### HYPOTHESIS

# Is mitochondrial bioenergetics and coenzyme $Q_{10}$ the target of a virus causing COVID-19?

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#### ABSTRACT

COVID-19 – a coronavirus disease, affected almost all countries in the world. It is a new virus disease, nobody has prior immunity to it, human population is prone to infections. In March 11 2020, WHO declared the pandemic status. The main symptoms include: fever, dry cough and fatigue. Virus proteins need mitochondrial energy for their own survival and replication. Upon viral infections, mitochondrial dynamics and metabolism can be modulated, which can influence the energy production in the host cells. Coenzyme  $Q_{10}$  is an integral component of mitochondrial respiratory chain and the key component of mitochondrial dynamics and metabolism with lower CoQ<sub>10</sub> levels in viral infections leads us to the hypothesis that one of the main pathobiochemical effects of SARS-Cov-2 virus could be mitochondrial bioenergetics dysfunction with CoQ<sub>10</sub> deficit leading to the reduction of its endogenous biosynthesis. The mechanism might be virus induced oxidative stress causing a mutation of one or more of the nine COQ genes, resulting in primary CoQ<sub>10</sub> deficiency. New perspective for patients with COVID-19 may be supportive targeting therapy with coenzyme  $Q_{10}$  to increase the energy production, immunity and decrease oxidative stress (*Fig. 1, Ref. 51*). Text in PDF *www.elis.sk* 

KEY WORDS: COVID-19, virus, mitochondrial bioenergetics, coenzyme Q<sub>10</sub> oxidative stress.

#### Introduction

COVID-19 – a coronavirus disease affected almost all countries in the world. In March 11 2020, WHO declared pandemic status. COVID-19 is the third new corona virus infection in the last years, which caused problems globally in the world. In 2003, a new coronavirus originated from southeast China, was named SARS (Severe Acute Respiratory Syndrome) coronavirus. In 2012, novel coronavirus originated from Middle East, was named MERS (Middle East Respiratory Syndrome) coronavirus (1). At the end of 2019, a novel virus SARS-CoV-2 (severe acute respiratory syndrome coronavirus 2) caused a series of pneumonia in Wuhan (Hubei, China) (2, 3). WHO named the diseases caused by SARS-CoV-2 as "COVID-19" (COrona VIrus Disease 2019) (4).

Coronaviruses (CoVs) belong to the family of *Coronaviridie*, which include four subfamilies:  $\alpha$ -coronavirus,  $\beta$ -coronavirus,

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 $\gamma$ -coronavirus and  $\delta$ -coronavirus. Coronaviruses primarily cause zoonotic infections in birds and mammals and they are capable to infect humans (5). Genome structure of SARS-CoV-2, SARS and MERS belong to the  $\beta$ -coronavirus subfamily (6, 7, 8).

SARS-CoV-2 virus causing COVID-19 is an enveloped positive strand RNA virus with the genome 30-32 kb, the largest known RNA virus. This virus mutates at a high rate and can survive up to 3 hours in droplets. COVID-19 affects mainly seniors with comorbidity, with non-communicable diseases, such as: cardiovascular diseases, diabetes, obesity, chronic lung disease and cancer (9). The structure of SARS-CoV-2 virus contains shaped surface proteins (Fig. 1). SARS-CoV-2 infection is triggered by binding of the spike protein of the virus to Angiotensin-converting enzyme 2 (ACE2), which has been identified as a functional receptor for coronaviruses (10). SARS-CoV-2 virus might pass through the nasal and larynx mucous, then enters the lung through respiratory tract. Virus SARS-CoV-2 primarily infects the lower respiratory tract (11, 12, 13). The main clinical symptoms of CO-VID-19 infections include fever, dry cough, dyspnoea and fatigue. Myalgia or fatigue was in 44–60 % of Asian cases (11, 14). The pathophysiology of COVID-19 includes an induced overproduction of early response proinflammatory cytokines (tumour necrosis factor (TNF), IL-6 and IL-1 $\beta$ ) in the host, what was described as "cytokine storm", leading to increased vascular hyperpermeability and multiorgan failure such as: heart, renal system and gastrointestinal tract (11, 15, 16).

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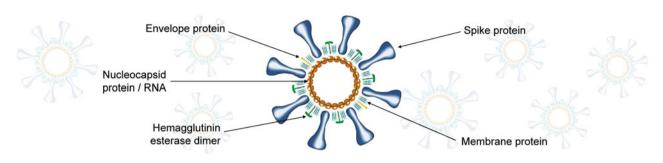


Fig. 1. Structure of SARS-CoV-2 virus (internet).

