doi: 10.4149/gpb_2014044

Editorial Commentary

The meth brain: methamphetamines alter brain functions *via* NMDA receptors

Juliane Proft and Norbert Weiss

Institute of Organic Chemistry and Biochemistry, Academy of Sciences of the Czech Republic, v.v.i., Prague, Czech Republic

Commentary to: Functional changes in pyramidal neurons in the chronic methamphetamine-treated rat. (Gen. Physiol. Biophys. 2015, pp. 5–12)

Key words: Ion channel — Methamphetamine — Piriform cortex — NMDA receptor — AMPA receptor

Methamphetamines (MAP) like crystal meth (MDA 3.4 methylendioxyamphetamine) and ecstasy (MDMA, 3.4 methylendioxymethamphetamine) are a group of neurotoxic drugs often used as a recreational drug and potentially to treat some neurological disorders. For instance, MDMA has been used as a therapeutic drug for posttraumatic stress disorder (PTSD) (Parrott 2014) as well as for attention deficit hyperactivity disorder (ADHD), although it has been declared as non-safe treatment due to its neurotoxicity and its addictive effect in human (Rusyniak 2013; Parrott 2014). Furthermore, addictive use of MAP derivatives has been shown to cause impaired learning and memory as well as other mental disorders (Schroder et al. 2003). In addition, an increased risk of Parkinson's disease (Bognar et al. 2013) has been documented in MPA users (Callaghan et al. 2012). Neurotoxicity of MPAs was explained by alteration of NMDA receptors and dopamine signaling pathways (Simoes et al. 2007; Ares-Santos et al. 2013). In addition, ecstasy binds to serotonin transporters and causes depletion of serotonin from its storage as well as release of dopamine and other neurotransmitters (White et al. 1996; Kish et al. 2010). Considerable efforts were made to characterize the influence of MAP derivatives

and investigated *via* electrophysiological recordings the influence of MPA on piriform cortex neurons, especially focusing on NMDA and AMPA receptors activity. The group observed the typical sniffing behavior and increase of movement in chronically-treated rats, the same behavior that is often observed in humans using MPA over a long period of time. These changes in behavior come with alterations of the morphology of dentrites of pyramidal cells. MPA-treated rats showed blebbing of the dentrites visible after staining with Lucifer yellow, to better identify

on hippocampal structures in the brain, but little is known

about the alterations in the sensory system, especially the

piriform cortex, the area that is mostly known to sense

In this issue of General Physiology and Biophysics, Hori et al. (pp. 5–12) treated rats chronically with MPA

odors (White et al. 1996).

breaks up causing an outward bulge of the cell membrane (Vermeulen et al. 2005). Blebbing can also play a role in other cellular processes like necrosis (Wyllie et al. 1980), chemical or physical stress, cell locomotion or division (Norman et al. 2010).

the soma and dentrites of neurons. Blebbing of the cell

typically occurs during apoptosis where the cytoskeleton

In addition, the authors observed a significant alteration of the electrical properties of the pyramidale neurons characterized by decrease of the membrane potential and input resistance of the cells. In order to further investigate the influence of MPA on neuronal network excitability and plasticity, transient post tetanic potentiation (PTP) and long-term potentiation (LTP) were analyzed (Gasparova et al. 2014). While PTP remains unaltered, LTP was significantly decreased in MPA-treated animals. In addition, ionotrophic application of AMPA and NMDA indicates an

Correspondence to: Juliane Proft, Institute of Organic Chemistry and Biochemistry, Academy of Sciences of the Czech Republic, v.v.i., Flemingovo nám. 2, 166 10 Prague 6 – Dejvice, Czech Republic

E-mail: proft@uochb.cas.cz

Norbert Weiss, Institute of Organic Chemistry and Biochemistry, Academy of Sciences of the Czech Republic, v.v.i., Flemingovo nám. 2, 166 10 Prague 6, Dejvice, Czech Republic E-mail: weiss@uochb.cas.cz

altered AMPA/NMDA receptors activity in MPA-treated rats. Considering that NMDA and AMPA receptors represent the molecular substrate of LTP, it is likely that alteration of NMDA/AMPA response contributes to the alteration of LTP induced by MPA treatment.

