

EXPERIMENTAL STUDY

Choline and citicoline ameliorate oxidative stress in acute kidney injury in rats

Elif BARIS¹, Oguzhan SIMSEK², Mualla Aylin ARICI³, Metiner TOSUN¹

Department of Medical Pharmacology, Faculty of Medicine, Izmir University of Economics, Izmir, Turkey.
metiner.tosun@ieu.edu.tr

ABSTRACT

OBJECTIVES: The purpose of this study is to investigate the effects of cholinergic anti-inflammatory pathway (CAP)-activating drugs, choline and citicoline (Cytidinediphosphate-choline, CDP-choline), on lipopolysaccharide (LPS)-induced acute kidney injury (AKI) parameters and the contribution of NADPH Oxidase4 (NOX4) p22phox.

BACKGROUND: Endotoxemia induces a systemic inflammatory response characterized by the production of pro-inflammatory mediators and reactive oxygen species (ROS), which eventually develops acute kidney injury (AKI). NADPH Oxidase4 (NOX4) p22phox pathway contributes to the development of endotoxemia-induced AKI. Inflammatory response can be controlled by CAP.

METHODS: Expressions levels of KIM-1, TNF- α , NOX4, p22phox and NF κ B in the kidney tissues of rats were analyzed via RT-PCR in experimental groups; 1. Control, 2. LPS (10 mg/kg) + saline, 3. LPS + CDP-choline (375 mg/kg) and 4. LPS + choline (90 mg/kg). Choline and ROS levels in kidney tissues were also measured by a spectrofluorometric assay.

RESULTS: LPS-induced elevations of ROS levels were decreased by CDP-choline or choline administration ($p < 0.001$). LPS-elevated KIM-1, TNF α , NOX4, p22 phox, and NF κ B expressions were significantly decreased by choline or CDP-choline treatments ($p < 0.001$).

CONCLUSION: Decreased ROS production in kidney tissues in treatment groups suggests that choline or CDP-choline may have therapeutic potential in endotoxemia-associated AKI via downregulating NOX4 and p22phox expressions (Tab. 1, Fig. 5, Ref. 45). Text in PDF www.elis.sk

KEY WORDS: endotoxemia, choline, cytidine diphosphate choline, acute kidney injury, reactive oxygen species.

Introduction

Acute kidney injury (AKI) is a very common clinical problem in intensive care units with the mortality rates ranging between 26–88 % and may require renal replacement therapy (1). Inflammation and oxidative stress are among the main mechanisms in AKI development. Inflammation can be triggered by some antigenic molecules as well as by lipopolysaccharide (LPS), a gram-negative bacterial endotoxin, which activates inflammatory cells by interacting with toll-like receptors (TLR) (2).

Systemic inflammatory response causes multiple organ dysfunctions due to uncontrolled production of proinflammatory

cytokines and reactive oxygen species (ROS) (3). Inflammation and oxidative stress are closely associated with many pathologies including cancer, cardiovascular, pulmonary and Alzheimer's diseases. Studies showed that expressions of Tumour Necrosis Factor (TNF) α , Interleukin (IL)1 β , Cyclooxygenase (COX)2, inducible Nitric Oxide Synthase (iNOS), Nicotinamide adenine dinucleotide phosphate oxidase (NOX)2, NOX4, and xanthine oxidase increased in heart and kidney tissues of endotoxemic animals (4–6). It has been also shown that LPS-mediated inflammation and oxidative stress contributes to AKI development in endotoxemic animals (7, 8).

NOX is an enzyme which catalyzes superoxide radical formation and ROS production. NOX4 is the most profound isoform of NADPH oxidases in kidney tissues and its stabilization and activation require p22phox, a membrane-bound protein. Studies proposed that NOX4 is regulated at the transcriptional level (9, 10). Previous data showed that, NOX4 expression is induced upon LPS activation in sepsis-associated AKI (11, 12). Malondialdehyde (MDA), hydrogen peroxide (H₂O₂), and cytokine levels increased, whereas catalase and glutathione levels decreased in the brain tissues of endotoxemic mice.

Cholinergic anti-inflammatory pathway (CAP) is a neural mechanism that controls inflammatory response by inhibiting the

¹Department of Medical Pharmacology, Faculty of Medicine, Izmir University of Economics, Izmir, Turkey, ²Institute of Health Sciences, Dokuz Eylul University, Izmir, Turkey, and ³Department of Medical Pharmacology, Faculty of Medicine, Dokuz Eylul University, Izmir, Turkey

Address for correspondence: Metiner TOSUN, Department of Medical Pharmacology, Faculty of Medicine, Izmir University of Economics, Sakarya Caddesi, No: 156, 35330, Balçova, Izmir, Turkey.
Phone: +9002324889843, Fax: +9002322792626

Acknowledgement: The present study was supported by scientific research projects funds from Izmir University of Economics (BAP 2019-03 to MT) and Dokuz Eylul University (2017.KB.SAG.010 to MAA).

