

## Bone marrow-derived cells: roles in solid tumor Minireview

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The role of cancer stem cells has been demonstrated for some cancers. Recently, research indicated that solid tumors may originate from bone marrow stem cells. Bone marrow-derived cells have recently been shown to contribute to stromal formation, especially angiogenesis and lymphvasculogenesis. Moreover, the interaction and the cell fusion between cancer cells and bone mesenchymal stem cells could enhance the aggregative ability of cancer cells. Bone marrow derived cells home to tumor-specific pre-metastatic sites to provide a permissive niche for incoming tumor cells. Since bone marrow-derived cells play an important role in carcinogenesis, angiogenesis and metastasis, bone marrow-derived cells are not only the tool for cancer therapy, but also the targets for cancer therapy.

**Key words:** bone marrow, stem cells, tumor, angiogenesis, metastasis, carcinogenesis

Transplantation of bone marrow cells has been successfully used in hematopoietic diseases. Recently, transplantation of genetically modified bone marrow progenitors was used as a vehicle for the transport of gene therapy in many diseases, including tumors [1]. Moreover transplantation of bone marrow cells has been hopeful by used for tissue regeneration and tissue repair [2,3]. Unfortunately, patients undergoing bone marrow transplantation have an increased risk of new solid cancers, especially malignant melanoma and cancers of the buccal cavity, liver, brain or other parts of the central nervous system, thyroid, bone, and connective tissue [4-8]. However, the origin of the cancer by bone marrow transplantation remains elusive. Research on bone marrow derived cell in carcinogenesis has entered a new era of controversy, excitement, and great expectations.

### Bone marrow-derived cells and carcinogenesis

The role of cancer stem cells has been demonstrated for some cancers, such as the cancer of hematopoietic system, breast and brain [9-11]. Recently, the clear similarities between stem cell and cancer stem cell genetic programs were

used as a basis for the proposal that some cancer stem cells could derive from adult stem cells [11]. Adult mesenchymal stem cells (MSC) may be targets for malignant transformation and undergo spontaneous transformation following long-term *in vitro* culture, supporting the hypothesis of cancer stem cell origin [12-14].

Chronic gastric inflammation, which develops as a consequence of *H. pylori*, leads over time to repetitive injury and repair resulting in hyperproliferation, an increased rate of mitotic error, and progression to adenocarcinoma. Wang *et al* reported that chronic infection of C57BL/6 mice with *H. pylori*, induced repopulation of the stomach epithelial cells with bone marrow-derived cells. Subsequently, these cells progressed through metaplasia and dysplasia to intraepithelial cancer [15]. Bone mesenchymal stem cells (BMSC) but not hematopoietic stem cells (HSC) are the target for malignant transformation.

Hepatic oval cells, the liver stem cells involved in some forms of liver regeneration, express many markers, such as C-kit, CD<sub>34</sub> and Thy-1, also found on HSC [16-18]. Moreover, bone marrow stem cells could differentiate into oval cells and hepatocytes [19-21]. Hepatoma cells could produce erythropoietin, colony-stimulating factor and myeloid differentiation antigens (CD14, CD68 and HLA-DR) [22,23]. Thus the hypothesis is that HCC(hepatoma cells) may originate from bone marrow stem

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cells [24]. In kidney allograft recipients, bone marrow derived-stem cells originating from a grafted kidney may migrate to the skin, differentiate, or fuse as keratinocytes that could, rarely, undergo cancer transformation [25]. Similarly, post-transplant Kaposi sarcoma could originate from bone marrow derived-stem cells originating from a grafted kidney [26]. Thus the hypothesis is that many solid tumors may originate from bone marrow stem cells [27].

Chronic infection by *H. pylori* could induce transformation of bone marrow-derived cells transformed to intraepithelial cancer, while bone marrow cells could not progress to HCC in transgenic mouse induced by treatment with diethylnitrosamine and phenobarbital [13, 28]. Moreover, HSC could fuse with a multipotent intestinal progenitor or stem cell. Local injury is important for BMDC fusion and that this fusion mechanism participates in epithelial regeneration [29]. Thus, the hypothesis is that chronic inflammation plays probably an essential role in the malignant transformation of bone marrow cells [24]. The stromal derived factor (SDF)-1 is a chemokines produced during chronic inflammation. SDF-1-CXCR4 (the receptor for SDF-1alpha) axis may be a master regulator of trafficking in both normal and cancer stem cells [30]. Stromal cell derived factor (SDF)-1 and stem cell factor-1 (two factors identified in mobilization and migration of marrow progenitor cells) were up-regulated in *H. pylori* infected mice compared with uninfected age-matched controls [15]. SDF-1-CXCR4 axis may play a pivotal role in the recruitment of bone marrow cells to origin and transformation to cancer cells.

