CLINICAL STUDY

Elevated nucleosome level and oxidative stress in schizophrenia patients

Bahceci B¹, Kokacya MH², Copoglu US², Bahceci I³, Sahin K³, Bagcioglu E⁴, Dokuyucu R⁵

Departments of Psychiatry, Recep Tayyip ErdoganUniversity, School of Medicine, Rize, Turkey. mhkokacya@mku.edu.tr

ABSTRACT

AIM: The aim of this study was to investigate the effect of oxidative stress on nucleosome levels and its relation with the clinical features in schizophrenia patients.

MATERIAL AND METHOD: Thirty schizophrenia patients and 30 healthy controls were enrolled in this study. Patients were diagnosed with schizophrenia according to the 4th edition of the Diagnostic and Statistical Manual for Mental Disorders (DSM-IV). The control group consisted of 30 healthy subjects matched to the patients with regard to age and gender and who had no history of any psychiatric disorder. The severity of schizophrenia symptoms in the patients was evaluated using the Positive and Negative Syndrome Scale (PANSS) and the Clinical Global Impression Severity Scale (CGI-S). Physical and neurological examinations were performed in each of the patients and controls.

RESULTS: Nucleosome, total oxidant levels and OSI values were higher in schizophrenia patients than in controls (p < 0.05). There was no significant difference in the total antioxidant levels. There was a positive correlation between the nucleosome level and PANSS positive subscale (p = 0.028, r = 0.402). There was a positive correlation between TAS and age (p = 0.025, r = 0.289), PANSS total (p < 0.001, r = 0.604). There was a negative correlation between OSI and PANSS total (p = 0.019, r = -0.427), PANSS positive subscale (p = 0.043, r = -0.372). There was a negative correlation between TOS and PANS total (p = 0.028, r = -0.402).

CONCLUSION: In this study we found a correlation between nucleosome level and PANSS positive subscale. To our knowledge, this is the first study that evaluates oxidative stress and nucleosomes released from apoptotic cells together (*Tab. 2, Ref. 50*). Text in PDF www.elis.sk.

KEY WORDS: nuclesome, schizophrenia, oxidative stress.

Introduction

Although the aetiology of schizophrenia is unknown for certain, some deficits like decreased gray matter volume, synaptic indicators and neurophil are reported. These changes show that an interruption of synaptic circuitry is central in schizophrenia patients(1). Generally, patients with schizophrenia have smaller temporal, parietal, and occipital gray matter. In addition, they have ventriculomegaly, and cortical and subcortical volume loss (2). Brain matter loss and neurodegeneration are involved in the pathophysiology of schizophrenia.

Therefore apoptosis is more prominent in schizophrenia patients than in others (3). Cortical Bcl-2 level is diminished in schizophrenia patients. Lower Bcl-2 levels might cause proapop-

¹Departments of Psychiatry, Recep Tayyip ErdoganUniversity, School of Medicine, Rize, Turkey, ²Departments of Psychiatry, Mustafa Kemal, University, School of Medicine, Hatay, Turkey, ³Departments of Microbiology, Recep Tayyip ErdoganUniversity, School of Medicine, Rize, Turkey, ⁴Department of Psychiatry, Afyon Kocatepe University, Faculty of Medicine, Turkey, and ⁵Department of Physiology, Faculty of Medicine, Mustafa Kemal University, Antakya, Turkey

