 <http://dx.doi.org/10.1023/A:1016366704630>

- Nestorović N., Lovren M., Sekulić M., Negić N., Šošić-Jurjević B., Filipović B., Milošević V. (2004a): Chronic somatostatin treatment affects pituitary gonadotrophs, ovaries and onset of puberty in rats. *Life Sci.* **74**, 1359–1373  
<http://dx.doi.org/10.1016/j.lfs.2003.07.042>
- Nestorović N., Lovren M., Sekulić M., Negić N., Šošić-Jurjević B., Manojlović-Stojanoski M., Filipović B., Milošević V. (2004b): Effects of intracerebroventricularly administered octreotide on gonadotrophic cells in female rats. *Jugoslav. Med. Biochem.* **23**, 149–153  
<http://dx.doi.org/10.2298/JMH0402149N>
- Nestorović N., Terzić M., Negić N., Starčević V., Milošević V. (2006): Neonatally applied SRIF-14 has immediate and prolonged inhibitory effect on pituitary GH cells. *Acta Vet.* **56**, 457–466  
<http://dx.doi.org/10.2298/AVB0606457N>
- Nestorović N., Manojlović-Stojanoski M., Ristić N., Sekulić M., Šošić-Jurjević B., Filipović B., Milošević V. (2008): Somatostatin-14 influences pituitary-ovarian axis in peripubertal rats. *Histochem. Cell. Biol.* **130**, 699–708  
<http://dx.doi.org/10.1007/s00418-008-0442-7>
- Nestorović N., Ristić N., Manojlović-Stojanoski M., Šošić-Jurjević B., Trifunović S., Savin S., Milošević V. (2011): Somatostatin 14 affects the pituitary-ovarian axis in infant rats. *Histol. Histopathol.* **26**, 157–166
- O'Carroll A. M., Krempeles K. (1995): Wide spread distribution of somatostatin receptor messenger ribonucleic acids in rat pituitary. *Endocrinology* **136**, 5224–5227
- Ojeda S. R., Ramirez V. D. (1972): Plasma level of LH and FSH in maturing rats: response to hemigonadectomy. *Endocrinology* **90**, 466–472  
<http://dx.doi.org/10.1210/endo-90-2-466>
- Ojeda S. R., Jameson H. E., McCann S. M. (1977): Developmental changes in pituitary responsiveness to luteinizing hormone-releasing hormone (LHRH) in the female rat: ovarian-adrenal influence during the infantile period. *Endocrinology* **100**, 440–451  
<http://dx.doi.org/10.1210/endo-100-2-440>
- Ojeda S. R., Urbanski H. E. (1994): Puberty in the rat. In: *The Physiology of Reproduction*. (2nd edition, Eds. E. Knobil and J. Neill), pp 363–409, Raven Press, New York
- Osman P. (1985): Rate and course of atresia during follicular development in the adult cyclic rat. *J. Reprod. Fertil.* **73**, 261–270  
<http://dx.doi.org/10.1530/jrf.0.0730261>
- Patel Y. C. (1999): Somatostatin and its receptor family. *Front. Neuroendocrinol.* **20**, 157–198.  