The mechanism of cellular fusion of bone marrow stem cell with host cells in organs has been illustrated by using serially transplanted cells and extensive karyotyping of engrafted cells [31, 32]. Other studies, however, suggest that bone marrow stem cell incorporate into organs through transdifferentiation into tissue specific stem cells [33,34]. Further evidence is presented that bone marrow in addition to CD45<sup>positive</sup> HSCs contains a rare population of heterogeneous CD45<sup>negative</sup> nonhematopoietic tissue committed stem cells. Thus some cancers may originate from the differentiation block of nonhematopoietic tissue committed stem cells [35]. Whether the mechanism occurs through differentiation block, trans-determination, or cell fusion is a key unresolved issue. Moreover, if cancer could originate from bone marrow stem cell, who (HCC, BMSC or tissue committed stem cells) is the real troublemaker?

### Bone marrow-derived cells and angiogenesis

Cancer-stromal interaction is well known to play important roles during cancer progression. The MSC self-renew by proliferation while maintaining their stem-cell phenotype and give rise to differentiated stromal cells. The recruitment of bone marrow-derived vascular endothelial cells and myofibroblasts is required for stromal formation during cancer progression [36]. Approximately 25% of myofibroblasts

in pancreatic insulinomas were bone marrow-derived, and these were concentrated toward the edge of the tumor [37].

The traditional view of angiogenesis and lymphvasculogenesis emphasizes proliferation and migration of vessel wall-associated endothelial cells or the pre-existing local lymphatic network. Recently, the role of bone marrow-derived cells in tumor angiogenesis and lymphvasculogenesis was currently the subject of intense research. The debate focus on the innocent supportive or constitutive role of bone marrow-derived cells in tumor angiogenesis and lymphvasculogenesis [38-44]. Bone-marrow-derived cells have recently been shown to contribute to tumor angiogenesis [45-53]. Transplanted bone marrow cells preferentially home to the vessels of in situ generated tumors rather than of normal organs [54].

A human pancreatic cancer cell line, Capan-1, was transplanted into 5 different sites: subcutaneous tissue, peritoneum, liver, spleen and lung. Dramatically, tumors in the subcutaneous tissue and peritoneum induced desmoplastic stroma and contained bone marrow-derived vascular endothelial cells and myofibroblasts, but weak stromal induction without recruitment of bone marrow-derived vascular endothelial cells or myofibroblasts was observed the liver, spleen and lung humors. Thus the cancer microenvironment is important in the recruitment of bone marrow-derived cells [36].

Recruitment of vascular endothelial growth factor (VEGF)-responsive bone- marrow-derived cells is necessary and sufficient for tumor angiogenesis [55]. Although targeting of either vascular endothelial growth factor receptor(VEGFR)1 or VEGFR2 alone partially blocks the growth of tumors, inhibition of both VEGFR1 and VEGFR2 was necessary to completely ablate the tumor growth [56]. Recruited bone marrow-derived circulating cells summoned by VEGF express a distinct different than endothelial progenitor cells. Retention of bone marrow-derived cells in close proximity to angiogenic vessels is mediated by SDF-1, a chemokine induced by VEGF in activated perivascular myofibroblasts [57].

Hypoxia is a central stimulus of VEGF gene expression [58]. The expression of SDF-1 is directly regulated by hypoxia-inducible factor-1 [59]. BMSC may have the capacity to participate in tumor angiogenesis through regulation of their angiogenic properties in the atmosphere of low oxygen that closely approximates the tumor microenvironment. Expression of membrane-type1-matrix metalloproteinase (MMP) was strongly induced by hypoxia. Functional inhibition of membrane-type1-MMP by a blocking antibody strongly suppressed BMSC ability to migrate and generate capillary-like structures [60]. Thus a hypoxic environment such as that encountered in tumor masses regulates BMSC angiogenic properties by pathways of VEGF-VEGFR and SDF-CXCR4 (Fig.1).