Tab. 1. Primers used in real-time quantitative RT-PCR.

Gene	Forward Sequence	Reverse Sequence
<i>NOX4</i>	GTGAACGCCCTGAACTTCTC	TTCTGGGATCCTCATTCTGG
<i>p22 phox</i>	TGTTGCAGGAGTGCTCATCTGTCT	AGGACAGCCCGGACGTAGTAATTT
<i>β-actin</i>	TGCAGAAGGAGATTACTGCC	CGCAGCTCAGTAACAGTCC
<i>TNF-α</i>	AAATGGGCTCCCTTCATCAGTTC	TCTGCTTGGTGGTTTGCTACGAC
<i>KIM-1</i>	ACTCCTGCAGACTGGAATGG	CAAAGCTCAGAGAGCCCATC
<i>NF-κB</i>	CAAAGCTCAGAGAGCCCATC	CCAGGTCATAGAGAGGCTCAA

release of inflammatory cytokines via activating vagus nerve and cholinergic receptors. It has been shown that alpha 7 nicotinic acetylcholine receptors ($\alpha 7nAChR$), members of cholinergic receptors, are required for communication between cholinergic system and immune system cells (13). Alpha7nAChR agonists inhibit the release of cytokines (i.e. TNF- α) and interleukins as well as increasing survival in *in vivo* and *in vitro* inflammation models. Therefore, potential therapeutic role of $\alpha 7nAChR$ agonists in inflammatory disorders has been suggested (13). The role of choline and CDP-choline, a choline donor, is extensively studied *in vivo* models of endotoxemia due, in part, to their agonistic effect on CAP via $\alpha 7nAChR$ interaction (14, 15). Studies demonstrated that CDP-choline and choline increase survival rates in endotoxemic animals and produce positive effects on endotoxin-induced tissue damage and multiple-organ failure (16–18) by CAP-mediated decrease in release of inflammatory mediators from mononuclear cells (19).

Regarding the oxidative stress and CAP interaction, citicoline (Cytidine Diphosphate Choline or CDP-choline), administration decreased MDA levels in the brain tissues of animals with head trauma (20). Administration of carbachol, a cholinomimetic agent, decreases xanthine oxidase and MDA levels in heart tissues of animals with sepsis (21). MDA levels decreased whereas SOD activity increased after citicoline administration in rats with myocardial damage (22). Citicoline reduced ischemia-reperfusion-induced oxidative stress and apoptosis in neonatal rats (23). Phosphorylcholine exerted antioxidant and renoprotective effects by decreasing MDA levels and increasing SOD activity on kidney tissues in rats with contrast induced AKI (24).

We previously showed that choline and CDP-choline administration downregulate LPS-induced Cyclooxygenase (COX)-2 pathway along with prostaglandin production while producing positive effects on sepsis severity and survival of animals with endotoxemia (25). COX pathway and prostaglandin production contribute to oxidative damage in animals with various disorders including inflammation (5, 26). There is insufficient evidence about the effects of choline and CDP-choline in LPS-induced AKI and oxidative stress mediated mechanisms. Therefore, this study investigates effects of CDP-choline and choline on the LPS-induced ROS production via NOX4 pathway in the kidney tissues of the endotoxemic rats.

Materials and methods

Archival kidney samples of male Wistar rats were used in this study. These samples were obtained from our previous study approved by Dokuz Eylul University Multidisciplinary Labo-

ratory Animal Studies Ethics Committee (51/2017). Animal Ethics Committee approval for this study about using collected kidney tissues of rats was also obtained (59/2021).

Four experimental groups archival kidney tissues stored in cryovials at -80°C for molecular analysis were used in this study:

1. Control (n = 6),
 2. LPS+saline (sublethal dose, 10 mg/kg, n = 12),
 3. LPS+ CDP-choline (375 mg/kg, n = 10) and
 4. LPS+ choline (90 mg/kg, n = 10).
- Saline or LPS were administered 5 min after the drug injections. Saline, choline, and CDP-choline injections were repeated at the 6th hour. The CDP-choline dose was determined as equieffective choline dose as reported earlier (25,27). Rats were sacrificed by decapitation at the 24th hour under mild ether anesthesia at the end of the experimental protocol and kidneys were removed and cryopreserved immediately for further analyses.

