# CYP19 gene expression and aromatase activity in endometrial cancer tissue: importance of the type of the disease<sup>\*</sup>

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Aromatase (CYP19) activity in malignant endometrium presents local mechanism with potential ability to support tumor growth. The data on interrelation between activity of this enzyme and its mRNA signal in endometrial cancer (EC) tissue are very scarce and inconclusive. To correct this gap we studied aromatase activity and gene expression totally in 19 samples of EC (17 of them – from postmenopausal women) collected during surgery. Aromatase activity was evaluated by tritium water release assay and CYP19 gene expression – with rt-PCR. Additionally, all studied EC cases were divided on the basis of case history and some characteristics of host and tumor and in accordance with existing classification into cases belonging to type I or II of the disease (correspondingly, 13 and 6 observations). Positive correlation between enzyme activity and CYP19 mRNA signal ( $R_s$ =+0.63, p<0.05) was revealed in the whole group of studied samples. Aromatase activity varied in evaluated material between 1.45 fM/mg prot/hr and 11.49 fM/mg prot/hr, and in type II cases it was higher (7.27±0.96 fM/mg prot/hr) than in type I observations (4.96±0.66 fM/mg prot/hr, p=0.066). CYP19 mRNA expression was not revealed in 6 cases and all of them belonged to the type I of disease. Thus, although type II of EC is frequently considered as hormone non-dependent, increased ability of this tumor type to estrogen biosynthesis (on CYP19 gene and protein level) may lead to reconsideration of such conclusion and warrants further investigation.

Key words: endometrial cancer, aromatase, activity, CYP19 mRNA

Estrogenic hyperstimulation is considered – together with progesterone deficiency, hyperinsulinemia/insulin resistance, etc. – among leading risk factors of endometrial cancer [9, 11, 12, 13, 25]. In this context, two main estrogen sources can be distinguished in relation to the target of interests. One of them may be called "exogenous" and, besides HRT and other estrogen containing drugs, includes also estrogen-like environmental substances as well as estrogens circulating in blood. Another source of estrogen is "endogenous", or intratissular, and reflects different processes, among which estrogen uptake and local biosynthesis are most essential [2, 17].

The final step in reactions leading to estrogen formation is based on conversion of androgens to estrogens and is catalyzed by enzyme aromatase. The latter belongs to the XIX class of cytochrome P450 (CYP19). Although not completely unanimously [21], the most popular contemporary concept claims that contrary to the situation with breast epithelium, normal endometrium does not contain aromatase mRNA transcripts, and ability to synthesize estrogens is characteristic only for endometrial cancer (EC) tissue itself [8, 18]. It deserves mentioning that aromatase activity in EC tissue was studied significantly much rarely than in breast cancer, and the scope of analysis of correlation between CYP19 activity and clinico-biological features of EC was understandably very far from exhaustive [5, 18, 23]. Additionally, according to our knowledge, expression of CYP19 gene was so far evaluated in less than 25 cases of EC [8, 19, 21], and practically no comparative studies relating levels of aromatase activity and CYP19 mRNA within endometrial cancer tissue have been performed [19].

Important and rather well known feature of EC is its heterogeneity. Several lines of evidence suggest that two different variants of EC can be distinguished [2, 7, 12, 15, 20]. One of them is traditionally called hormone-dependent (type I) or es-

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trogen-dependent and more rarely – as usual type, and the other one – hormone-independent (type II) or estrogen-independent and as a special type [7, 12, 15, 23]. Initial task of the present study was to compare aromatase activity and CYP19 gene expression in EC tissue and to undertake preliminary steps in direction of the evaluation of these parameters separately in cases belonging to type I or II of endometrial cancer.

# Material and methods

Subjects. Tumor samples from 19 patients with endometrial cancer were evaluated. Patients' age varied from 43 to 77, 17 patients were postmenopausal. According to existing surgical classification, stage of the disease varied between FIGO IA–IIIC. In 17 cases, or in 89.5 %, it was in a range of IA–IIB (Tab. 1). Morphologically mostly endometrioid adenocarcinomas were presented in this material. To distinguish between patients with type I and type II of EC following criteria were used: absence or presence of infertility/low pregnancy number, late menopause, obesity, hypertension, hyperlipidemia, uterine myoma, endometriosis, ovarian theca hy-

pertrophy, well-, modestly- or low-differentiated carcinomas, superficial (less than 5 mm) or deep invasion into myometrium, involvement of regional lymph nodes. As proposed, patients who are infertile, obese, hypertensive, hyperlipidemic, have menopause timing  $\geq$ 53 years, myoma or endometriosis and well-differentiated node-negative adenocarcinoma with superficial invasion (or are characterized with more than 50–60 % of these features) belong to type I of disease [7]. As a result, in the studied group of patients 13 women had type I of EC and 6 women had type II (Tab. 1).