Tumor derived plateletderived growth factor receptor (PDGFR) beta (+) progenitor perivascular cells (PPC) that have the ability to differentiate into pericytes and regulate vessel stability and vascular survival in tumors. A subset of PDGFR beta (+) PPC is recruited from bone marrow to

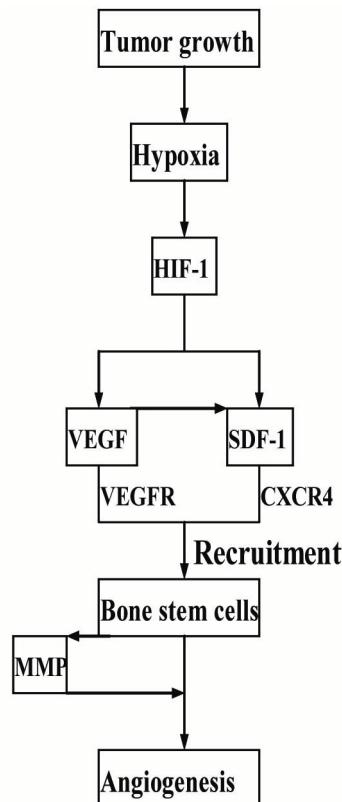
perivascular sites in tumors [61]. In particular, basic fibroblast growth factor, a key mediator of angiogenesis, was found to be the most potent growth factor for inducing BMSC proliferation, migration, and tubulogenesis [62]. Granulocyte colony-stimulating factor (G-CSF) markedly promoted growth of the colon cancer inoculated into the subcutaneous space of mice, whereas G-CSF had no effect on cancer cell proliferation in vitro. G-CSF may have a potential to promote the tumor growth, at least in part, by stimulating angiogenesis in which bone marrow-derived EPC play a role [63]. Loss of Id1 in tumor endothelial cells results in down-regulation of several proangiogenic genes, including alpha6 and beta4 integrins, matrix metalloprotease-2, and fibroblast growth factor receptor-1 [64]. BMSC could be recruited at the sites of active tumor neovascularization through paracrine regulation of their angiogenic properties.

Inhibition of angiogenesis is a promising strategy for treatment of cancer and several other disorders [65,66]. Transplantation of genetically modified bone marrow progenitors may represent a vehicle for the transport of gene therapy to tumors [67-73].

### Bone marrow-derived cells and metastasis

Metastasis is a sequential process including breaking off the cells from the primary tumor, traveling through the bloodstream and stopping at a distant site. Dramatically, bone marrow-derived hematopoietic progenitor cells that express VEGFR1 home to tumor-specific pre-metastatic sites and form cellular clusters before the arrival of tumor cells. VEGFR1+ cells express integrin alpha4beta1, adhesion molecules which joint the cells and ECM, and that tumor-specific growth factors up-regulate fibronectin-a integrin alpha4beta1 ligand-in resident fibroblasts, providing a permissive niche for incoming tumor cells [74]. MMP-9 cell production by multiple myeloma cells is up-regulated in vivo by the interaction of multiple myeloma cells with BMEC [75]. The expression of MMP-9 from bone marrow-derived cells could promote lung tumor colonization of injected tumor cells. MMP-9 from the bone marrow contributes to early survival and establishment of tumors in the lung and has no effect on subsequent growth [76].

After co-culture with BMSC, changes were detected in the morphology, proliferative capacity and aggregation pattern of human breast cancer MCF-7 cells, but these parameters were not affected after the co-culture of BMSC with a non-tumorigenic breast epithelial cell line, MCF-10. VEGF and IL-6 cells mimic the effects produced by BMSC or its products on the proliferation and aggregation properties of MCF-7, cells, respectively [77]. When epithelial metastatic breast cancer MCF-7s cell were set in co-culture with BMSC or in feeder layer of 3T3 fibroblasts, MCF-7 cells grow in clusters on 3T3, but disperse on BMSC. Concomitant with the loss of their aggregation status, MCF-7 cells on BMSC expresses low levels of the intercellular adhesion molecules, E-cadherin and



**Fig.1 Bone stem cells and tumorous angiogenesis**

The expression of SDF-1 and VEGF is directly regulated by HIF-1, which induced by hypoxia. SDF-1 is also induced by VEGF in activated perivascular myofibroblasts. VEGFR(+) and/or CXCR4(+) BMSC may be recruited and have the capacity to participate in angiogenesis under hypoxic microenvironment. Expression of membrane-type1- MMP, which play an important role of migrating and generating capillary-like structures, was strongly induced by hypoxia. Thus a hypoxic environment of tumor masses recruits and regulates BMSC angiogenic properties by pathways of VEGF-VEGFR and SDF-CXCR4. HIF-1: hypoxia-inducible factor-1, MMP: matrix metalloproteinase, SDF-1: Stromal cell derived factor-1, VEGF: vascular endothelial growth factor, VEGFR: vascular endothelial growth factor receptor.

epithelial-specific antigen [78]. Thus the interaction between cancer cells and BMSC could enhance the aggregative ability of cancer cells.