Address for correspondence: M.H. Kokacya, MD, Department of Psychiatry, Mustafa Kemal University, Faculty of Medicine, Turkey. Phone: +90.505.2585954, Fax: +90.326. 224556

totic effects and neuronal atrophy (4). The ratio of Bax to Bcl-2is is increased in schizophrenia, which is indicative of apoptotic tendency (5). Apoptosis increases in cases of synaptic and neuronal loss (1). Therefore, apoptosis may have a role in developing gray matter loss seen in psychoses. Apoptosis is a physiological process of genetically programmed cell death, first described by Kerr et al. (6). Neuronal cell death is observed in neurodegenerative diseases (7). Injured or diseased neurons are eradicated by apoptosis in neurodegenerative disorders (1). Nucleosome, which is composed of double-stranded DNA and histones, is the basic packing unit of DNA (8). The main role of the nucleosome is the packing of DNA, which controls genetic information by regulating protein access. This control mechanism of nucleosome is regulated by post-translational modifications like methylation, acetylation and phosphorylation of histones (8, 9). During apoptosis, nucleosomes are released into the circulation by nuclear endonucleases activation (8). Elevated levels of nucleosomes in the plasma have been found in malignant diseases, sepsis, septic shock, cerebral stroke, and systemic lupus erythematous(9).

Oxidative stress is considered to be involved in the pathophysiology of schizophrenia(6). Dopamine is one of the possible reactive products (6). Excessive oxidant production and/ or insufficient enzymatic and non-enzymatic antioxidant defense leads to oxidative stress (4). Pathophysiological involvement of 587 - 590

oxidative stress in some diseases is due to interference with the metabolism of carbohydrates, proteins, lipids and DNA (4). One of the effects of oxidative stress on DNA is its role in apoptosis. There is an evidence that oxidative stress leads to apoptosis in some cell types(10-12).

The aim of this study was to investigate the effect of oxidative stress on nucleosome levels and its relation with the clinical features in schizophrenia patients.

Materials and method

Patients and controls

Thirty schizophrenic patients (18 female and 12 male) and 30 healthy controls (19 female and 11 male) were enrolled in this study, which was approved by the local ethics committee. Patients were diagnosed with schizophrenia according to the 4th edition of the Diagnostic and Statistical Manual for Mental Disorders (DSM-IV) (13). The control group consisted of 30 healthy subjects matched to the patients with regard to age and gender and who had no history of any psychiatric disorder. Patients were recruited from the Psychiatry Outpatients Clinic of the Department of Psychiatry, School Medicine, RecepTayyip Erdogan University. All subjects were advised about the procedure and signed the informed consent to participate in the study. The severity of schizophrenia symptoms in the patients was evaluated using the Positive and Negative Syndrome Scale (PANSS) (14) and the Clinical Global Impression Severity Scale (CGI-S) (15). Physical and neurological examinations were performed in each of the patients and controls.

Blood sampling

Venous blood samples were collected from the left forearm vein into heparinized tubes. The blood samples were centrifuged at 3.000 rpm for 10 min at 4 $^{\circ}$ C in order to remove plasma. The buffy coat on the erythrocyte sediment was separated carefully. Plasma samples were stored at -80 $^{\circ}$ C until analysis.

Measurement of TAS, TOS and OSI levels

The total antioxidant status (TAS) was determined by the method of Erel, which allows fully automated measurement of the total antioxidant capacity of the body against strong free radicals. Similarly, the total oxidant status (TOS) was determined in an automated system developed by Erel. Oxidative stress index (OSI) was calculated from the TOS/TAS ratio (16).

Measurement of nucleosome level

The serum samples for nucleosome determination were centrifuged at 3,000g for 15 min and treated with 10 mmol/l EDTA (ethylene diamine tetraacetic acid) immediately after centrifugation and stored at –70 °C until further analysis. The commercially available Cell Death Detection ELISAPLUS Kit (Roche Diagnostics Mannheim, Germany) was used according to the manufacturer's instructions for quantification of nucleosome concentrations in serum. Absorbance (OD) of each well was determined with a microtiter plate reader (Multiskan GO, Thermo Scientific) within 5 minutes. Standard curves were fitted using Titri ELISA software,

then the fitted curve was used to convert sample absorbance readings to nucleosome concentration.