Glutamatergic NMDA and AMPA receptors represent essential component of synaptic plasticity and long-term potentiation and depression (Luscher and Malenka 2012; Mokrushin and Pavlinova 2013). The observation that chronic treatment with MPA alters NMDA/AMPA response certainly represents an interesting molecular substrate for MPA-dependent alteration of cognitive functions. In addition, it is well accepted that alteration of NMDA and AMPA receptors significantly contribute to neurodegenerative disorders like Parkinson's and Alzheimer's diseases (You et al. 2012; Proft and Weiss 2014). Interestingly, a MPA-induced animal model for Parkinson's disease (Proft et al. 2011; Curtin et al. 2014; Tai et al. 2014) has been described. Moreover, a binding of MPA to α-synuclein has been reported and causes missfolding of αsynuclein, a key protein in Parkinson's disease (Tavassoly and Lee 2012). It is possible that missfolded α -synuclein could alter gluramatergic NMDA-dependent signaling pathway like it has been shown for missfolded amyloid (Proft and Weiss 2012; Stys et al. 2012; You et al. 2012).

Overall the results described in the paper by Hori et al. represent an interesting molecular substrate of how drug abuse might cause neurodegenerative disorders and a better understanding of the interaction of those drugs with key neuronal proteins will certainly highlight not only the molecular mechanism of drug-induced cognitive disorders but also potentially translate to a better basic understanding of those diseases.

Acknowledgment. The work in N.W.'s laboratory was supported by the Czech Science Foundation [Grant 15-13556S] and the Institute of Organic Chemistry and Biochemistry (IOCB). J.P. is supported by a postdoctoral fellowship from IOCB.

References

Ares-Santos S., Granado N., Moratalla R. (2013): The role of dopamine receptors in the neurotoxicity of methamphetamine. J. Intern. Med. **273**, 437–453

http://dx.doi.org/10.1111/joim.12049

Bognar C., Baldovic M., Benetin J., Kadasi L., Zatkova A. (2013): Analysis of Leucine-rich repeat kinase 2 (LRRK2) and Parkinson protein 2 (parkin, PARK2) genes mutations in Slovak Parkinson disease patients. Gen. Physiol. Biophys. 32, 55–66 http://dx.doi.org/10.4149/gpb_2013006

Callaghan R. C., Cunningham J. K., Sykes J., Kish S. J. (2012): Increased risk of Parkinson's disease in individuals hospitalized with conditions related to the use of methamphetamine or other amphetamine-type drugs. Drug Alcohol Depend. 120, 35–40

http://dx.doi.org/10.1016/j.drugalcdep.2011.06.013

Curtin K., Fleckenstein A. E., Robison R. J., Crookston M. J., Smith K. R., Hanson G. R. (2015): Methamphetamine/amphetamine abuse and risk of Parkinson's disease in Utah: A population-based assessment. Drug Alcohol Depend. 146, 30–38

http://dx.doi.org/10.1016/j.drugalcdep.2014.10.027

Gasparova Z., Stara V., Stolc S. (2014): Effect of antioxidants on functional recovery after in vitro-induced ischemia and long-term potentiation recorded in the pyramidal layer of the CA1 area of rat hippocampus. Gen. Physiol. Biophys. 33, 43–52

http://dx.doi.org/10.4149/gpb_2013062

Hori N., Kadota T., Akaike N. (2014): Functional changes in piriform cortex pyramidal neurons in the chronic methamphetamine-treated rat. Gen. Physiol. Biophys.

Kish S. J., Lerch J., Furukawa Y., Tong J., McCluskey T., Wilkins D., Houle S., Meyer J., Mundo E., Wilson A. A. et al. (2010): Decreased cerebral cortical serotonin transporter binding in ecstasy users: a positron emission tomography/[(11)C]DASB and structural brain imaging study. Brain 133, 1779–1797 http://dx.doi.org/10.1093/brain/awq103

Luscher C., Malenka R. C. (2012): NMDA receptor-dependent long-term potentiation and long-term depression (LTP/LTD). Cold Spring Harb. Perspect. Biol. 4