Tumor cell becomes metastatic by fusion to normal cells that travel throughout the body freely, such as lymphocytes or macrophages [79, 80]. It has been hypothesized that bone marrow stem cells could fuse with transformed intestinal epithelium to confer phenotypic and genotypic diversity to the tumor [81,82]. HSC but not BMSC, could fuse with a multipotent intestinal progenitor or stem cell [82]. HSC are not involved in tumor initiation in the small intestine, but with undifferentiated or stem cell-like tumor cells [82]. These fusion events may lead to a cell type that possesses greater metastatic characteristics [81, 83].

## Conclusion

Studies have shown that some cancer cells and even stromal-connected especially with angiogenesis and lymphvasculogenesis, may originate from bone marrow stem cells. Moreover, bone marrow derived cells could enhance the aggregative and metastatic ability of cancer cells. Since bone marrow-derived cells play an important role in carcinogenesis, angiogenesis and metastasis, bone marrow-derived cells are not only the tool for cancer therapy, but also potential targets for cancer therapy. The long biosafety of bone marrow transplant or bone marrow derived cells for patients with or without cancer seems to need still more study.

## References

- [1] KASSEM M. Mesenchymal stem cells: biological characteristics and potential clinical applications. *Cloning Stem Cells* 2004;6:369-74.
- [2] GRENIER G, RUDNICKI MA. The potential use of myogenic stem cells in regenerative medicine. *Handb Exp Pharmacol* 2006;174:299-317.
- [3] KEMP KC, HOWS J, DONALDSON C. Bone marrow-derived mesenchymal stem cells. *Leuk Lymphoma* 2005;46:1531-44.
- [4] BAKER KS, DEFORTE, BURNS LJ, RAMSAY NK, NEGELIA JP, ROBISON LL. New malignancies after blood or marrow stem-cell transplantation in children and adults: incidence and risk factors. *J Clin Oncol* 2003;21:1352-8.
- [5] AU WY, LIE AK, LIANG R, LIU CL, SHEK TW, LAU GK. Aggressive hepatocellular carcinoma complicating pregnancy after autologous bone marrow transplantation for non-Hodgkin's lymphoma. *Bone Marrow Transplant* 2002;29:177-9.
- [6] BHATIA S, LOUIE AD, BHATIA R, O'DONNELL MR, FUNG H, KASHYAP A, et al. Solid cancers after bone marrow transplantation. *J Clin Oncol* 2001;19:464-71.
- [7] SOCIE G, CURTIS RE, DEEG HJ, SOBOCINSKI KA, FILIPOVICH AH, TRAVIS LB, et al. New malignant diseases after allogeneic marrow transplantation for childhood acute leukemia. *J Clin Oncol* 2000;18:348-57.
- [8] CURTIS RE, ROWLINGS PA, DEEG HJ, SHRINER DA, SOCIE G, TRAVIS LB, et al. Solid cancers after bone marrow transplantation. *N Engl J Med* 1997;336:897-904.
- [9] REYA T, MORRISON SJ, CLARKE MF, WEISSMAN IL. Stem cells, cancer, and cancer stem cells. *Nature* 2001;414:105-11.
- [10] PARDAL R, CLARKE MF, MORRISON SJ. Applying the principles of stem-cell biology to cancer. *Nat Rev Cancer* 2003;3:895-902.
- [11] SOLTYSOVA A, ALTANEROVA V, ALTANER C. Cancer stem cells. Minireview. *Neoplasma* 2005;52:435-40.
- [12] RUBIO D, GARCIA-CASTRO J, MARTIN MC, DE LA FUENTE R, CIGUDOSA JC, LLOYD AC, et al. Spontaneous human adult stem cell transformation. *Cancer Res* 2005;65: 3035-9.