Statistical analyses

The data were evaluated by the SPSS version 18.0 (SPSS Inc., Chicago, IL, USA). Both descriptive and analytical statistics were used. Chi-square / Fisher's Exact test was used for comparisons between categorical variables. Normal distribution of continues variables were tested with the one way Kolmogorov–Smirnov test. Between-group comparisons with normal distribution values were made by the Student'st-test. The Mann–Whitney U test was employed to compare the not normal distribution values. The 2-tailed significance level was set at 0.05. For correlation evaluations, Pearson's and Spearmen's correlation (2-tailed) was used.

Results

30 patients with schizophrenia and 30 age and gender-matched control subjects were included in the study. The socio-demographic characteristics of the study participants are shown in the Table 1.

Nucleosome, total oxidant levels and OSI values were higher in schizophrenia patients than in controls (p < 0.05). There was no significant difference in the total antioxidant levels. Nucleosome, TAS and TOS levels and OSI values are shown in the Table 2.

There was a positive correlation between the nucleosome level and PANSS positive subscale (p = 0.028, r = 0.402). There was a positive correlation between TAS and age (p = 0.025, r = 0.289), PANSS total (p<0.001, r = 0.604). There was a negative correlation between OSI and PANSS total (p = 0.019, r = -0.427), PANSS

Tab. 1. Socio-demographic characteristics of the study participants.

a		
Schizophrenia	Control	p
patients (n=30)	(n=30)	
27.8±3.4	29.1± (6.1)	0.327
		0.791
18 % (48.6)	19 % (51.4)	
12 % (52.2)	11 % (47.8)	
		0.018
7 % (23.3)	17 % (56.7)	
21 % (70.0)	13 % (43.3)	
2 % (6.7)	0	
		0.063
13 % (43.3)	10 % (33.3)	
17 % (56.7)	15 % (50.0)	
0	5 % (16.7)	
		0.108
8 % (26.7)	14 % (46.7)	
22 % (73.3)	16 % (53.3)	
	patients (n=30) 27.8±3.4 18 % (48.6) 12 % (52.2) 7 % (23.3) 21 % (70.0) 2 % (6.7) 13 % (43.3) 17 % (56.7) 0 8 % (26.7)	patients (n=30) (n=30) 27.8±3.4 29.1± (6.1) 18 % (48.6) 19 % (51.4) 12 % (52.2) 11 % (47.8) 7 % (23.3) 17 % (56.7) 21 % (70.0) 13 % (43.3) 2 % (6.7) 0 13 % (43.3) 10 % (33.3) 17 % (56.7) 15 % (50.0) 0 5 % (16.7) 8 % (26.7) 14 % (46.7)

Tab. 2. Nucleosome, TAS and TOS levels and OSI value of cases.

	Schizophrenia patients (n=30)	Control (n=30)	p
Nucleosome (mean±sd)	2.9±0.7	2.6±0.4	0.014
TAS (median. min-max)	1.4. 0.8-4.0	$1.2.\ 1-1.8$	0.096
TOS (mean±sd)	17.9±10.6	10.3 ± 4.8	0.001
OSI (median. min-max)	9.7. 2.7-55.7	7.2. 4-22.6	0.005

 $TAS-Total\ antioxidant\ status,\ TOS-Total\ oxidant\ status,\ OSI-Oxidative\ Stress\ Index$

positive subscale (p = 0.043, r = -0.372). There was a negative correlation between TOS and PANS total (p = 0.028, r = -0.402).

Discussion

TOS level and OSI value were higher in schizophrenia patients compared to controls, but there was no difference in TAS levels. Studies examining the oxidative metabolism in patients with schizophrenia established that TOS level and OSI values and other oxidant parameters were significantly higher in patients with schizophrenia than those in controls (17–20). Although there are studies generally demonstrating increase of oxidants in patients with schizophrenia, a few have shown that there was no change (21, 22). Therefore, studies examining antioxidant parameters in patients with schizophrenia are conflicting. Some studies have shown increased antioxidant levels in patients with schizophrenia (23–26), while some have shown a decrease (22, 27, 28), and still some have found no change (29–31). Our results showed an increase in oxidant levels and no change in antioxidants in patients with schizophrenia.