Aromatase activity in tumor tissue was estimated by measuring of  ${}^{3}\text{H}_{2}\text{O}$  release from  ${}^{3}\text{H}-1$ -androstenedione (NEN, Boston, MA; specific activity, 94 GBq/mM) as described [22]. Briefly, the reaction mixture (which contained tumor homogenate, NADPH regeneration system and labeled androgenic precursor) was incubated for 2 hr at 37 °C. Then reaction was stopped by adding 5 vol of cold chloroform, and 5 % suspension of activated charcoal (Norit A) was added to the water phase. The fraction containing  ${}^{3}\text{H}_{2}\text{O}$  was separated by centrifugation, and counting was performed with dioxane scintillator. Results were presented in fM/mg protein/hr.

*CYP19 gene expression* was studied in the same 19 samples of malignant endometrium. Total RNA was isolated with guanidinium thiocyanate-phenol-chloroform method and checked by spectrophotometry and electrophoresis. Reverse transcription was performed in standard way and its efficiency was verified with PCR with primers to GAPDH-gene. Expession of the coding site of aromatase mRNA was

Table 1. Data on patients, type of disease,	aromatase activity	and CYP19	expression in
tumor tissue			

Initials	Age	MP timing	Stage (FIGO)	Type of disease	AA (fM/mg prot/hr)	CYP19 expression (cond. units)
K.L.A.	43	_	IB	1	3.75	2 (±)
O.K.	74	42	IIB	1	2.09	1 (-)
K.N.A.	65	50	IB	1	2.98	1 (-)
L.Z.	61	53	IC	1	4.05	3 (+)
B.G.	51*	48	IB	2	7.26	2 (±)
K.N.	52	50	IB	1	7.93	2 (±)
V.M.	63	50	IB	1	4.90	2 (±)
F.N.	52	47	IA	1	3.13	1 (-)
B.L.	38	_	IB	1	1.45	1 (-)
J.T.	52	49	IB	2	5.81	2 (±)
T.G.	55	50	IB	1	4.22	1 (-)
T.E.	66	45	IIIC	2	8.06	2 (±)
E.V.P.	52	47	IA	1	6.45	3 (+)
G.L.	70	45	IC	2	6.19	2 (±)
T.P.	77	45	IB	2	11.49	3 (+)
K.M.	63	50	IC	1	8.76	3 (+)
K.L.	62	55	IIIB	1	6.52	1 (-)
Sh.T.	69	52	IB	1	8.25	3 (+)
G.M.	76	50	IIB	2	4.81	3 (+)

evaluated examining the region across exons II and III with primers described by KOOS et al [14] – sense: 5'-GAATATTGGAAGGATGCACAGACT-3', antisense: 5'-GGGTAAAGATCATTTCCAGCATGT-3'. The conditions of reaction were the following: 37 cycles, denaturation – 30 sec. at 95 °C, annealing – 30 sec. at 61 °C, synthesis – 1.5 min. at 72 °C. The products were separated in the polyacrylamide gel and visualized by ethidium bromide staining. The expected size of CYP19 product was 293 bp and as a positive control RNA from human placenta was used. Results were presented in a semiquantitative manner using arbitrary or conditional units: 1 ("–" no signal), 2 ("±" weak positive signal) and 3 ("+" distinct positive signal) (Fig. 1).



Figure 1. Examples of rt-PCR for aromatase gene in endometrial cancer tissue. Sample 1: (+); sample 2: (±); sample 3: (±); sample 4: (-); sample 5: mol. weight markers; sample 6: internal control, water; sample 7: (-); sample 8: (+); sample 9: (±); sample 10: (±).

Statistical analysis was performed by methods allowing for means, standard errors,  $\chi^2$ -values and Spearman correlation on the basis of SigmaPlot program. The differences with p $\leq$ 0.05 were considered as significant.

# Results

Within studied samples of malignant endometrium (n=19)aromatase activity varied between 1.45 fM/mg prot./hr and 11.49 fM/mg prot./hr, respectively (Tab. 1) with average (±SE) on the level of 5.69±0.59 fM/mg prot./hr. Inclination to more higher aromatase activity was revealed in tumors from patients belonging to type II of disease (7.27±0.96 fM/mg prot./hr, n=6) vs. tumors from patients with type I of disease (4.96±0.66 fM/mg prot./hr, n=13; p=0.066). Expression of CYP19 mRNA was not revealed in 6 cases of EC, all of which belonged to type I of disease; 7 cases demonstrated signal on the level "±" and 6 cases – on the level "+". Difference in distribution of aromatase gene signaling between patients/tumors belonging to type I or II of the disease is seen in Tables 1 and 2. In the whole group of studied samples aromatase activity and CYP19 mRNA signal correlated with each other (rank correlation coefficient, or  $R_s = +0.63$ , p); this correlation was more pronounced in type I samples  $(R_s=+0.67)$  than in type II material  $(R_s=+0.33)$ .