- [13] BURNS JS, ABDALLAH BM, GULDBERG P, RYGAARD J, SCHRODER HD, KASSEM M. Tumorigenic heterogeneity in cancer stem cells evolved from long-term cultures of telomerase-immortalized human mesenchymal stem cells. *Cancer Res* 2005; 65: 3126-35.
- [14] SERAKINCI N, GULDBERG P, HATCH H, PI L, DENG J, SCOTT EW. Mouse A6-positive hepatic oval cells also express several hematopoietic stem cell markers. *Hepatology* 2003;37:632-40.
- [19] PETERSEN BE, BOWEN WC, PATRENE KD, MARS WM, SULLIVAN AK, MURASE N, et al. Bone marrow as a potential source of hepatic oval cells. *Science* 1999; 284: 1168-70.
- [20] WU XZ, ZHAO LS, XU Q, ZHANG Y, TANG H. Differentiation of Bone Marrow Mesenchymal Stem Cells into Hepatocytes in Hepatectomized Mouse. *J Biomed Eng* 2005; 22: 1234-7.
- [21] JIANG Y, JAHAGIRDAR BN, REINHARDT RL, SCHWARTZ RE, KEENE CD, ORTIZ-GONZALEZ XR, et al. Pluripotency of mesenchymal stem cells derived from adult marrow. *Nature* 2002; 418:41-9.
- [22] GABRILOVE JL, WELTE K, LU L, CASTRO-MALASPINA H, MOORE MA. Constitutive production of leukemia differentiation, colony-stimulating, erythroid burst-promoting, and pluripotent factors by a human hepatoma cell line: characterization of the leukemia differentiation factor. *Blood* 1985;66:407-15.
- [23] WANG SY, CHEN LY, TSAI TF, SU TS, CHOO KB, HO CK. Constitutive production of colony-stimulating factors by human hepatoma cell lines: possible correlation with cell differentiation. *Exp Hematol* 1996; 24: 437-44.
- [24] WU XZ, CHEN D. Origin of hepatocellular carcinoma: Role of stem cells. *J Gastroenterol Hepatol*. 2006; In press.
- [25] ARACTINGI S, KANITAKIS J, EUVRARD S, et al. Skin carcinoma arising from donor cells in a kidney transplant recipient. *Cancer Res* 2005; 65: 1755-60
- [26] BAROZZI P, LUPPI M, FACCHETTI F, et al. Post-transplant Kaposi sarcoma originates from the seeding of donor-derived progenitors. *Nat Med* 2003; 9: 554-61
- [27] LI HC, STOICOV C, ROGERS AB, HOUGHTON JM. Stem cells and cancer: Evidence for bone marrow stem cells in epithelial cancers. *World J Gastroenterol* 2006;12:363-71.
- [28] ISHIKAWA H, NAKAO K, MATSUMOTO K, NISHIMURA D, ICHIKAWA T, HAMASAKI K, et al. Bone marrow engraftment in a rodent model of chemical carcinogenesis but no role in the histogenesis of hepatocellular carcinoma. *Gut* 2004; 53: 884-9.
- [29] RIZVI AZ, SWAIN JR, DAVIES PS, BAILEY AS, DECKER AD, WILLENBRING H, et al. Bone marrow-derived cells fuse with normal and transformed intestinal stem cells. *Proc Natl Acad Sci U S A* 2006;103:6321-5.
- [30] KUCIA M, RECA R, MIEKUS K, WANZECK J, WOJAKOWSKI W, JANOWSKA-WIECZOREK A, et al. Trafficking of normal stem cells and metastasis of cancer stem cells involve similar mechanisms: pivotal role of the SDF-1-CXCR4 axis. *Stem Cells* 2005;23:879-94.
- [31] VASSILOPOULOS G, WANG PR, RUSSELL DW. Transplanted bone marrow regenerates liver by cell fusion. *Nature* 2003;422:901-4.
- [32] WANG X, WILLENBRING H, AKKARI Y, TORIMARU Y, FOSTER M, AL-DHALIMY M, et al. Cell fusion is the

