EXPERIMENTAL STUDY

Are there any effects of Sevoflurane and Desflurane anaesthesia on blood glucose levels in acute hyperglycemic diabetic rats?

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ABSTRACT

AIM: The aim of this study was to investigate blood glucose level of desflurane and sevoflurane on blood glucose in diabetic rats undergoing acute hyperglycemia.

MATERIALS AND METHODS: In this study, 30 male Wistar albino rats were included. Diabetes was induced by a single IP injection of streptozotocin. After the effects of chronic diabetes encountered, diabetic rats were randomly assigned into diabetic control (group DC), diabetic hyperglycemia group (group DH), diabetic hyperglycemia group with desflurane (group DH-D), and diabetic hyperglycemia group with sevoflurane (group DH-S) groups. The normoglycemic groups received an IP injection of the same amount of saline. Hyperglycemic diabetic rats were anaesthetized by desflurane 6% or sevoflurane 2% at a dose, by which minimal alveolar concentration (MAC) for rats would be one. The drugs were given for 4 hours within 100% oxygen at a rate of 6 L.min⁻¹. One hour after cessation of inhalation anaesthesia, blood glucose levels were determined at 1st, 4th and 24th hours. 24 hours after the anaesthesia.

RESULTS: Serum glucose was detected to be significantly lower in Group C, when compared to Groups DC, DH, DH-D and DH-S (p = 0.002, p = 0.001, p = 0.002, p = 0.003, respectively).

Blood glucose levels in the diabetic groups were similar at the end of 6 weeks period, after hyperglycemia and anaesthesia induction.

CONCLUSION: We found out that sevoflurane and desflurane administration in hyperglycemic rats were both related with insignificant blood glucose level increase at early post anaesthesia period and at post anaesthesia 24th hours. We still think that patients undergoing anaesthesia protocols with acute hyperglycemia need relatively longer follow up periods (Tab. 1, Ref. 28). Text in PDF www.ells.sk.

KEY WORDS: diabetes mellitus, acute hyperglycemia, sevoflurane, desflurane, blood glucose level, rat.

Introduction

In the recent two or three decades, the prevalence of diabetes mellitus (DM) has rapidly increased throughout the world, the estimation being that it will increase by 200% in the next few decades (1–5). As a result, physicians will be faced with an increasing population of diabetic patients undergoing anaesthesia and surgery. These patients are carrying high risks for serious cardiovascular complications eventually leading to significant increases in mortality and length of stay rates in hospital (1–3).

Hyperglycemic state of body is generating serious complications, which include impaired wound healing and immune system functions, dehydration and proteolysis at cellular and tissue level. Beside these, hyperosmolar non-ketotic coma is a well-known effect of dehydration due to high serum glucose levels. Poor wound healing in patients with diabetes is a consequence of several factors related to hyperglycemia including impaired blood flow, fibroblast activity and vitamin C uptake, which plays an important role in collagen synthesis (6).

Diabetes mellitus is the leading, but not the only cause of hyperglycemia in vivo. Several others include surgery and general anesthesia leads to a prompt neuroendocrine stress response identified by elevated levels of stress hormones such as: epinephrine, glucagon, cortisol, growth hormone with interleukin-6, tumour necrosis factor-alpha bursts. Activation of stress response results in increased insulin resistance in certain tissues with higher insulin levels, elevated lipolysis and protein degradation, impaired peripheral glucose utilization that results in hyperglycemia and ketosis in some states (7–10).

Different anaesthesia protocols (epidural anaesthesia is related with lower metabolic disarrangements than that general anesthesia) and surgery types (renal and liver transplantation surgery or cardiovascular bypass protocols are commonly associated with hyperglycemia and insulin resistance) lead to different levels of counter regulatory hormone release. Also, limitation or cessation
of the caloric intake at postoperative period can result in impaired glycemic balance (10).

In vitro, volatile anaesthetics such as halothane, enflurane and isoflurane inhibit insulin responses to glucose in a reversible and dose-dependent manner (11–15). The clinical study by Diltoer and Camu showed that glucose tolerance was impaired by isoflurane. In the experimental study (14), halogenated anaesthetic agents, such as halothane or sevoflurane, produced higher negative inotropic effects in diabetic patients compared to normal myocardium, possibly because diabetes exacerbates anaesthetic-induced alterations in troponin-tropomyosin complex activity.