Chemicals

Lipopolysaccharide (LPS; from *E. coli*), Choline chloride, CDP-choline (Cytidine 5- diphosphocholine sodium salt dihydrate) obtained from Sigma-Aldrich (St. Louis, Mo., USA). All drugs were dissolved in saline (0.9 % sodium chloride) and administered intraperitoneally.

Measurement of reactive oxygen species

Reactive oxygen species (ROS) levels in kidney tissue analyzed with the fluorometric ROS kit (Elabsience, E-BC-K138-F, USA) according to the instructions in the kit (28).

RT-qPCR analysis on *KIM-1*, *TNF-α*, *NOX4*, *p22phox* and *NFκB* expression

Total RNA extraction in kidney homogenates was performed according to the manufacturer instructions (GeneJET RNA Purification Kit, ThermoScientific, USA). cDNA of each sample was synthesized by a commercial kit (ABM 6236, USA) and amplified in an RT-PCR device (Bio-Rad CFXconnect) by using Master Mix (ABM G013, USA). Beta-actin was used as a positive internal control for normalization of the gene transcription. The oligonucleotide primer sequences used in the study are shown in Table 1. $2^{-\Delta\Delta Ct}$ method was used to calculate the relative expression levels (29).

Measurement of total choline levels

Total choline levels in kidney tissue were analyzed via a spectrophotometric total choline/acetylcholine kit (Biovision, K615, USA) according to the manufacturer instructions (25).

Statistical analysis

One-way analysis of variance analysis (ANOVA) with post hoc Tukey–Kramer multiple comparisons for parametric results were employed for group comparisons. Statistical analyses were performed by using GraphPad 5 (La Jolla, USA). Data were shown as mean \pm standard error of the mean (SEM) and $p < 0.05$ was accepted as significant.

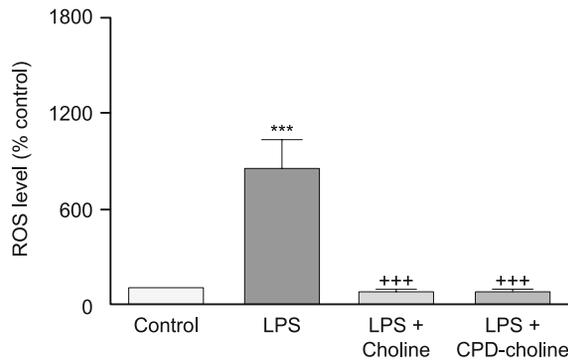


Fig. 1. Relative tissue reactive oxygen levels in kidney tissues of experimental groups. One-way analysis of variance (ANOVA) with post hoc Turkey-Kramer multiple comparison was used for statistical analyses. Data are shown as mean and S.E.M (n = 4–6 in each group). ***, p < 0.001 vs control group; †††, p < 0.001 vs LPS group.

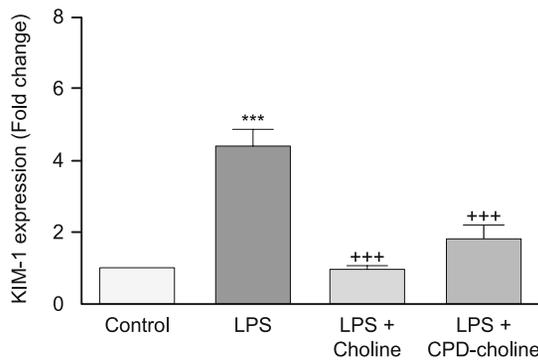


Fig. 2. KIM-1 mRNA expression (fold change) in kidney tissues of experimental groups. $2^{-\Delta\Delta Ct}$ method was employed for the relative quantification of mRNA expression. One-way analysis of variance (ANOVA) with post hoc Turkey-Kramer multiple comparison was used for statistical analyses. Data were shown as mean and S.E.M (n = 4–6 per group). ***, p < 0.001 vs control group; †††, p < 0.001 vs LPS-group.