- principal source of bone-marrow-derived hepatocytes. *Nature*. 2003;422:897-901.
- [33] FERRARI G, CUSELLA-DE ANGELIS G, COLETTA M, PAOLUCCI E, STORNAIUOLO A, COSSU G, *et al*. Muscle Regeneration by Bone Marrow-Derived Myogenic Progenitors. *Science* 1998; 279, 1528-30.
- [34] IANUS A, HOLZ GG, THEISE N D, HUSSAIN MA. In vivo derivation of glucose-competent pancreatic endocrine cells from bone marrow without evidence of cell fusion. *J Clin Invest* 2003;111, 843-50.
- [35] KUCIA M, RECA R, JALA VR, DAWN B, RATAJCZAK J, RATAJCZAK MZ. Bone marrow as a home of heterogeneous populations of nonhematopoietic stem cells. *Leukemia* 2005;19:1118-27.
- [36] SANGAI T, ISHII G, KODAMA K, MIYAMOTO S, AOYAGI Y, ITO T, *et al*. Effect of differences in cancer cells and tumor growth sites on recruiting bone marrow-derived endothelial cells and myofibroblasts in cancer-induced stroma. *Int J Cancer* 2005;115:885-92.
- [37] DIREKZE NC, HODIVALA-DILKE K, JEFFERY R, HUNT T, POULSOM R, OUKRIF D, *et al*. Bone marrow contribution to tumor-associated myofibroblasts and fibroblasts. *Cancer Res* 2004;64:8492-5.
- [38] SHINDE PATIL VR, FRIEDRICH EB, WOLLEY AE, GERSZTEN RE, ALLPORT JR, WEISSLEDER R. Bone marrow-derived lin(-)c-kit(+)Sca-1+ stem cells do not contribute to vasculogenesis in Lewis lung carcinoma. *Neoplasia* 2005;7:234-40.
- [39] MACHEIN MR, RENNINGER S, DE LIMA-HAHN E, PLATE KH. Minor contribution of bone marrow-derived endothelial progenitors to the vascularization of murine gliomas. *Brain Pathol* 2003;13:582-97.
- [40] YUNG YC, CHESHIER S, SANTARELLI JG, HUANG Z, WAGERS A, WEISSMAN I, *et al*. Incorporation of naive bone marrow derived cells into the vascular architecture of brain tumor. *Microcirculation* 2004;11:699-708.
- [41] ZIEGELHOEFFER T, FERNANDEZ B, KOSTIN S, HEIL M, VOSWINCKEL R, HELISCH A, *et al*. Bone marrow-derived cells do not incorporate into the adult growing vasculature. *Circ Res* 2004;94:230-8.
- [42] RELIGA P, CAO R, BJORNDAHL M, ZHOU Z, ZHU Z, CAO Y. Presence of bone marrow-derived circulating progenitor endothelial cells in the newly formed lymphatic vessels. *Blood* 2005;106:4184-90.
- [43] HE Y, RAJANTIE I, ILMONEN M, MAKINEN T, KARK-KAINEN MJ, HAIKOP, *et al*. Preexisting lymphatic endothelium but not endothelial progenitor cells are essential for tumor lymphangiogenesis and lymphatic metastasis. *Cancer Res* 2004;64:3737-40.
- [44] JAIN RK, DUDA DG.. Role of bone marrow-derived cells in tumor angiogenesis and treatment. *Cancer Cell* 2003;3:515-6.
- [45] DAVIDOFF AM, NG CY, BROWN P, LEARY MA, SPURBECK WW, ZHOU J, *et al*. Bone marrow-derived cells contribute to tumor neovasculature and, when modified to express an angiogenesis inhibitor, can restrict tumor growth in mice. *Clin Cancer Res* 2001; 7:2870-2879.