Nucleosome level was higher in schizophrenia patients compared to controls. Nucleosome is considered to be a marker of apoptosis. Even though apoptosis is frequently determined in schizophrenia and other neurodegenerative diseases, nucleosome measurements have not been widely used. In a recent study, a susceptibility to apoptosis in antipsychotic-naive schizophrenia patients has been found (32). Another study showed an increased apoptosis in euthymic bipolar disorder patients (33). Apoptotic neurons have been shown in some neurodegenerative disorders like Alzheimer's and Huntington's diseases (4, 34, 35).

Oxidative products can cross the membranes and diffuse into the nucleus. They activate the endonucleases by increasing intracellular calcium and cause DNA fragmentation and apoptosis in nucleus (36). Dopamine's neurotoxic and neurodegenerative effects have been shown (6–9, 37). These effects are associated with oxidative metabolism (10–12, 37). Dopamine and 6-OH-DA might induce apoptosis in the presence of oxidants in various neural and non-neural cells (37).

Apoptosis can be activated by a variety of causes in addition to intracellular oxidants and oxidative stress (1, 10, 38). Fatty acid oxidation products are postulated to play a role in apoptosis in a review article (39). Dysregulation in oxidant metabolism such as reduced antioxidants and increased oxidants are observed in schizophrenia (22). Oxidative stress is implicated in the pathophysiology of schizophrenia. Lack of antioxidants might have a role in the aetiology by increasing the apoptosis and causing neuron loss. Several studies support this hypothesis showing a link between decreased GSH (which is an antioxidant enzyme) and increased apoptosis (40). Because of the role of oxidative stress in apoptosis, some antioxidants have been used for preventing apoptosis (41, 42)

We found a positive correlation between nucleosome level and PANSS positive subscale. To our knowledge, there is no data about nucleosome level and disease severity in psychiatric or neurological disorders. However, there is a correlation between disease severity and nucleosome level in some other diseases such as SLE, psoriasis and severe sepsis and septic shock (43–46).

The effect of antipsychotics in apoptosis is controversial. Haloperidol has been found to cause apoptotic neuronal death in a study (47). Atypical antipsychotics up-regulated bcl-2 mRNA which is a potent inhibitor of apoptosis in rat brain (4, 48). This means that typical antipsychotics induce and atypical antipsychotics inhibit apoptosis. However, in an experimental study antipsychotics including haloperidol, clozapine, and quetiapine were shown to activate caspase-3, which is a mediator of apoptosis without an increased DNA fragmentation. Also, it has been found that there was no association between increased Bcl-2 activity and these antipsychotics. This data suggests that antipsychotics may have non-lethal apoptotic effects (49). Another study shows that antipsychotics like haloperidol, eticlopride, raclopride, chlorpromazine and risperidone protect neuronal cells against cell death (50). Caspase-3 activity has been found to be higher in antipsychotic-naïve first episode schizophrenia patients (32). Antipsychotics can be a confounding factor in this study. Therefore, further studies are needed in patients with first-episode and drug naive schizophrenia patients.

In conclusion, in this study we found a correlation between nucleosome level and PANSS positive subscale. To our knowledge, this was the first study that evaluated oxidative stress and nucleosomes released from apoptotic cells together.