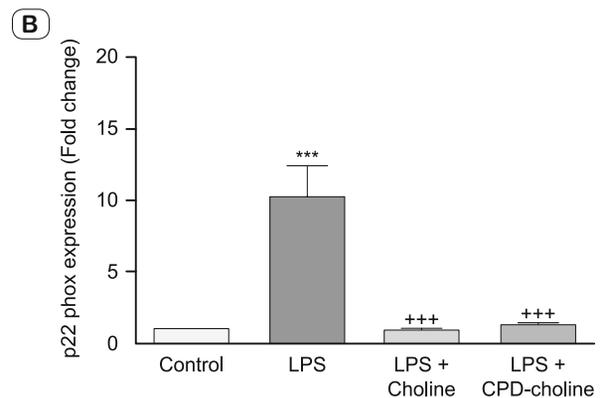
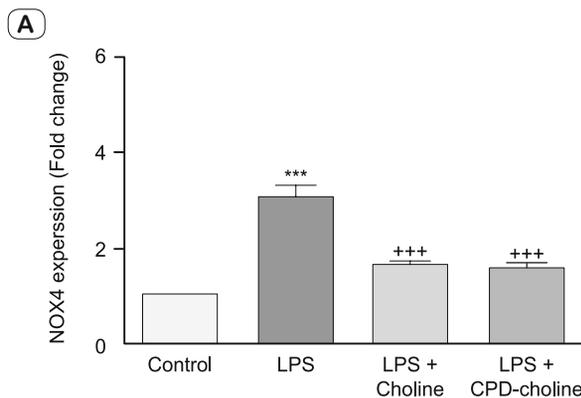


Fig. 3. NOX4 and p22 phox mRNA expression (fold change) in kidney tissues of experimental groups. $2^{-\Delta\Delta Ct}$ method was employed for the relative quantification of mRNA expression. One-way analysis of variance (ANOVA) with post hoc Turkey-Kramer multiple comparison were used for statistical analyses. Data are shown as mean and S.E.M (n = 4–6 per group). ***, p < 0.001 vs control group; †††, p < 0.001 vs LPS-group

Results

ROS levels in kidney tissues

Tissue ROS levels significantly increased in LPS+Saline treated group compared to controls (p < 0.001) while significantly decreased in LPS+Choline and LPS+CDP-choline treated groups compared to LPS+Saline group (p < 0.001) (Fig. 1).

KIM-1 mRNA expression in kidney tissues of endotoxemic animals

Kidney Injury Molecule (KIM)-1, marker of AKI, significantly increased in LPS+Saline treated group compared to control (p < 0.001) while significantly decreased in LPS+Choline and LPS+CDP-choline treated groups compared to LPS+Saline group (p < 0.001) (Fig. 2).

NOX 4 and p22 phox mRNA expressions in kidney tissues

NOX 4 mRNA and p22phox mRNA expressions significantly increased in LPS+Saline treated group compared to controls (p < 0.001), while significantly decreased in LPS+Choline and LPS+CDP-choline treated groups compared to LPS+Saline group (p < 0.001) (Fig. 3).

TNF α and NF- κ B mRNA expressions in kidney tissues

TNF α and NF- κ B expressions, significantly increased in LPS+Saline treated group compared to controls (p < 0.001) while decreased significantly in LPS+Choline and LPS+CDP-choline treated groups comparable to LPS+Saline group (p < 0.001) (Fig. 4).

Total choline levels in kidney tissues

Total Ch/ACh levels significantly increased in LPS+Choline and LPS+CDP-choline treated groups comparable to control group (p < 0.01 and p < 0.05, respectively) (Fig. 5).