- [46] STOLL BR, MIGLIORINI C, KADAMBI A, MUNN LL, JAIN RK. A mathematical model of the contribution of endothelial progenitor cells to angiogenesis in tumors: implications for antiangiogenic therapy. *Blood* 2003; 102: 2555- 2561.
- [47] RUZINOVA MB, SCHÖER RA, GERALD W, EGAN JE, PANDOLFI PP, RAFII S, *et al*. Effect of angiogenesis inhibition by Id loss and the contribution of bone-marrow-derived endothelial cells in spontaneous murine tumors. *Cancer Cell* 2003; 4:277-289.
- [48] PETERS BA, DIAZ LA, POLYAK K, MESZLER L, ROMANS K, GUINAN EC, *et al*. Contribution of bone marrow-derived endothelial cells to human tumor vasculature. *Nat Med* 2005;20:261-2.
- [49] SUN B, ZHANG S, NI C, ZHANG D, LIU Y, ZHANG W, *et al*. Correlation between melanoma angiogenesis and the mesenchymal stem cells and endothelial progenitor cells derived from bone marrow. *Stem Cells Dev* 2005;14:292-8.
- [50] OKAMOTO R, UENO M, YAMADA Y, TAKAHASHI N, SANO H, SUDA T, *et al*. Hematopoietic cells regulate the angiogenic switch during tumorigenesis. *Blood* 2005;105:2757-63.
- [51] RAJANTIE I, ILMONEN M, ALMINAITE A, OZERDEM U, ALITALO K, SALVEN P. Adult bone marrow-derived cells recruited during angiogenesis comprise precursors for periendothelial vascular mural cells. *Blood* 2004;104:2084-6.
- [52] REYES M, DUDEK A, JAHAGIRDAR B, KOODIE L, MARKER PH, VERFAILLIE CM. Origin of endothelial progenitors in human postnatal bone marrow. *J Clin Invest* 2002;109:337-46.
- [53] DE PALMA M, VENNERI MA, ROCA C, NALDINI L. Targeting exogenous genes to tumor angiogenesis by transplantation of genetically modified hematopoietic stem cells. *Nat Med* 2003;9:789-95.
- [54] DWENGER A, ROSENTHAL F, MACHEIN M, WALLER C, Spyridonidis A. Transplanted bone marrow cells preferentially home to the vessels of in situ generated murine tumors rather than of normal organs. *Stem Cells* 2004;22:86-92.
- [55] LYDEN D, HATTORI K, DIAS S, COSTA C, BLAIKIE P, BUTROS L, *et al*. Impaired recruitment of bone-marrow-derived endothelial and hematopoietic precursor cells blocks tumor angiogenesis and growth. *Nat Med* 2001; 7:1194-1201.
- [56] DAVIDOFF AM, NG CY, BROWN P, LEARY MA, SPURBECK WW, ZHOU J, *et al*. Bone marrow-derived cells contribute to tumor neovasculature and, when modified to express an angiogenesis inhibitor, can restrict tumor growth in mice. *Clin Cancer Res* 2001;7:2870-9.
- [57] GRUNEWALD M, AVRAHAM I, DOR Y, BACHAR-LUSTIG E, ITIN A, YUNG S, *et al*. VEGF-induced adult neovascularization: recruitment, retention, and role of accessory cells. *Cell* 2006;124:175-89.
- [58] von MARSCHALL Z, CRAMER T, HOCKER M, FINKENZELLER G, WIEDENMANN B, ROSEWICZ S. Dual mechanism of vascular endothelial growth factor upregulation by hypoxia in human hepatocellular carcinoma. *Gut* 2001;48:87-96.
- [59] ANNABI B, LEE YT, TURCOTTE S, NAUD E, DESROSIERS RR, CHAMPAGNE M, *et al*. Hypoxia promotes murine