References

- 1. Glantz LA, Gilmore JH, Lieberman JA, Jarskog LF. Apoptotic mechanisms and the synaptic pathology of schizophrenia. Schizophrenia Res 2006; 81 (1): 47–63.
- 2. Filippi M, Canu E, Gasparotti R, Agosta F, Valsecchi P, Lodoli G et al. Patterns of brain structural changes in first-contact, antipsychotic drug-naive patients with schizophrenia. AJNR Amer J Neuroradiol 2014; 35 (1): 30–37.
- 3. Pérez–Neri I, Ramírez–Bermúdez J, Montes S, Ríos C. Possible mechanisms of neurodegeneration in schizophrenia. Neurochem Res 2006; 31 (10): 1279–1294.
- **4. Jarskog LF, Gilmore JH, Selinger ES, Lieberman JA.** Cortical bcl–2 protein expression and apoptotic regulation in schizophrenia. Biol Psychiat 2000; 48 (7): 641–650.
- **5. Fatemi SH, Folsom TD.** The neurodevelopmental hypothesis of schizophrenia, revisited. Schizophr Bull 2009; **35** (3): 528–548.
- **6. Catts VS, Catts SV.** Apoptosis and schizophrenia: is the tumour suppressor gene, p53, a candidate susceptibility gene? Schizophr Res 2000; 41 (3): 405–415.
- **7. Friedlander RM.** Apoptosis and caspases in neurodegenerative diseases. New Engl J Med 2003; 348 (14): 1365–1375.
- **8. Decker P.** Nucleosome autoantibodies. Clin Chim Acta 2006; 366 (1): 48–60
- 9. Chen Q, Ye L, Jin Y, Zhang N, Lou T, Qiu Z et al. Circulating nucleosomes as a predictor of sepsis and organ dysfunction in critically ill patients. Intern J Infect Dis 2012; 16 (7): 558–564.
- **10.** Aizenman E, Stout AK, Hartnett KA, Dineley KE, McLaughlin B, Reynolds IJ. Induction of neuronal apoptosis by thiol oxidation. J Neurochem 2000; 75 (5): 1878–1888.
- 11. Esteve J, Mompo J, De La Asuncion JG, Sastre J, Asensi M, Boix J et al. Oxidative damage to mitochondrial DNA and glutathione oxidation in apoptosis: studies in vivo and in vitro. FASEB J 1999; 13 (9): 1055–1064.
- 12. Kagan VE, Gleiss B, Tyurina YY, Tyurin VA, Elenström-Magnusson C, Liu SX et al. A role for oxidative stress in apoptosis: oxidation and ex-