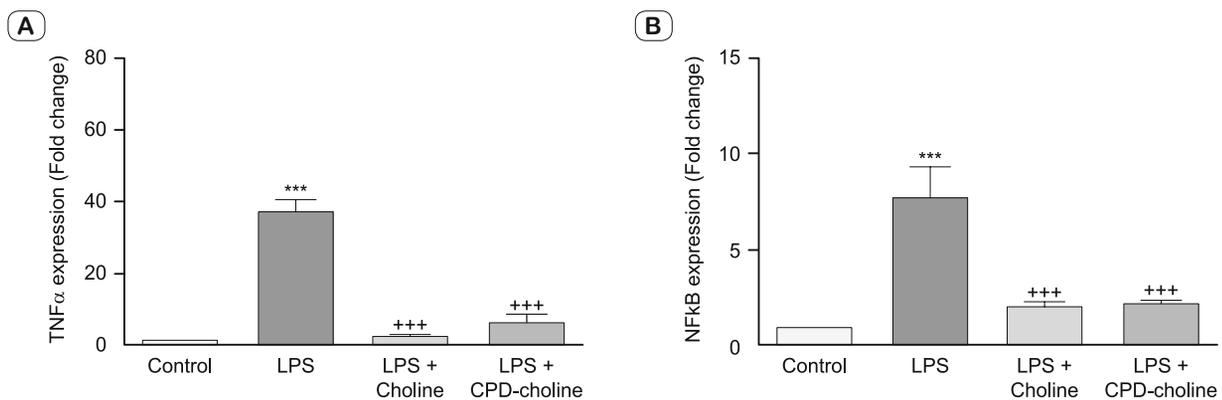


Fig. 4. TNF- α and NF- κ B mRNA expression (fold change) in kidney tissues of experimental groups. $2^{-\Delta\Delta Ct}$ method was employed for the relative quantification of mRNA expression. One-way analysis of variance (ANOVA) with post hoc Turkey-Kramer multiple comparison were used for statistical analyses. Data are shown as mean and S.E.M (n = 4–6 per group). ***, p < 0.001 vs control group; †††, p < 0.001 vs LPS-group.

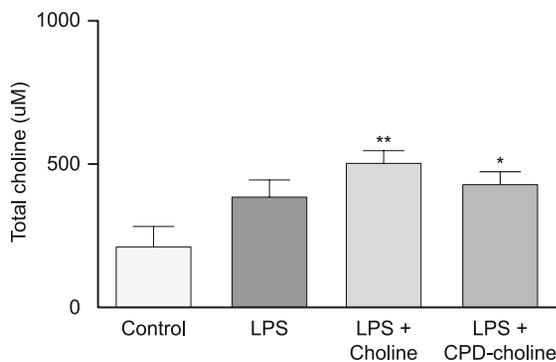


Fig. 5. Total choline levels in kidney tissues of experimental groups. One-way analysis of variance (ANOVA) with post hoc Turkey-Kramer multiple comparison were used for statistical analyses. Data are shown as mean and S.E.M (n = 4–6 in each group). *, p < 0.05; **, p < 0.01 vs control group.

Discussion

This study shows the inhibitory effects of CDP-choline and choline on the LPS-induced ROS production via NOX4 pathway in the cryopreserved kidney tissue samples isolated from the endotoxemic rats.

Kidney Injury Molecule-1 (KIM-1) is a transmembrane protein expressed in the proximal tubules which increases with acute kidney injury (AKI) and accepted as an early biomarker (30, 31) induced in LPS-induced AKI (32, 33). Studies showed that elevated KIM-1 expressions are correlated with acute tubular damage and acute tubular necrosis (34). KIM-1 was demonstrated as an early biomarker in the diagnosis of septic AKI and correlated with poor prognosis in clinical practice (35). There is a lack of evidence regarding the interaction between KIM-1 and choline-containing compounds. Earlier study showed an increase in choline radioactivity in regenerating kidney tissues in rats with nephropathy induced by mercury, suggesting an association between renal regeneration

and choline-containing phospholipids (36). In this study, KIM-1 levels significantly increased in LPS group, comparable to control. Briefly, significant decrease in KIM-1 levels by CDP-choline and choline suggests that these agents may have a clinical impact on prophylaxis of endotoxemia-induced AKI.

The pathophysiology of sepsis-induced AKI includes ischemic or toxic injury to the renal tubular epithelium leading to necrosis, apoptosis and production of reactive oxygen species (ROS), which is characterized clinically as tubular damage (37, 38). NADPH oxidases (NOX) are important sources of ROS production, and NOX4 isoform is widely expressed in the kidney. Stabilization and activation of NOX4 requires p22 phox protein interaction (39). Elevations of NOX4 and p22 phox expressions were demonstrated in different kidney injury models *in vivo* (29, 40, 41). Previous data showed that NOX4 expression induced upon LPS activation in sepsis was associated with AKI (11, 12). Choline downregulates angiotensin II-mediated NOX expression and mitochondrial ROS via muscarinic ACh receptor activation in vascular smooth muscle cells *in vitro* (42). In summary, significant decreases in LPS-induced NOX4 and p22phox expressions may account for the possible therapeutic effectiveness of choline and CDP-choline in endotoxemia.

Inflammation and oxidative stress are closely related with sepsis-induced AKI. Studies showed that expressions of TNF α , IL1 β , COX2, iNOS, NOX2, NOX4, and xanthine oxidase increased in heart and kidney tissues of endotoxemic animals (4, 6, 43). Malondialdehyde (MDA) and hydrogen peroxide (H₂O₂), cytokine levels increased, and catalase and glutathione levels decreased in brain tissues of endotoxemic mice while $\alpha 7nAChR$ protein, ACh levels and choline acetyl transferase activities decreased (44). These observations were supported by the previous histomorphological evidence that endotoxemia-induced kidney damage along with hemodynamic parameters were significantly improved by CDP-choline administration (45). In addition to previous findings, the present study further suggests a therapeutic potential of choline and CDP-choline prevention of LPS-elevated

ROS, TNF α and NF- κ B levels possibly due to the inhibition of NOX4 and p22phox expressions in rat kidney.