At present, it is uncertain, which anaesthetic agents facilitate adequate glucose control and hemodynamic stability during the perioperative period.

The aim of this study was to investigate blood glucose level of desflurane and sevoflurane on blood glucose in diabetic rats undergoing acute hyperglycemia.

**Material and methods**

**Animals and experimental protocol**

The study was performed upon the approval of Gazi University Experimental Animals Ethics Committee in Gazi University Experimental and Clinical Research Centre (GUDAM). All the procedures were performed according to the accepted standards of the Guide for the Care and Use of Laboratory Animals.

In this study, 30 male Wistar albino rats weighing between 200 and 250 g, raised under the same environmental conditions were used. The rats were kept under 20–21 °C at cycles of 12-hour daylight and 12-hour darkness and had free access to food until 2 hours before the anaesthesia procedure.

The animals were randomly separated into five groups, each containing 6 rats: diabetic control (group DC), diabetic hyperglycemia group (group DH), diabetic hyperglycemia group with desflurane (group DH-D), and diabetic hyperglycemia group with sevoflurane (group DH-S). Another 6 rats without diabetes were assigned as the control group (group C).

STZ (Sigma Chemicals, St. Louis, MO, USA) were prepared by dissolving in saline solution (0.9 % NaCl). STZ was freshly prepared just before the treatment at a dose of 55 mg.kg⁻¹ body weight. Blood glucose levels of diabetic rats were checked by glucometer (mg/dl) 3 days after administration of STZ. Rats were classified as diabetic if their fasting blood glucose (FBG) levels exceeded 250 mg.dl⁻¹, and only animals with FBGs of > 250 mg.dl⁻¹ were included in the diabetic groups (diabetes only, diabetes hyperglycemia, diabetes hyperglycemia plus sevoflurane and diabetes hyperglycemia plus desflurane). The rats were kept alive 6 weeks after streptozotocin injection to allow the development of chronic diabetes before they were exposed to sevoflurane and desflurane.

**Hyperglycemia**

Hyperglycemia was induced with an intraperitoneal injection of 2.5 g.kg⁻¹ of glucose. The normoglycemic groups received an IP injection of the same amount of saline.

Before the study was started, anaesthetic gas vapourisers were calibrated. Anaesthetic gases were set at a minimum alveolar concentration (MAC) of 1 and desflurane 6 % and sevoflurane 2 % were administered. The anaesthesia procedure was conducted with the rats in a transparent plastic container of 40 x 40 x 70 cm in size. The container, which allowed for observations of the rats, was connected to a half open anaesthesia machine with static hoses. The anaesthetic gases were released into the container in 100 % O₂.

The rats were divided into five groups (n = 6). The control, the DC and the DH groups were not subjected to any application. Desflurane (Suprane, Eczacıbaşı, Istanbul, Türkiye) was administered at 6 % inspiratory concentration, 6 L.min⁻¹ in 100 % O₂ for 2 hours, and sevoflurane (Sevorane, Abbot, Istanbul, Turkey) was administered at 2 % inspiratory concentration, 6 L.min⁻¹ in 100 % O₂ for 4 hours. One hour after cessation of inhalation anaesthesia, blood glucose levels were determined at 1st, 4th and 24th hours.

After anaesthesia procedure, all rats were given ketamine 100 mg.kg⁻¹ intraperitoneally.

**Statistical analysis**

Statistical Package for the Social Sciences (SPSS, Chicago, IL, USA) 20.0 program was used for statistical analysis. The importance of the difference of the mean blood glucose levels were assessed by using the Kruskal–Wallis test. Bonferroni adjusted Mann–Whitney U test was used after significant Kruskal–Wallis to determine, which group differs from the other. Results were expressed as the mean ± standard deviation (mean ± SD). Statistical significance was set at a p < 0.05.

**Results**

Blood glucose measurements were 102.50 ± 5.24, 282.25 ± 32.53, 316.25 ± 54.94, 298.33 ± 30.18, 276.25 ± 263.90 mg/dL for Group C, DC, DH, DH-D and DH-S, respectively (Tab. 1).