### Viruses effect on mitochondrial function in organism

Mitochondria, the main source of cells energy production, are found in the cytoplasm of almost all eukaryotic cells. They are important for regulation of the metabolism of carbohydrates, amino acids and fatty acids. Under physiological conditions, mitochondrial oxidative phosphorylation generates a sufficient amount of ATP for cells (17). However, rapidly dividing cells, as activated immune cells or cancer cells, which have the highest request for energy production, receive ATP also by upregulated alternative way, aerobic glycolysis (18, 19). Mitochondria are dynamic structures. The dynamics of mitochondria is controlled by the processes that regulate the morphology of mitochondria. Mitochondrial fission is important process for removing old or damaged mitochondria implicated in stress response and apoptosis. Damaged mitochondria are removed by mitophagy. A small number of large and prolonged mitochondria are formed by fusion. The loss of mitochondrial fusion protein leads to mitochondrial fragmentation (20). Under mitochondrial fragmentation dynamin related protein 1 (DRP-1) is transported from cytosol to the outer mitochondrial membrane and released cytochrome c from mitochondria into the cytoplasm leads to apoptosis (21).

Mitochondria have been implicated in many metabolic functions, in many diseases, in antiviral responses. Virus proteins need mitochondria for their own survival and replication. Mitochondrial antiviral signalling protein (MAVS), associated with the outer mitochondrial membrane, mediates the activation of NF<sub>k</sub>-B and the induction of interferons in response to viral infection (22, 23). Many viruses target mitochondrial dynamics and metabolism, modulate mitochondrial bioenergetics, mitochondrial membrane potential, mitochondrial ion permeability, induce reactive oxygen species production, alter the Ca<sup>2+</sup> regulatory activity and cause oxidative stress in host cells. Viruses can modulate apoptosis and mitochondrial antiviral immunity, alter intracellular distribution of mitochondria, cause host mitochondrial DNA depletion for their survival in the cell (24, 25, 26, 27).

Viral infections activate immune cells for energy production by aerobic glycolysis. Different effect of virus infection on mitochondrial respiratory chain and ATP production was found in the beginning and the progress of the infections. At the beginning of the viral infections, mitochondrial respiration was enhanced related to complex I of the electron transport chain and after the progress of the infection, complex I and complex II of mitochondrial respiratory chain and ATP content decreased (28). Viruses can modulate the host mitochondrial metabolism by different ways. Herpes virus human cytomegalovirus (HCMV) enhances glycolytic flux, directly elevates mitochondrial biogenesis and increases mitochondrial respiration. Herpes simplex virus type-1 (HSV-1) induces Krebs cycle. Hepatitis C virus (HCV) infection enhances mitochondrial fatty acid oxidation (29). Sindbis virus infection causes mitochondrial bioenergetics alterations, which participate in the molecular mechanism of encephalitis (21, 26). Influenza A viral protein targets mitochondria, leads to mitochondrial fragmentation and loss of mitochondrial membrane potential (30). Dengue virus inhibits DRP 1 protein and induces mitochondrial fusion and elongation. Mitochondria play the central role in the primary host defence mechanisms against viral infections and in these processes a number of novel viral and mitochondrial proteins are involved.

SARS-CoV is a large single-positive-strand RNA virus. The virus genome encodes accessory eight open reading frame proteins (ORF). ORF-3a and ORG-8a trigger cellular apoptosis; ORF-7a activates NF-kB; ORF-3b upregulates the expression of several cytokines and chemokines; ORF-6 reduces IFN production; ORF-8b induces cellular DNA synthesis (31). Protein ORF-9b localizes to the outer mitochondrial membrane, alters host cell mitochondrial antiviral signalling. SARS-CoV ORF-9b manipulates host cell mitochondria and mitochondrial functions (32).

Viral infections induce production of reactive oxygen species (ROS) that control replication, as different viruses are able to modulate antioxidative enzymes. Increased ROS production might contribute to the alterations in mitochondrial bioenergetics (21, 32). Exact pathobiochemical mechanisms of SARS-CoV-2 virus effect on mitochondrial bioenergetics is not known. We assume that SARS-CoV-2 virus might manipulate mitochondrial dynamics and metabolism like the SARS-CoV.