Mokrushin A. A., Pavlinova L. I. (2013): Effects of the blood components on the AMPA and NMDA synaptic responses in brain slices in the onset of hemorrhagic stroke. Gen. Physiol. Biophys. **32**, 489–504

http://dx.doi.org/10.4149/gpb_2013038

Norman L. L., Brugues J., Sengupta K., Sens P., Aranda-Espinoza H. (2010): Cell blebbing and membrane area homeostasis in spreading and retracting cells. Biophys. J. 99, 1726–1733 http://dx.doi.org/10.1016/j.bpj.2010.07.031

Parrott A. C. (2014): The potential dangers of using MDMA for psychotherapy. J. Psychoactive Drugs **46**, 37–43 http://dx.doi.org/10.1080/02791072.2014.873690

Proft J., Faraji J., Robbins J. C., Zucchi F. C., Zhao X., Metz G. A., Braun J. E. (2011): Identification of bilateral changes in TID1 expression in the 6-OHDA rat model of Parkinson's disease. PLoS One **6**, e26045

http://dx.doi.org/10.1371/journal.pone.0026045

Proft J., Weiss N. (2012): Jekyll and Hide: The two faces of amyloid beta. Commun. Integr. Biol. **5,** 405–407 http://dx.doi.org/10.4161/cib.22571

Proft J., Weiss N. (2014): Rectifying rectifier channels in Huntington disease. Commun. Integr. Biol. 7, e29410 http://dx.doi.org/10.4161/cib.29410

Rusyniak D. E. (2013): Neurologic manifestations of chronic methamphetamine abuse. Psychiatr. Clin. North Am. **36**, 261–275

http://dx.doi.org/10.1016/j.psc.2013.02.005

Schröder N., O'Dell S. J., Marshall J. F. (2003): Neurotoxic methamphetamine regimen severely impairs recognition memory in rats. Synapse **49**, 89–96

http://dx.doi.org/10.1002/syn.10210

Simoes P. F., Silva A. P., Pereira F. C., Marques E., Grade S., Milhazes N., Borges F., Ribeiro C. F., Macedo T. R. (2007): Methampheta-

mine induces alterations on hippocampal NMDA and AMPA receptor subunit levels and impairs spatial working memory. Neuroscience **150**, 433–441

http://dx.doi.org/10.1016/j.neuroscience.2007.09.044

Stys P. K., You H., Zamponi G. W. (2012): Copper-dependent regulation of NMDA receptors by cellular prion protein: implications for neurodegenerative disorders. J. Physiol. **590**, 1357–1368

http://dx.doi.org/10.1113/jphysiol.2011.225276

Tai Y., Chen L., Huang E., Liu C., Yang X., Qiu P., Wang H. (2014): Protective effect of alpha-synuclein knockdown on methamphetamine-induced neurotoxicity in dopaminergic neurons. Neural. Regen. Res. 9, 951–958

http://dx.doi.org/10.4103/1673-5374.133146

Tavassoly O., Lee J. S. (2012): Methamphetamine binds to alphasynuclein and causes a conformational change wich can be detected by nanopore analysis. FEBS Lett. **586**, 3222–3228 http://dx.doi.org/10.1016/j.febslet.2012.06.040

Vermeulen K., Van Bockstaele D. R., Berneman Z. N. (2005): Apoptosis: mechanisms and relevance in cancer. Ann. Hematol. **84,** 627–639

http://dx.doi.org/10.1007/s00277-005-1065-x

White S. R., Obradovic T., Imel K. M., Wheaton M. J. (1996): The effects of methylenedioxymethamphetamine (MDMA, "Ecstasy") on monoaminergic neurotransmission in the central nervous system. Prog. Neurobiol. **49**, 455–479 http://dx.doi.org/10.1016/0301-0082(96)00027-5

Wyllie A. H., Kerr J. F., Currie A. R. (1980): Cell death: the significance of apoptosis. Int. Rev. Cytol. **68**, 251–306

http://dx.doi.org/10.1016/S0074-7696(08)62312-8

You H., Tsutsui S., Hameed S., Kannanayakal T. J., Chen L., Xia P., Engbers J. D., Lipton S. A., Stys P. K., Zamponi G. W. (2012): Abeta neurotoxicity depends on interactions between copper ions, prion protein, and N-methyl-D-aspartate receptors. Proc. Natl. Acad. Sci. U.S.A. 109, 1737–174 http://dx.doi.org/10.1073/pnas.1110789109