Conclusion

Taken together, this study showed that CDP-choline and choline administrations significantly decreased LPS-induced NOX4 and p22phox expressions by increasing total choline levels resulting in decrease in the ROS levels in rat kidney tissues. Together with previous data, our results suggest that CDP-choline and choline may have a therapeutic value in LPS-induced AKI and that further studies can be conducted to evaluate the clinical implications of these findings.

Learning points

Citicoline (CDP-choline) and choline treatments decreased KIM-1 levels that indicate acute kidney injury (AKI) in rats with endotoxemia.

Citicoline (CDP-choline) and choline decreased AKI via down-regulating LPS induced NOX4 and p22phox expressions along with ROS.

Citicoline (CDP-choline) and choline decreased ROS levels via increasing total choline levels in kidney tissues.

References

1. Ympa YP, Sakr Y, Reinhart K, Vincent JL. Has mortality from acute renal failure decreased? A systematic review of the literature. *Amer J Med* 2005; 118 (8): 827–832.
2. Stearns-Kurosawa DJ, Osuchowski MF, Valentine C, Kurosawa S, Remick DG. The pathogenesis of sepsis. *Ann Rev Pathol* 2011; 6: 19–48.
3. Proniewski B, Kij A, Sitek B, Kelley EE, Chlopicki S. Multiorgan Development of Oxidative and Nitrosative Stress in LPS-Induced Endotoxemia in C57Bl/6 Mice: DHE-Based In Vivo Approach. *Oxidative Med Cell Longevity* 2019; (10): 1–11.
4. Li S-Y, Zhou Y-L, He D-H, Liu W, Fan X-Z, Wang Q et al. Centipeda minima extract exerts antineuroinflammatory effects via the inhibition of NF- κ B signaling pathway. *PhytoMed* 2020; (67): 153164.
5. Lee S Bin, Lee WS, Shin JS, Jang DS, Lee KT. Xanthotoxin suppresses LPS-induced expression of iNOS, COX-2, TNF- α , and IL-6 via AP-1, NF- κ B, and JAK-STAT inactivation in RAW 264.7 macrophages. *International Immunopharmacol* 2017; (49): 21–29.
6. Zhao H, Zhang M, Zhou F, Cao W, Bi L, Xie Y et al. Cinnamaldehyde ameliorates LPS-induced cardiac dysfunction via TLR4-NOX4 pathway: The regulation of autophagy and ROS production. *J Mol Cell Cardiol* 2016; (101): 11–24.
7. Liu X, Lu J, Liao Y, Liu S, Chen Y, He R et al. Dihydroartemisinin attenuates lipopolysaccharide-induced acute kidney injury by inhibiting inflammation and oxidative stress. *BioMed Pharmacother* 2019; (117): 109070.
8. Takahashi K, Mizukami H, Kamata K, Inaba W, Kato N, Hibi C et al. Amelioration of acute kidney injury in lipopolysaccharide-induced systemic inflammatory response syndrome by an aldose reductase inhibitor, Fidarestat. *PLoS One* 2012; 7 (1): e30134.
9. Gorin Y. Nox4 as a potential therapeutic target for treatment of uremic toxicity associated to chronic kidney disease. *Kidney Internat* 2013; 83 (4): 541–543.
10. Zana M, Péterfi Z, Kovács HA, Tóth ZE, Enyedi B, Morel F et al. Interaction between p22phox and Nox4 in the endoplasmic reticulum suggests a unique mechanism of NADPH oxidase complex formation. *Free Rad Biol Med* 2018; (116): 41–49.