# Is coenzyme Q<sub>10</sub> the target of SARS-CoV-2 virus causing CO-VID-19?

Coenzyme  $Q_{10}$  (Co $Q_{10}$ ) was discovered by Frederick Loring Crane in the year 1957. Co $Q_{10}$ , located within the inner mitochondrial membrane, is an integral component of the mitochondrial respiratory chain that transports electrons from complex I and complex II to complex III.  $CoQ_{10}$  is the key component of ATP production in mitochondria. FL Crane showed its effect between a plasma membrane and autism (33, 34, 35, 36).

Primarily, CoQ<sub>10</sub> is endogenously synthetized in the endoplasmic reticulum from tyrosine by mevalonate pathway, including vitamins C,  $B_2$ ,  $B_6$ ,  $B_9$ ,  $B_{12}$  and transported in the plasma by low-density lipoproteins. Currently, nine genes are known to be involved in endogenous CoQ<sub>10</sub> biosynthesis, which are called "COQ genes": COQ2 (coenzyme Q2 - polyprenyltransferase), COO4 (coenzyme O4), COO6 (coenzyme O6 - monooxygenase), COQ7 (coenzyme Q7 - hydroxylase), COQ8A (coenzyme Q8A), COQ8B (coenzyme Q8B), COQ9 (coenzyme Q9), PDSS1 (prenyl diphosphate synthase, subunit 1), PDSS2 (prenyl diphosphate synthase, subunit 2). CoQ<sub>10</sub> biosynthesis involves a number of metabolic reactions such as: methylation, decarboxylation and hydroxylation (37). Low levels of  $CoQ_{10}$  may be caused by a damage to endogenous  $\text{CoQ}_{10}$  synthesis, or by mutation of one or more of the COQ genes, or by its increased utilization. Significantly lower serum level of CoQ<sub>10</sub> were found in patients with an acute influenza infection (15), in patients with chronic kidney disease (38, 39), in endomyocardial biopsies of patients after heart transplantation (40) in patients with cardiomyopathy (41), in infertile men (42, 43).  $CoQ_{10}$  has shown the potential to decrease pain and fatigue in patients with fibromyalgia (44). In patients with septic shock, a significant association between CoQ10 and IL-2 and TNF-alpha was found (45). Statins therapy may also decrease the levels of CoQ<sub>10</sub>, induce mitochondrial dysfunction, fatigue, myopathy and myalgia (46, 47). New roles of  $CoQ_{10}$  in cardiovascular disease discovered by a single group were summarized (35, 48). Targeted therapy of mitochondrial disturbances with  $CoQ_{10}$  was documented (49). A beneficial role of  $CoQ_{10}$  is related to its antioxidant activity, and its effect on cytokine production by human peripheral blood mononuclear cells (PMBC) may modulate human immune function. Authors incubated PMBC with varying doses of CoQ<sub>10</sub> for 24 hours and reported a decreased TNF-alpha and IL-2 secretion in PMBC (50). Coenzyme Q<sub>10</sub> decreases inflammatory biomarkers and cytokines (51).

# Conclusion

Modulated mitochondrial dynamics and metabolism with lower  $CoQ_{10}$  levels in viral infections leads us to the hypothesis that one of the main pathobiochemical effects of SARS-Cov-2 virus could be mitochondrial bioenergetics dysfunction with  $CoQ_{10}$ deficit leading to its reduced endogenous biosynthesis. The mechanism may be virus induced oxidative stress causing a mutation of one or more of nine COQ genes, resulting into primary  $CoQ_{10}$ deficiency. New perspective for patients with COVID-19 may be supportive targeted therapy with coenzyme  $Q_{10}$  to increase energy production, immunity and decrease oxidative stress.

#### References

1. Hilgefeld R, Peiris M. From SARS to MERS: 10 years of research on highly pathogenic human coronaviruses. Antiviral Res 2013; 100: 286–295.

**2. World Health Organization.** WHO Director-General's opening remarks at the media briefing on COVID-19 – 11 March 2020.

**3. Lu H, Stratton CW, Tang YW.** Outbreak of pneumonia of unknown etiology in Wuhan, China. The mystery and the miracle. J Med Virol 2020; 92: 401–402.