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<tr>
<th>Tab. 1. Blood glucose levels in rats (mean ± SD).</th>
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<td>Group C (n = 6)</td>
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p – statistical significance was set at a p value < 0.05 for Kruskal–Wallis test, * p < 0.05 – When compared to Group C
Serum glucose was detected to be significantly lower in Group C, when compared to Groups DC, DH, DH-D and DH-S (p = 0.002, p = 0.001, p = 0.002, p = 0.003, respectively).

Blood serum glucose levels after hyperglycemia and anesthesia were measured and similar results were found between study groups (Tab. 1).

Discussion

Emergency surgery rates in diabetic population during life span is reported as 5%. Coronary artery bypass grafting, appendectomy, cholecystectomy, bariatric surgery, laparotomy, abscess drainage, embolectomy and lower extremity amputation are the most commonly performed procedures in this population. Although major procedures such as coronary artery bypass grafting have additional risks, minor interventions can be associated with unwanted surgical and anesthetic outcomes (16).

Pre-anesthetic examination of the patients in diabetic population have to be done with extreme caution. In emergency situations, extensive medical history has to be learned and careful physical examination supported with blood studies have to be done. However, glycemic control in the diabetic patients at the time of emergency surgery is usually insufficient and suboptimal. So precautions have to be done, any impaired glucose level with blood electrolytes, signs of dehydration and acid-base derangements have to be identified before surgery. When identified once, insulin therapy with neutralization of dextrose solutions, correction of acid base disorders and electrolyte imbalances have to be treated (16).

Previous studies showed minimal increases in blood glucose levels with halothane in fed rats (17), and with isoflurane in fed dogs (18). However, in another study, authors showed significant increases in the blood glucose levels for a 3 hr period in fed rats when compared to blood glucose levels determined in fasted rats (200 mg/dl versus 130 mg/dl). These results are similar with those observed with ketamine-xylazine, although both studies have not investigated hyperglycemic states determined after isoflurane and ketamine-xylazine (19).

Acute hyperglycemia is an important prognostic factor in several critical illnesses. Many studies showed strong correlation with acute hyperglycemia and higher mortality rates in acute myocardial infarction (20, 21), cardiovascular surgery (22) and stroke (23, 24). Similarly in animal models of myocardial infarction, deleterious effects of the high blood glucose levels were identified (25, 26). Saha et al (19) showed significantly elevated blood glucose levels in fed rats treated with ketamine plus xylazine (KX) and isoflurane. In contrast, they found that ketamine alone and pentobarbital sodium did not cause any increase in blood glucose levels in both fed and fasted rats. In KX and isoflurane group, decreased plasma levels of insulin, adrenocorticotropic hormone (ACTH), and corticosterone and increased levels of glucagon and growth hormone (GH) were measured. When animals in the KX group were treated with yohimbine (specific alpha2 adrenergic receptor antagonist), they found significant alteration in serum levels of insulin, GH, ACTH and corticosterone. As a consequence, the authors concluded that hyperglycemic effect of KX was driven by its effect on specific alpha2 adrenergic receptors. Additionally, they recommended that agents such as the KX and isoflurane, which resulted in increased blood glucose levels in fed rats, had to be used with caution.

Carraretto et al (27) showed protective effects of propofol and isoflurane on ischaemia reperfusion injury while they couldn’t show the same effect at transient hyperglycemic state. The transient hyperglycemia had a potential injury effect on the kidney after an episode of ischaemia reperfusion. The control of blood glucose levels may be clinically used for limiting organ dysfunction after periods of tissue ischaemia.

Efрати et al (28) studied halothane 2% and reported increased blood glucose levels between 5–60 minutes interval after induction and after 60th minutes they showed decreasing glucose levels which returned pre-anesthesia levels at 120th minutes.

In our study, sevoflurane and desflurane administrations at acute hyperglycemic state insignificantly increased blood glucose levels and these increased levels maintained during 24 hours postoperative period.

In conclusion, we can state that anesthesia protocols in diabetics with acute hyperglycemia can increase the blood glucose levels – an insignificant, but numerically determined effect – and this effect may maintain longer than 24 hours and much longer follow up periods are needed in the patients with acute hyperglycemic state undergoing anesthesia. Additionally we can conclude that both desflurane and sevoflurane can be used safely during acute hyperglycemic state, because of their non-increasing blood glucose effects. Future investigations are needed to further identify the effects of different anesthesia and surgical protocols in acute hyperglycemia.

References


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