 Table 2. Distribution of the data on CYP19 gene expression in endometrial cancers belonging to the different types of the disease

CYP19 expression	Type I	Type II
Negative (-)	6	0
Weak positive (±)	3	4
Distinct positive (+)	4	2

Data are presented as number of corresponding cases.

Difference in distribution of reactions ("-" vs. " $\pm$ " and "+") between type I and type II cases is significant ( $\chi^2 = 4.05$ , p = 0.044).

#### Discussion

According to epidemiological observations, in the developed countries endometrial cancer ranks fourth among female cancers and first among gynecological cancers [12]. Effects of the known EC risk factors (obesity, anovulation, infertility, late menopause, etc.) are frequently thought to be mediated through a leading hormonal mechanism, that is, relatively high estrogen levels along with absolute or relative progesterone insufficiency affecting endometrium [13, 20, 25]. There are two sources of estrogen production in female organism, namely gonadal and extragonadal, and significance of the latter increases especially in postmenopausal period [2, 17]. As demonstrated, similarly to breast, epithelial tumors EC tissue also possesses ability for estrogen biosynthesis [5, 8, 18, 19]. Although idea on heterogeneity of EC and existence of its two variants (one of which is usually considered as hormone/estrogen dependent and the other one as hormone/estrogen independent) became rather popular during last 40–50 years [1, 6, 7, 12, 15, 20, 23], nobody so far studied aromatase (estrogen synthetase) activity and aromatase gene signal separately in tumor samples belonging to type I or II of the disease. Meanwhile, such approach might help at least partly to solve the problem, whether the hormonal imbalance associated with risk of endometrial cancer and excessive estrogenic stimulation is peculiar only for so called type I of this disease.

In studies described above two principal results were received: aromatase activity as well as CYP19 gene signal in EC tissue were increased in cases belonging to the type II of disease and besides there was a positive correlation between these two parameters reaching significance (p<0,05) in the whole group of samples. In one of the previous reports tendency to increased aromatase activity in low-differentiated tumors was revealed [24], and according to our recent observations the ratio of incidence of CYP19 A6A6 genotype (presenting a combination of longer and probably more biologically active allelic polymorphisms) to the frequency of A1A6 and A3A6 genotypes was higher in type II patients (1.0) than in type I patients (0.3) [4].

Thus, notwithstanding that we studied relatively small number of cases, it may be suggested that more aggressive clinically and frequently receptor-negative type II of endometrial cancer [7, 12, 14, 15, 20, 23] is associated with the signs of intratumoral hyperproduction of estrogens both on the level of aromatase protein and gene expression. These data additionally support earlier made assumptions that: 1) EC of type II actually may be also estrogen dependent - with the dependence of neoplastic transformation of endometrium and tumor growth progression only of locally produced estrogens [3], and 2) the existence of two variants of EC is a reflection of two types of hormonal carcinogenesis, i.g. promotional and genotoxic [2, 10, 16]. In case that further studies confirm at least first of these assumptions, such property of endometrial cancer probably might be a sign of increased sensitivity of type II of this disease to the treatment with aromatase inhibitors.

## References

- BERSTEIN LM. Excretion of classical estrogens and total phenolsteroids in endometrial cancer patients. Cand Med Sci Thesis, St. Petersburg 1967.
- [2] BERSTEIN LM. Hormonal carcinogenesis. St. Peterburg: Nauka Publishers, 2000.
- [3] BERSTEIN LM, IMYANITOV EN, KOVALEVSKIJ AJ, MAXIMOV SJ, VASILYEV DA et al. CYP17 and CYP19 genetic polymorphisms in endometrial cancer: association with intratumoral aromatase activity. Cancer Lett 2004; 207: 191–196.
- [4] BERSTEIN LM, IMYANITOV EN, ZIMARINA T, KOVALEVSKIJ A, MAXIMOV S et al. Allelic polymorphism of steroidogenic en-

zymes in two types of endometrial cancer. Internat. Conf. "Sex steroids: an update on estrogen and progesterone actions. Role in cancer, ageing and in reproductive medicine (from research to clinical practice)", Montpellier, France, April 2004: 32.