- bone-marrow-derived stromal cell migration and tube formation. *Stem Cells* 2003;21:337-47.
- [60] CERADINI DJ, GURTNER GC. Homing to hypoxia: HIF-1 as a mediator of progenitor cell recruitment to injured tissue. *Trends Cardiovasc Med* 2005;15:57-63.
- [61] SONG S, EWALD AJ, STALLCUP W, WERB Z, BERGERS G. PDGFRbeta+ perivascular progenitor cells in tumors regulate pericyte differentiation and vascular survival. *Nat Cell Biol* 2005;7:870-9.
- [62] ANNABI B, NAUD E, LEE YT, ELIOPoulos N, GALIPEAU J. Vascular progenitors derived from murine bone marrow stromal cells are regulated by fibroblast growth factor and are avidly recruited by vascularizing tumors. *J Cell Biochem* 2004;91:1146-58.
- [63] NATORI T, SATA M, WASHIDA M, HIRATA Y, NAGAI R, MAKUCHI M. G-CSF stimulates angiogenesis and promotes tumor growth: potential contribution of bone marrow-derived endothelial progenitor cells. *Biochem Biophys Res Commun* 2002;297:1058-61.
- [64] RUZINOVA MB, SCHOER RA, GERALD W, EGAN JE, PANDOLFI PP, RAFII S, et al. Effect of angiogenesis inhibition by Id loss and the contribution of bone-marrow-derived endothelial cells in spontaneous murine tumors. *Cancer Cell* 2003;4:277-89.
- [65] FERRARA N, KERBEL RS. Angiogenesis as a therapeutic target. *Nature* 2005;438: 967-74.
- [66] CARMELIET P. Angiogenesis in life, disease and medicine. *Nature* 2005;438:932-6.
- [67] DE PALMA M, VENNERI MA, ROCA C, NALDINI L. Targeting exogenous genes to tumor angiogenesis by transplantation of genetically modified hematopoietic stem cells. *Nat Med* 2003;9:789-95.
- [68] VAN DAMME A, VANDEN DRIESSCHE T, COLLEN D, CHUAH MK. Bone marrow stromal cells as targets for gene therapy. *Curr Gene Ther* 2002;2:195-209.
- [69] MOSCA JD, HENDRICKS JK, BUYANER D, DAVIS-SPROUL J, CHUANG LC, MAJUMDAR MK, et al. Mesenchymal stem cells as vehicles for gene delivery. *Clin Orthop Relat Res* 2000;379:S71-90.
- [70] SCHULER T, KORNIG S, BLANKENSTEIN T. Tumor rejection by modulation of tumor stromal fibroblasts. *J Exp Med* 2003;198:1487-93.
- [71] SPIOTTO MT, YU P, ROWLEY DA, NISHIMURA MI, MEREDITH SC, GAJEWSKI TF, et al. Increasing tumor antigen expression overcomes "ignorance" to solid tumors via crosspresentation by bone marrow-derived stromal cells. *Immunity* 2002;17:737-47.
- [72] FU P, CHEN J, TIAN Y, WATKINS T, CUI X, ZHAO B. Anti-tumor effect of hematopoietic cells carrying the gene of ribonuclease inhibitor. *Cancer Gene Ther* 2005;12: 268-75.
- [73] JAIN RK, DUDA DG.. Role of bone marrow-derived cells in tumor angiogenesis and treatment. *Cancer Cell* 2003;3:515-6.
- [74] KAPLAN RN, RIBA RD, ZACHAROULIS S, BRAMLEY AH, VINCENT L, COSTA C, et al. VEGFR1-positive hematopoietic bone marrow progenitors initiate the pre-metastatic niche. *Nature* 2005;438:820- 7.
- [75] VAN VALCKENBORGH E, BAKKUS M, MUNAUT C, NOEL A, ST PIERRE Y, ASOSINGH K, et al. Upregulation of matrix metalloproteinase-9 in murine 5T33 multiple myeloma cells by interaction with bone marrow endothelial cells. *Int J Cancer* 2002;101:512-8.
- [76] ACUFF HB, CARTER KJ, FINGLETON B, GORDEN DL, MATRISIAN LM. Matrix metalloproteinase-9 from bone marrow-derived cells contributes to survival but not growth of tumor cells in the lung microenvironment. *Cancer Res* 2006;66: 259-66.
- [77] FIERRO FA, SIERRALTA WD, EPUNAN MJ, MINGUELL JJ. Marrow-derived mesenchymal stem cells: role in epithelial tumor cell determination. *Clin Exp Metastasis* 2004;21:313-9.
- [78] HOMBAUER H, MINGUELL JJ. Selective interactions between epithelial tumour cells and bone marrow mesenchymal stem cells. *Br J Cancer* 2000;82:1290-6.
- [79] PAWELEK JM. Tumour cell hybridization and metastasis revisited. *Melanoma Res* 2000;10: 507-14.
- [80] RACHKOVSKY M, SODI S, CHAKRABORTY A, AVIS-SAR Y, BOLOGNIA J, MCNIFF JM, et al.. Melanoma x macrophage hybrids with enhanced metastatic potential. *Clin Exp Metastasis* 1998;16: 299-312.
- [81] DUELLI D, LAZEBNIK Y. Cell fusion: a hidden enemy? *Cancer Cell* 2003;3:445-8.
- [82] A RIZVI AZ, SWAIN JR, DAVIES PS, BAILEY AS, DECKER AD, WILLENBRING H, et al. Bone marrow-derived cells fuse with normal and transformed intestinal stem cells. *Proc Natl Acad Sci U S A* 2006;103:6321-5.
- [83] KERBEL RS, LAGARDE AE, DENNIS JW, DONAGHUE TP. Spontaneous fusion in vivo between normal host and tumor cells: possible contribution to tumor progression and metastasis studied with a lectin-resistant mutant tumor. *Mol Cell Biol* 1983;3:523-38.