<http://dx.doi.org/10.1006/frne.1999.0183>
- Prelević G. M., Wurzbürger M. I., Balint-Perić L., Nešić J. S. (1990): Inhibitory effect of sandostatin on secretion of luteinising hormone and ovarian steroids in polycystic ovary syndrome. *Lancet* **336**, 900–903  
[http://dx.doi.org/10.1016/0140-6736\(90\)92270-R](http://dx.doi.org/10.1016/0140-6736(90)92270-R)
- Rajkumar K., Kerr D. E., Kirkwood R. N., Laarveld B. (1992): Inhibitory action of somatostatin-14 on hormone-stimulated cyclic adenosine monophosphate induction in porcine granulosa and luteal cells. *J. Endocrinol.* **134**, 297–306  
<http://dx.doi.org/10.1677/joe.0.1340297>
- Rebuffat P., Robba C., Mazzocchi G., Nussdorfer G. G. (1984): Inhibitory effect of somatostatin on the growth and steroidogenic capacity of rat adrenal zona glomerulosa. *J. Steroid. Biochem.* **21**, 387–390  
[http://dx.doi.org/10.1016/0022-4731\(84\)90301-7](http://dx.doi.org/10.1016/0022-4731(84)90301-7)
- Reed D. K., Korytko A. I., Hipkin R. W., Wehrenberg W. B., Schonbrunn A., Cuttler L. (1999): Pituitary somatostatin receptor (sst)1-5 expression during rat development: age-dependent expression of sst2. *Endocrinology* **140**, 4739–4744
- Richards J. S. (1980): Maturation of ovarian follicles: action and interactions of pituitary and ovarian hormones on follicular cell development. *Recent. Prog. Horm. Res.* **32**, 477–527
- Samuels M. H., Henry P., Ridgway E. C. (1992): Effects of dopamine and somatostatin on pulsatile pituitary glycoprotein secretion. *J. Clin. Endocrinol. Metab.* **74**, 217–222
- Starčević V., Milošević V., Brkić B., Severs W. B. (2002): Somatostatin affects morphology and secretion of pituitary luteinizing hormone (LH) cells in male rats. *Life Sci.* **70**, 3019–3027  
[http://dx.doi.org/10.1016/S0024-3205\(02\)01551-5](http://dx.doi.org/10.1016/S0024-3205(02)01551-5)
- Strauss M. C., Seelig A. S., Weiss J. M., Diedrich K., Ortmann O. (2003): Expression of somatostatin and its receptors in human ovary. *Hum. Reprod.* **18**, 495
- Taya K., Sasamoto S. (1988): Age-related changes in the secretory pattern of FSH and LH in response to LH-RH in prepubertal female rats. *Endocrinol. Jpn.* **35**, 349–355  
<http://dx.doi.org/10.1507/endocrj1954.35.349>
- Thoss V. S., Perez J., Duc D., Hoyer D. (1995): Embryonic and postnatal mRNA distribution of five somatostatin receptor subtypes in the rat brain. *Neuropharmacol.* **34**, 1673–1688  
[http://dx.doi.org/10.1016/0028-3908\(95\)00135-2](http://dx.doi.org/10.1016/0028-3908(95)00135-2)
- Todman M. G., Han S. K., Herbison A. E. (2005): Profiling neurotransmitter receptor expression in mouse gonadotropin-releasing hormone neurons using green fluorescent protein promoter transgenics and microarrays. *Neuroscience* **132**, 703–712  
<http://dx.doi.org/10.1016/j.neuroscience.2005.01.035>
- Van Vugt H. H., Swarts H. J., Van de Heijning B. J., Van der Beek E. M. (2004): Centrally applied somatostatin inhibits the estrogen-induced luteinizing hormone surge via hypothalamic gonadotropin-releasing hormone cell activation in female rats. *Biol. Reprod.* **71**, 813–819  
<http://dx.doi.org/10.1095/biolreprod.104.028936>
- Wandji S. A., Srsen V., Nathanielsz P. W., Eppig J. J., Fortune J. E. (1997): Initiation of growth of baboon primordial follicles in vitro. *Hum. Reprod.* **12**, 1993–2001  
<http://dx.doi.org/10.1093/humrep/12.9.1993>
- Weibel E. R. (1979): Practical methods for biological morphometry. In: *Stereological Methods*. Vol. 1, pp. 1–415, Academic Press, London
- Wilson M. E., Tanner J. M. (1994): Somatostatin analog treatment slows growth and the tempo of reproductive maturation in female rhesus monkeys. *J. Clin. Endocrinol. Metab.* **79**, 495–501
- Zhang L., Li W., Hong X., Lin H. (2009): Regulation of preprosomatostatin 1 (PSS1) gene expression by 17 $\beta$ -estradiol and identification of the PSS1 promoter region in orange-spotted grouper (*Epinephelus coioides*). *Mol. Cell. Endocrinol.* **311**, 87–93  
<http://dx.doi.org/10.1016/j.mce.2009.06.008>

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