- [5] BERSTEIN LM, TCHERNOBROVKINA AE, GAMAJUNOVA VB, KOVALEVSKIJ AJ, VASILYEV DA et al. Tumor estrogen content and clinico-morphological and endocrine features of endometrial cancer. J Cancer Res Clin Oncol 2003; 129: 245–249.
- [6] BERTHELSEN H, SVANE H. Cancer of the endometrium and hyperestrinism. Two different modes of origin of endometrial cancer. Danish Med Bull 1956; 3: 236–239.
- [7] BOKHMAN JV. Two pathogenetic types of endometrial carcinoma. Gynecol Oncol 1983; 15: 10–17.
- [8] BULUN SE, ECONOMOS K, MILLER D, SIMPSON ER. CYP19 (aromatase cytochrome P450) gene expression in human malignant endometrial tumors. J Clin Endocrinol Metab 1994; 79: 1831–1834.
- [9] GAMAJUNOVA VB, BOBROV JF, TSYRLINA EV, EVTUSHENKO TP, BERSTEIN LM. Comparative study of blood insulin levels in breast and endometrial cancer. Neoplasma 1997; 44: 123–126.
- [10] CAVALIERI E, FRENKEL K, LIEHR JG, ROGAN E, ROY D. Estrogens as endogenous genotoxic agents – DNA adducts and mutations. J Natl Cancer Inst Monograph 2000; 27: 75–94.
- [11] DILMAN VM. Development, ageing and disease. A new rationale for an intervention strategy. Chur (Switzerland): Harwood Acad Publ 1994.
- [12] EMONS G, FLECKENSTEIN G, HINNEY B, HUSCHMAND A, HEYL W. Hormonal interactions in endometrial cancer. Endocrine-Related Cancer 2000; 7: 227–242.
- [13] GUSBERG S. Estrogens and endometrial cancer an epilogue a la recherch du temps perdu. Gynecol Oncol 1994; 52: 3–9.
- [14] KOOS RD, BANKS PK, INKSTER SE, YUE W, BRODIE AM. Detection of aromatase and keratinocyte growth factor expression in breast tumors using reverse transcription-polymerase

chain reaction. J Steroid Biochem Mol Biol 1993; 45: 217–225.

- [15] KOSHIYAMA M, KONISHII I, FUJII S. Pathology, hormonal aspects, and molecular genetics of the two types of endometrial cancer. The Cancer J 1998; 11: 277–283.
- [16] LIEHR JG. Dual role of oestrogens as hormones and pro-carcinogens: tumour initiation by metabolic activation of oestrogens. Eur J Cancer Prev 1997; 6: 3–10.
- [17] MILLER WR. Estrogen and breast cancer. Austin: R.G. Landes Comp, 1996.
- [18] SASANO H, HARADA N. Intratumoral aromatase in human breast, endometrial, and ovarian malignancies. Endocrine Rev 1998; 19: 593–607.
- [19] SASANO H, KAGA K, SATO S, YAJIMA A, NAGURA H, HARADA N. Aromatase cytochrome P450 gene expression in endometrial carcinoma. Br J Cancer 1996; 74: 1541–1544.
- [20] SHERMAN ME. Theories of endometrial carcinogenesis: a multidisciplinary approach. Mod Pathol 2000; 13: 295–308.
- [21] TARKOWSKI R, SKRZYPCZAK M, WINIARCZYK S, KOTARSKI J, JAKOWICKI JA, JAKIMIUK AJ. Aromatase (P450arom) mRNA expression in normal, hyperplastic and malignant endometrium and aromatase activity in endometrial cancer tissue culture. Ginekol Pol 2000; 71(3): 130–135.
- [22] TILSON-MALLETT N, SANTNER SJ, FEIL PD, SANTEN RJ. Biological significance of aromatase activity in human breast tumors. J Clin Endocrinol Metab 1983; 57: 1125–1128.
- [23] TRITZ D, PIERETTI M, TURNER S, POWELL D. Loss of heterozygosity in usual and special variant carcinomas of the endometrium. Hum Pathol 1997; 28: 607–612.
- [24] WATANABE K, SASANO H, HARADA N, OZAKI M, NIIKURA H et al. Aromatase in human endometrial carcinoma and hyperplasia. Immunohistochemical, in situ hybridization, and biochemical studies. Am J Pathol 1995; 146: 491–500.
- [25] ZELENIUCH-JACQUOTTE A, AKHMEDKHANOV A, KATO I, KOENIG KL, SHORE RE et al. Postmenopausal endogenous oestrogens and risk of endometrial cancer: results of a prospective study. Br J Cancer 2001; 84: 975–981.