**4. Wu YC, Chen CS, Chan YJ**. The outbreak of COVID-19: An overview. J Chin Med Assoc 2020; 83: 217–220.

5. Schoeman D, Fielding BC. Coronavirus envelope protein: current knowledge. Virol J 2019; 16: 19. https://doi.org/10.1016/S0140-3736 (15)60454-8.

6. Cohen J, Normille D. New SARS-like virus in China triggers alarm. Science 2020; 367: 234–235. https://doi.org/10.1126/science.367.6475.234.

7. Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, Zhao X, Huang B, Shi W, Lu R, Niu P, Zhan F, Ma X, Wang D, Xu W, Wu G, Gao GF, Tan W. A novel coronavirus from patients with pneumonia in China, 2019. N Engl J Med 2020; 382 (8): 727–733. https://doi.org/10.1056/NEJMoa2001017.

8. Chen Y, Liu Q, Guo D. Coronaviruses, genome structure, replication and pathogenesis. J Med Virol 2020; 92 (4): 418–423. https://doi.org/10.1002/jmv.25681.

9. Zhang L, Liu Y. Potential interventions for novel coronavirus in China: A systematic review. J Med Virol 2020; 92: 479–490.

10. Shi TT, Yang FY, Liu C, Cao X, Lu J, Zhang XL, Yuan MX, Chen C, Yang JK. Angiotensis-converting enzyme 2 regulates mitochondrial function in pancreatic beta-cells. Biochem Biophys Res Commun 2018; 495 (1): 860–866.

11. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu ZY, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, Wei Y, Wu W, Xie X, Yin W, Li H, Liu M<sup>0</sup>, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Jin Q, Wang J, Cao B. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet 2020; 395: 497–506. Published online 2020/01/24 DOI: 10.1016/S0140-6736 (20)30183-5.

12. Li JY, You Z, Wang Q, Zhou ZJ, Qiu Y, Luo R, Ge XY. The epidemic of 2019-novel-coronavirus (2019-nCoV) pneumonia and insight for emerging infectious diseases in the future. Microbes and Infection 2020; 22: 80–85.

**13. Lupia T, Scabini S, Pinna SM, Di Perri G, De Rosa FG, Corcione S.** 2019 novel coronavirus (2019-nCoV) outbreak: A new challenge. J Global Antimicrobial Resistance 2020; 21: 22–27.

14. Ren LL, Wang YM, Wu ZQ, Xiang ZC, Guo L, Xu T, Yong-Zhong J, Yan X, Yong-Jun L, Xing-Wang L, Hui L, Gou-Hui F, Xiao-Ying G, Yan X, Hong G, Jiu-Yang X, Fan Y, Xin-Ming W, Chaiňo W, Lan C, Yi-Wei L, Bo L, Jian Y, Xiao-Rui W, Jie D, Li L, Chao-Lin H, Jian-Ping Z, Yi H, Zhen-Shun C, Lin-Lin L, Zhao-Hui Q, Chuan Q, Qi J, Bin C, Jian-Wei W. Identification of a novel coronavirus causing severe pneumonia in human: a desriptive study. Chinese Medical Journal 2020; 133 (9): 1015–1024. 10.1097/CM9.0000000000722.

**15. Turner AJ, Hiscox JA, Hooper NM**. ACE2: from vasopeptidase to SARS virus receptor. Trends Pharmacol Sci 2004; 25: 291–294.

16. Chase M, Cocchi MN, Liu X, Andersen LW, Holmberg MJ, Donnino MW. Coenzyme Q10 in acute influenza. Influenza Oher Respi Viruses 2019; 13: 64–70.

**17. Gvozdjakova A, Cornelissen G, Singh RB (Eds).** Recent advances in mitochondrial medicine and coenzyme Q<sub>10</sub>. NOVA Science, NY, USA, 2018, pp.418.

**18. Warburg O, Wind F, Negelein E**: The metabolism of tumors in the body. J Gen Physiol, 1927; 8/6: 519–530.

**19. Zhao H, Raines LN, Ching-Cheng Huang S.** Carbohydrate and amino acid metabolism as hallmarks for innate immune cll activation and function. Cells 2020; 9: 562. DOI: 10.3390/cells9030562

# Bratisl Med J 2020; 121 (11)

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**20. Longo DL, Archer SJ.** Mitochondrial dynamics – mitochondrial fission and fusion in human diseases. N Engl J Med 2013; 369: 2236–2251.

**21. Tiku V, Tan MW, Dikic I**. Mitochondrial functions in infection and immunity. Trends Cell Biol 2020; 30 (4): 263–275.