- ternalization of phosphatidylserine is required for macrophage clearance of cells undergoing Fas-mediated apoptosis. J Immunol 2002; 169 (1): 487–499.
- **13. American Psychiatric A.** Diagnostic and statistical manual of mental disorders: DSM–IV–TR®: American Psychiatric Pub; 2000.
- **14. Kay SR, Flszbein A, Opfer LA.** The positive and negative syndrome scale (PANSS) for schizophrenia. Schizophr Bull 1987; 13 (2): 261.
- **15. Guy W.** Clinical global impression scale. The ECDEU Assessment Manual for Psychopharmacology Revised Volume DHEW Publ No ADM 76. 1976: 338: 218–222.
- **16.** Erel O. A new automated colorimetric method for measuring total oxidant status. Clin Biochem 2005; 38 (12): 1103–1111.
- 17. Akyol Ö, Herken H, Uz E, Fadıllıoğlu E, Ünal S, Söğüt S et al. The indices of endogenous oxidative and antioxidative processes in plasma from schizophrenic patients: the possible role of oxidant/antioxidant imbalance. Progr Neuro-Psychopharmacol Biol Psychiat 2002; 26 (5): 995–1005.
- **18. Grignon S, Chianetta JM.** Assessment of malondialdehyde levels in schizophrenia: a meta-analysis and some methodological considerations. Progr Neuro–Psychopharmacol Biol Psychiat 2007; 31 (2): 365–369.
- **19. Zhang M, Zhao Z, He L, Wan C.** A meta–analysis of oxidative stress markers in schizophrenia. Sci China Life Sci 2010; 53 (1): 112–124.
- **20. Zhang XY, Tan YL, Cao LY, Wu GY, Xu Q, Shen Y et al.** Antioxidant enzymes and lipid peroxidation in different forms of schizophrenia treated with typical and atypical antipsychotics. Schizophr Res 2006; 81 (2): 291–300.
- 21. Pazvantoglu O, Selek S, Okay IT, Sengul C, Karabekiroglu K, Dilbaz N et al. Oxidative mechanisms in schizophrenia and their relationship with illness subtype and symptom profile. Psychiat Clin Neurosci 2009; 63 (5): 693–700.
- 22. Virit O, Altindag A, Yumru M, Dalkilic A, Savas HA, Selek S et al. A defect in the antioxidant defense system in schizophrenia. Neuropsychobiol 2009; 60 (2): 87–93.
- **23.** Kuloglu M, Ustundag B, Atmaca M, Canatan H, Tezcan AE, Cinkilinc N. Lipid peroxidation and antioxidant enzyme levels in patients with schizophrenia and bipolar disorder. Cell Biochem Func 2002; 20 (2): 171–175.
- 24. Kunz M, Gama CS, Andreazza AC, Salvador M, Ceresér KM, Gomes FA et al. Elevated serum superoxide dismutase and thiobarbituric acid reactive substances in different phases of bipolar disorder and in schizophrenia. Progr Neuro–Psychopharmacol Biol Psychiat 2008; 32 (7): 1677–1681.
- **25.** Sarandol A, Kirli S, Akkaya C, Altin A, Demirci M, Sarandol E. Oxidative—antioxidative systems and their relation with serum S100 B levels in patients with schizophrenia: effects of short term antipsychotic treatment. Progr Neuro—Psychopharmacol Biol Psychiat 2007; 31 (6): 1164–1169.
- **26.** Zhang XY, Zhou DF, Qi LY, Chen S, Cao LY, Xiu MH et al. Superoxide dismutase and cytokines in chronic patients with schizophrenia: association with psychopathology and response to antipsychotics. Psychopharmacol 2009; 204 (1): 177–184.
- 27. Ustundag B, Atmaca M, Kirtas O, Selek S, Metin K, Tezcan E. Total antioxidant response in patients with schizophrenia. Psychiat Clin Neurosci 2006; 60 (4): 458–464.
- **28.** Yao JK, Reddy R, McElhinny LG, van Kammen DP. Reduced status of plasma total antioxidant capacity in schizophrenia. Schizophr Res 1998; 32 (1): 1–8.
- 29. Çöpoğlu ÜS, Vırıt O, Bülbül F, Ünal A, Alpak G, Savaş HA. An Investigation on Oxidative Metabolism and Oxidative DNA Damage Among Schizophrenia Patients with or without Symptomatic Remission. Bull Clin Psychopharmacol 2012; 22 (1): S129.
- **30.** Matsuzawa D, Obata T, Shirayama Y, Nonaka H, Kanazawa Y, Yoshitome E et al. Negative correlation between brain glutathione level and negative symptoms in schizophrenia: a 3T 1H–MRS study. PLoS One 2008; 3 (4): e1944.
- **31.** Yao JK, Reddy RD, van Kammen DP. Human plasma glutathione peroxidase and symptom severity in schizophrenia. Biol Psychiat 1999; 45 (11): 1512–1515.