11. Yoo J-Y, Cha DR, Kim B, An EJ, Lee SR, Cha JJ et al. LPS-Induced Acute Kidney Injury Is Mediated by Nox4-SH3YL1. *Cell Reports* 2020; (3): 108245.
12. Kim JY, Leem J, Hong HL. Melittin Ameliorates Endotoxin-Induced Acute Kidney Injury by Inhibiting Inflammation, Oxidative Stress, and Cell Death in Mice. *Oxidative Med Cell Longevity* 2021; 1–14.
13. Pavlov VA, Wang H, Czura CJ, Friedman SG, Tracey KJ. The cholinergic anti-inflammatory pathway: a missing link in neuroimmunomodulation. *Mol Med* 2003; 9 (5–8): 125–134
14. Bagdas D, Sonat FA, Hamurtekin E, Sonal S, Gurun MS. The antihyperalgesic effect of cytidine-5'-diphosphate-choline in neuropathic and inflammatory pain models. *Behav Pharmacol* 2011; (5–6): 589–598.
15. Gurun M, Parker R, C Eisenach J, Vincler M. The Effect of Peripherally Administered CDP-Choline in an Acute Inflammatory Pain Model: The Role of α 7 Nicotinic Acetylcholine Receptor. *Anesthesia Analgesia* 2009; 108 (5): 1680–1687.
16. Li XD, Buccafusco JJ. Role of α 7 Nicotinic Acetylcholine Receptors in the Pressor Response to Intracerebroventricular Injection of Choline: Blockade by Amyloid Peptide A β 1-42. *J Pharmacol Exp Ther* 2004; 309 (3): 1206–1212.
17. Pan Z-Y, Wang H. Synergistic interaction between choline and aspirin against acute inflammation induced by carrageenan and lipopolysaccharide. *International Immunopharmacol* 2014; 20 (1): 229–237.
18. Parrish WR, Rosas-Ballina M, Gallowitsch-Puerta M, Ochani M, Ochani K, Yang L-H et al. Modulation of TNF release by choline requires alpha7 subunit nicotinic acetylcholine receptor-mediated signaling. *Mol Med* 2008; 14 (9–10): 567–574.
19. Ilcol YO, Yilmaz Z, Ulus IH. Endotoxin alters serum-free choline and phospholipid-bound choline concentrations, and choline administration attenuates endotoxin-induced organ injury in dogs. *Shock* 2005; 24 (3): 288–293.
20. Menku A, Ogden M, Saraymen R. The protective effects of propofol and citicoline combination in experimental head injury in rats. *Turkish Neurosurg* 2010; 20 (1): 57–62.
21. Hu S, Zhang L, Bai H, Bao C. The protective effects of carbachol on visceral ischemia-induced lipid peroxidation injury in rats with sepsis. *Chin Crit Care Med* 2009; 21 (9): 552–554.
22. Huang Y, He Q, Zhan L. The effects of CDP-Choline on the improvement of the successful rate of cardiopulmonary resuscitation and post-resuscitation cardiac function. *Zhonghua Wei Zhong Bing Ji Jiu Yi Xue* 2013; 25 (2): 80–83.
23. González-Pacheco H, Méndez-Domínguez A, Hernández S, López-Marure R, Vazquez-Mellado MJ, Aguilar C et al. Pre-conditioning with CDP-choline attenuates oxidative stress-induced cardiac myocyte death in a hypoxia/reperfusion model. *ScientificWorldJ* 2014; 187071.
24. Aksu F, Aksu B, Unlu N, Karaca T, Ayvaz S, Erman H et al. Antioxidant and renoprotective effects of sphingosylphosphorylcholine on contrast-induced nephropathy in rats. *Renal Fail* 2016; 38 (7): 1089–1098.