**22.** Seth RB, Sun L, Ea CK, Chen ZJ. Identification and characterization of MAVS, a mitochondrial antiviral signaling protein that activates NF-*k*B and IRF3. Cell 2005; 122 (5): 669–682.

**23.** Sun Q, Sun L, Liu HH, Chen X, Seth RB, Forman J, Chen ZJ. The specific and essential role of MAVS in antiviral innate immune responses. Immunity 2006; 24: 633–642. DOI: 10.1016/j.immuni.2006.04.004.

**24. Anand SK, Tikoo SK**. Viruses as modulators of mitochondrial functions. Hindawi, Advances in Virology, Volume 2013; Article ID 738794, 17 pages; http://dx.doi.org/10.1155/2013/738794.

**25.** Ohta A, Nishiyama Y. Mitochondria and viruses. Mitochondrion 2011; 11 (1): 1–12.

26. Ripoli M, D'Aprile A, Quarato G, Sarasin-Filipowicz M, Gouttenoire J, Scrima R, Cela O, Boffoli D, Heim MH, Moradpour D, Capitanio N, Piccoli C. Hepatitis C-virus linked mitochondrial dysfunction promotes hypoxia-inductible factor 1 alpha-mediated glycolytic adaptation. J Virology 2010; 84 (1): 647–660.

27. Di Gennaro F, Pizzol D, Marotta C, Antunes M, Racalbuto V, Veronese N, Smith L. Coronavirus diseases (COVID-19) current status and future perspectives: A narrative review. Int J Environ Res Public Health 2020; 17, 2690. DOI: 10.3390/ijerph17082690.

28. Kaarbo M, Ager-Wick E, Osenbroch PO, Kilander A, Skinnes R, Muller F, Eidee L. Human cytomegalovirus infection increases mitochondrial bioegenesis. Mitochondrion 2011; 11 (6): 935–945.

**29. Vastag L, Koyuncu E, Grady SL, Shenk TE, Rabinowitz JD.** Divergent effects of human cytomegalovirus and herpes simplex virus-1 on cellular metabolism. PLOS Pathogens 2011; 7 (7): e1002124. https://doi. org/10.1371/journal.ppat.1002124.

**30. Kalil AC, Thomas PG.** Influenza virus-related critical illness: pathophysiology and epidemiology. Critical care 2019; 23: 258, 7 p.

**31.** McBride R, Fielding BC. The role of severe acute respiratory syndrome (SARS)-coronavirus accessory proteins in virus pathogenesis. Viruses 2012; 4: 2902–2923.

**32.** Shi CH, Qi HY, Boularan C, Huang NN, Abu-Asab M, Shelhamer JH, Kehrl JH. SARS-Coronavirus open reading frame-9b suppresses innate immunity by targeting mitochondria and the MAVS/TRAF3/TRAF6 signalosome. J Immunol 2014; 193: 3080–3089.

**33.** Crane FL, Löw H, Sun I, Navas P, Gvozdjáková A. Plasma membrane coenzyme Q: evidence for a role in autism. Biologics 2014; 8: 199–205.

34. Gvozdjáková A, Kucharská J, Ostatníková D, Babinská K, Nakládal D, Crane FL. Ubiquinol improves symptoms in children with autism. HINDAWI, Oxidative Medicine and Cellular Longevity, 2014; Article ID 798 957, 6 pages, http://dx.doi.org/10.1155/2014/798957

**35.** Gvozdjáková A, Takahashi T, Singh RB, De Meester F, Wilson DG, Crane FL. New roles of coenzyme  $Q_{10}$  in cardiovascular diseases discovered by a single group. World Heart J 2013, 5/3: 159–171.

**36. Teske BF, Sun IL, Gvozdjáková A, Low H, Crane FL.** Plasma membrane CoQ, Porin, and Redox Control of Autism. In: Quinones, eds: ER Price and SC Johnson, 2013; 157–172, Nova Science Publishers, Inc. ISBN: 978-1-62618-323-0

**37. Laredj LN, Licitra F, Puccio HM**. The molecular genetics of coenzyme Q biosynthesis in health and disease. Biochimie 2014; 100: 76–87.