- **32.** Gasso P, Mas S, Molina O, Lafuente A, Bernardo M, Parellada E. Increased susceptibility to apoptosis in cultured fibroblasts from antipsychotic-naive first-episode schizophrenia patients. J Psychiat Res 2014; 48 (1): 94–101.
- 33. Fries GR, Vasconcelos-Moreno MP, Gubert C, Santos BT, da Rosa AL, Eisele B et al. Early apoptosis in peripheral blood mononuclear cells from patients with bipolar disorder. J Affect Disord 2014; 152–154: 474–477.
- 34. Bredesen DE. Neural apoptosis. Ann Neurol 1995; 38 (6): 839-851.
- **35.** Dragunow M, Faull RLM, Lawlor P, Beilharz EJ, Singleton K, Walker EB et al. In situ evidence for DNA fragmentation in Huntington's disease striatum and Alzheimer's disease temporal lobes. Neuroreport 1995; 6 (7): 1053–1057.
- **36. Halliwell B, Aruoma OI.** DNA damage by oxygen-derived species Its mechanism and measurement in mammalian systems. FEBS Lett 1991; 281 (1): 9–19.
- **37.** Luo Y, Umegaki H, Wang X, Abe R, Roth GS. Dopamine induces apoptosis through an oxidation–involved SAPK/JNK activation pathway. J Biol Chem 1998; 273 (6): 3756–3764.
- **38.** Trinei M, Giorgio M, Cicalese A, Barozzi S, Ventura A, Migliaccio E et al. A p53–p66Shc signalling pathway controls intracellular redox status, levels of oxidation-damaged DNA and oxidative stress–induced apoptosis. Oncogene 2002; 21 (24): 3872–3878.
- **39.** Tang DG, La E, Kern J, Kehrer JP. Fatty acid oxidation and signaling in apoptosis. Biol Chem 2002; 383 (3–4): 425–442.
- **40. Franco R, Cidlowski JA.** Apoptosis and glutathione: beyond an antioxidant. Cell Death Diff 2009; 16 (10): 1303–1314.
- **41. Hockenbery DM, Oltvai ZN, Yin X.M, Milliman CL, Korsmeyer SJ.** Bcl-2 functions in an antioxidant pathway to prevent apoptosis. Cell 1993; 75 (2): 241–251.
- **42.** Liu M, Pelling JC, Ju J, Chu E, Brash DE. Antioxidant action via p53-mediated apoptosis. Cancer Res 1998; 58 (8): 1723–1729.
- **43. Decker P, Wolburg H, Rammensee HG.** Nucleosomes induce lymphocyte necrosis. Eur J Immunol 2003; 33 (7): 1978–1987.
- **44.** Dilek AR, Dilek N, Saral Y, Yuksel D. The relationship between severity of the disease and circulating nucleosomes in psoriasis patients. Arch Dermatol Res 2013; 305 (6): 483–487.
- **45.** Zeerleder S, Stephan F, Emonts M, de Kleijn ED, Esmon CT, Varadi K et al. Circulating nucleosomes and severity of illness in children suffering from meningococcal sepsis treated with protein C. Crit Care Med 2012; 40 (12): 3224–3229.
- **46.** Zeerleder S, Zwart B, Wuillemin WA, Aarden LA, Groeneveld AB, Caliezi C et al. Elevated nucleosome levels in systemic inflammation and sepsis. Crit Care Med 2003; 31 (7): 1947–1951.
- **47. Noh JS, Kang HJ, Kim EY, Sohn S, Chung YK, Kim SU et al.** Haloperidol-induced neuronal apoptosis: role of p38 and c-Jun-NH (2)-terminal protein kinase. J Neurochem 2000; 75 (6): 2327–2334.
- **48. Bai O, Zhang H, Li XM.** Antipsychotic drugs clozapine and olanzapine upregulate bcl-2 mRNA and protein in rat frontal cortex and hippocampus. Brain Res 2004; 1010 (1–2): 81–86.
- **49.** Jarskog LF, Gilmore JH, Glantz LA, Gable KL, German TT, Tong RI et al. Caspase-3 activation in rat frontal cortex following treatment with typical and atypical antipsychotics. Neuropsychopharmacology: official publication of the American College of Neuropsychopharmacology 2007; 32 (1): 95–102.
- **50. Bastianetto S, Danik M, Mennicken F, Williams S, Quirion R.** Prototypical antipsychotic drugs protect hippocampal neuronal cultures against cell death induced by growth medium deprivation. BMC Neurosci 2006; 7: 28.

Received August 20, 2014. Accepted October 17, 2014.