25. Baris E, Simsek O, Efe H, Oncu S, Gelal A, Hamurtekin E et al. Effects of CDP-Choline and Choline on COX Pathway in LPS-Induced Inflammatory Response in Rats. *Internat J Pharmacol* 2021; 17: 84–96.
26. Muñoz M, Sánchez A, Pilar Martínez M, Benedito S, López-Oliva M-E, García-Sacristán A et al. COX-2 is involved in vascular oxidative stress and endothelial dysfunction of renal interlobar arteries from obese Zucker rats. *Free Rad Biol Med* 2015; 84: 77–90.
27. Ilcol Y, Yilmaz Z, Cansev M, H. Ulus I. Choline or CDP-choline alters serum lipid responses to endotoxin in dogs and rats: Involvement of the peripheral nicotinic acetylcholine receptors. *Shock* 2009; 32 (3): 286–294.
28. Eid RA, Khalil MA, Alkhateeb MA, Eleawa SM, Zaki MSA, El-Kott AF et al. Exendin-4 Attenuates Remodeling in the Remote Myocardium of Rats After an Acute Myocardial Infarction by Activating β -Arrestin-2, Protein Phosphatase 2A, and Glycogen Synthase Kinase-3 and Inhibiting β -Catenin. *Cardiovascular Drugs Ther* 2020; 35 (6): 1095–1110.
29. Cavdar Z, Oktan MA, Ural C, Kocak A, Calisir M, Heybeli C et al. Alpha lipoic acid attenuates iron induced oxidative acute kidney injury in rats. *Biotech Histochem* 2020; 96 (6): 409–417.
30. Tian L, Shao X, Xie Y, Wang Q, Che X, Zhang M et al. Kidney Injury Molecule-1 is Elevated in Nephropathy and Mediates Macrophage Activation via the Mapk Signalling Pathway. *Cell Physiol Biochem* 2017; (41): 769–783.
31. Jin Y, Shao X, Sun B, Miao C, Li Z, Shi Y. Urinary kidney injury molecule-1 as an early diagnostic biomarker of obstructive acute kidney injury and development of a rapid detection method. *Mol Med Reports* 2017; 15 (3): 1229–1235.
32. Nakano D, Kitada K, Wan N, Zhang Y, Wiig H, Wararat K et al. Lipopolysaccharide induces filtrate leakage from renal tubular lumina into the interstitial space via a proximal tubular Toll-like receptor 4-dependent pathway and limits sensitivity to fluid therapy in mice. *Kidney Internat* 2020; (97): 904–912.
33. Zhang J, Yang S, Chen F, Li H, Chen B. Ginkgetin aglycone ameliorates LPS-induced acute kidney injury by activating SIRT1 via inhibiting the NF- κ B signaling pathway. *Cell Biosci* 2017; 7 (1): 44.
34. Cai J, Jiao X, Luo W, Chen J, Xu X, Fang Y et al. Kidney injury molecule-1 expression predicts structural damage and outcome in histological acute tubular injury. *Renal Failure* 2019; (41): 80–87.
35. Tu Y, Wang H, Sun R, Ni Y, Ma L, Xv F et al. Urinary netrin-1 and KIM-1 as early biomarkers for septic acute kidney injury. *Renal Failure* 2014; (36): 1559–1563.
36. Toback FG, Havener LJ, Dodd RC, Spargo BH. Phospholipid metabolism during renal regeneration after acute tubular necrosis. *Amer J Physiol* 1977; 232 (2): 216–222.
37. Klenzak J, Himmelfarb J. Sepsis and the kidney. *Crit Care Clin* 2005; 21 (2): 211–222.
38. Kim JY, Leem J, Park KK. Antioxidative, antiapoptotic, and anti-inflammatory effects of Apamin in a murine model of lipopolysaccharide-induced acute kidney injury. *Molecules* 2020; 25 (23): 5717.
39. O'Neill S, Knaus UG. Protein-Protein Interaction Assay to Analyze NOX4/p22 (phox) Heterodimerization. *Methods Mol Biol* 2019; (1982): 447–458.
40. Oh SW, Lee ES, Kim S, Na KY, Chae DW, Kim S et al. Bilirubin attenuates the renal tubular injury by inhibition of oxidative stress and apoptosis. *BMC Nephrol* 2013; (14): 105.
41. Zhang H-F, Wang J-H, Wang Y-L, Gao C, Gu Y-T, Huang J et al. Salvianolic Acid A Protects the Kidney against Oxidative Stress by Activating the Akt/GSK-3 β /Nrf2 Signaling Pathway and Inhibiting the NF- κ B Signaling Pathway in 5/6 Nephrectomized Rats. *Oxidative Med Cell Longevity* 2019; 2853534.
42. He X, Deng J, Yu X-J, Yang S, Yang Y, Zang W-J. Activation of M3AChR (Type 3 Muscarinic Acetylcholine Receptor) and Nrf2 (Nuclear Factor Erythroid 2-related Factor 2) Signaling by Choline Alleviates Vascular Smooth Muscle Cell Phenotypic Switching and Vascular Remodeling. *Arterioscler Thrombosis Vasc Biol* 2020; 40 (11): 2649–2664.
43. Sirijariyawat K, Ontawong A, Palee S, Thummasorn S, Maneechote C, Boonphang O et al. Impaired renal organic anion transport 1 (SLC22A6) and its regulation following acute myocardial infarction and reperfusion injury in rats. *Biochim Biophys Acta – Molecular Basis Dis* 2019; 1865 (9): 2342–2355.
44. Han Y-G, Qin X, Zhang T, Lei M, Sun F-Y, Sun J-J et al. Electroacupuncture prevents cognitive impairment induced by lipopolysaccharide via inhibition of oxidative stress and neuroinflammation. *Neurosci Lett* 2018; 683: 190–195.
45. Sevim Ç, Altınbaş B, Yalcın M, Inan S, Özgür Özyiğit M, Arıcan I et al. Protective effect of CDP-choline on hypotension and tissue injury in septic shock model. *Ankara Üniversitesi Veteriner Fakültesi Dergisi* 2017; 64 (2): 103–110.

Received July 19, 2022.

Accepted August 24, 2022.