**38. Gvozdjáková A, Sumbalová Z, Kucharská J, Komlósi M, Rausová Z, Vančová O, Számošová M, Mojto V**. Platelet mitochondrial respiration, endogenous coenzyme  $Q_{10}$  and oxidative stress in patients with chronic kidney disease. Diagnostics 2020, 10, 176. DOI: 10.3390/diagnostics10030176

**39.** Gvozdjáková A, Sumbalová Z, Kucharská J, Chládeková A, Rausová Z, Vančová O, Komlosi M, Uličná O, Mojto V. Platelet mitochondrial bioenergetics analysis in patients with nephropathies and non-communicable diseases: a new method. Bratisl Med J 2019; 120/9: 630–635.

**40.** Gvozdjáková A, Kucharská J. Implication of coenzyme Q depletion in heart transplantation. In: Coenzyme Q: Molecular Mechanisms in Health and Disease, eds. Kagan VE, Quinn PJ, CRC Press, Boca Raton, London, New York, Washington, D.C. 2001: 293–304.

**41. Gvozdjáková A, Kucharská J, Dhalla NS, Šimko F.** Mitochondrial cardiology. Gvozdjáková A, Cornélissen G, Singh RB (eds): Recent Advances in Mitochondrial Medicine and Coenzyme Q<sub>10</sub>. NOVA Science, USA, 2018; 131–144.

**42. Gvozdjáková A, Kucharská J, Dúbravický J, Mojto V, Singh RB.** Coenzyme  $Q_{10}$ ,  $\alpha$ -tocopherol, and oxidative stress could be important metabolic biomarkers of male infertility. Disease Markers, Volume 2015, Article ID 827941.

**43. Gvozdjáková A, Dúbravický J, Singh RB.** Mitochondrial reproductive medicne. Gvozdjáková A, Cornélissen G, Singh RB (Eds). Recent Advances in Mitochondrial Medicine and Coenzyme Q<sub>10</sub>. NOVA Science, USA, 2018; 229–240.

**44. Cordero MD, Alcocer-Gómez E, de Miguel M, Culic O, Carrión AM, Alvarez-Suarez JM, Bullón P, Battino M, Fernández-Redriguez A, Sánchz-Alcazar JA**. Can coenzyme Q<sub>10</sub> improve clinical and molecular parameters in fibromyalgia? Antioxidants Redox Signaling, 2013; 19 (12): 1356–1361. DOI: 10.1089/ars.2013.5260 PMID: 23458405.

**45.** Donnino MW, Cocchi MN, Salciccioli JD, Kim D, Naini A, Buetter C, Akuthota P. Coenzyme  $Q_{10}$  levels are low and may be assocaited with the inflammatory cascade in septic shock. Crit Care 2011; 15 (4): R189. http://ccforum.com/content/15/4/R189.

**46.** Gvozdjáková A, Kucharská J, Singh RB, Vančová O, Uličná O, Mojto V, Fedačko J, Pella D, Verma NS, Cornélissen G. Statin-induced mitochondrial dysfunction and targeting coenzyme Q<sub>10</sub> therapy. World Heart J 2016; 8/2: 171–181.

**47. Littarru GP, Tiano L.** Clinical aspects of coenzyme  $Q_{10}$ : an update. Nutrition 2010; 26: 250–254.

**48. Gvozdjáková A, Mikulecký M, Crane FL, Kucharská J, Cornelissen G, Kumar A, Palacka P, Singh RB.** Mitochondrial cardiomyopathy and coenzyme Q<sub>10</sub>. World Heart J 2014; 6 (1): 29–46.

**49. Gvozdjáková A, Kucharská J, de Cabo R, Tiano L, Navas P.** Coenzyme Q<sub>10</sub> targeting therapy of mitochondrial disturbances. Gvozdjáková A, Cornélissen G, Singh RB (eds): Recent Advances in Mitochondrial Medicine and Coenzyme Q<sub>10</sub>. NOVA Science, USA, 2018; 269–292.

**50.** Bessler H, Bergman M, Blumberger N, Djaldetti M, Salman H. Coenzyme  $Q_{10}$  decreases TNF- $\alpha$  and IL-2 secretion by human peripheral blood mononuclear cells. J Nutr Sci Vitaminol 2010; 56 (1): 77–81.

**51. Fan L, Feng Y, Chen GC, Qin LQ, Fu LC, Chen LH**. Effects of coenzyme Q<sub>10</sub> supplementation on inflammatory markers: A systematic metaanalysis of randomized controlled trials. Pharmacol Res 2017; 119: 128–136.

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