GENOTYPING OF URUGUAYAN HUMAN ADENOVIRUS ISOLATES COLLECTED BETWEEN 1994 AND 1998

S. FRABASILE¹, N. VITUREIRA¹, G. PEREZ¹, S. MATEOS², J. ARBIZA^{1*}

¹Sección Virología, Facultad de Ciencias, Universidad de la República, Iguá 4225., Montevideo, Uruguay; ²Departamento de Bacteriologia y Virologia, Instituto de Higiene, Facultad de Medicina, Montevideo, Uruguay

Received October 8, 2004; accepted April 18, 2005

Summary. – Adenoviruses are one of the most frequent causative agents of acute lower respiratory infections in infants and young children. Twenty-three adenovirus isolates from nasopharyngeal aspirates of children hospitalized for acute lower respiratory infections in Uruguay between 1994 and 1998 were studied by restriction enzyme analysis. The genomic analysis showed that 60.9% (n = 14) of isolates belonged to the species *Human adenovirus C (HAdV-C)* and 31.9% (n = 9) to the species *Human adenovirus B (HAdV-B)*. Whereas some isolates could be classified according to the published profiles into genotype or genomic variant, others displayed migration patterns not allowing classification. Eight isolates (89%) of *HAdV-B* corresponded to the Ad7h genotype that has been associated with severe and fatal pneumonia and necrotizing bronchiolitis in children in South America. The isolates of *HAdV-C* showed a great variability in accordance with the data published earlier.

Key words: human adenoviruses; genotyping; restriction analysis

Introduction

Human adenoviruses (the *Mastadenovirus* genus, the *Adenoviridae* family) (van Regenmortel *et al.*, 2000) belong to the most frequent causative agents of acute lower respiratory infections in infants and young children (Horwitz, 2001; Brandt *et al.*, 1969). In developing countries, adenoviruses cause serious illness and death in childhood (Pio *et al.*, 1984; Berman, 1991). Among them, those classified within the species *HAdV-B* (serotypes or simply types 3 and 7), *HAdV-C* (types 1, 2 and 5) and *HAdV-E* (type 4) are regarded

as the cause of respiratory diseases (Sharp and Wadell, 1995). In South American countries, adenoviruses were found to be the second major group of viruses after Respiratory syncytial virus which infect the respiratory tract of children (Suarez *et al.*, 1988a,b; Avila *et al.*, 1989, 1990; Nascimento *et al.*, 1991; Kajón *et al.*, 1996).

Restriction analysis performed on 23 adenovirus isolates from Argentina, Chile and Uruguay between 1991 and 1994 revealed that, whereas a majority of them belonged to the *HAdV-B* species, the remaining ones were classified mostly as *HAdV-C* and rarely as *HAdV-E* (Kajón *et al.*, 1996).

In this report we present the results of genotyping by restriction analysis of 23 adenovirus isolates originating from children with acute lower respiratory infections from Uruguay between 1994 and 1998.

Materials and Methods

Virus isolation was performed on HEp-2 cells from nasopharyngeal aspirates collected from children under five years with acute lower respiratory diseases between 1994 and 1998 in Uru-

^{*}Corresponding author. E-mail: jarbiza@fcien.edu.uy; fax: +5982-5258617.

Present address: Laboratório de Vírus Respiratórios, Pavilhão Cardoso Fontes, Instituto Oswaldo Cruz (FIOCRUZ/CNPq), Avenida Brasil 4365, CEP: 21.045-900, Manguinhos, Rio de Janeiro, Brazil.

Abbreviations: CPE = cytopathic affect; IFA = immunofluorescence assay; $HAdV-B = Human \ adenovirus \ B; \ HAdV-C$ = Human adenovirus C; $HAdV-E = Human \ adenovirus \ E$

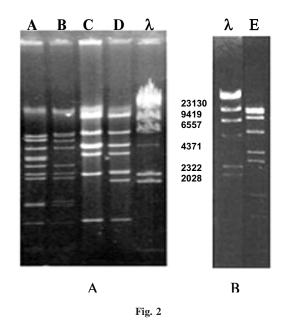
guay. The cells were grown in Eagle's Minimal Essential Medium containing 10% of fetal bovine serum, 30 mg/ml glutamine, 100 μ l peniciline and 50 μ l gentamicine. Upon appearance of cytopathic effect (CPE), the cultures were frozen.

Immunofluorescence assay (IFA). Identification of adenovirus infection was performed by an indirect IFA (Ballew *et al.*, 1984) on infected culture cells showing CPE using commercial monoclonal antibodies (Chemicon International, Inc.) conjugated with fluorescein.

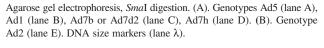
Viral DNA extraction. Each viral isolate was inoculated onto 75–80% confluent monolayers of HEp-2 cells in 75 cm² flasks. Upon appearance of an extensive CPE, the cells were harvested and the DNA was extracted according to Shinagawa *et al.* (1983) with modifications.

Restriction analysis of viral DNA and agarose gel electrophoresis. One μ g of viral DNA extracted from each sample was digested with 10 U of *Bam*HI, *Sma*I and *Bg*/II under conditions specified by the manufacturer (Promega). The digested viral DNAs were loaded onto 0.7% agarose gel containing 0.25 µg/ml ethidium bromide and run at 80 V for 3 hrs in TBE buffer pH 8.0. Restriction profiles were visualized under UV light and photographed. The resulting restriction profiles were assigned to corresponding virus species and genotypes according to the literature data.

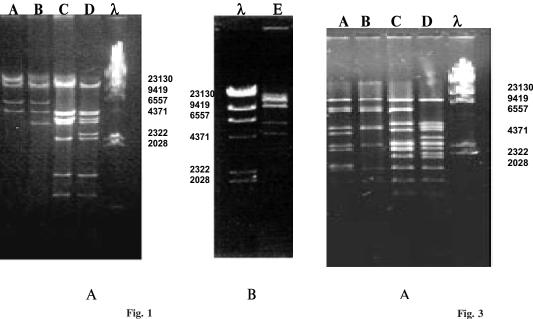


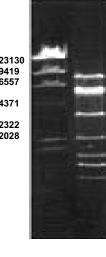


Restriction profiles of standard genotypes of some human adenoviruses









В

F

Restriction profiles of standard genotypes of some human adenoviruses

Agarose gel electrophoresis, *Bam*HI digestion. (A). Genotypes Ad5 (lane A), Ad1 (lane B), Ad7b or Ad7d2 (lane C), Ad7h (lane D). (B). Genotype Ad2 (lane E). DNA size markers (lane λ).

Restriction profiles of standard genotypes of some human

adenoviruses

Agarose gel electrophoresis, *BgI*II digestion. (A). Genotypes Ad5 (lane A), Ad1 (lane B), Ad7b or Ad7d2 (lane C), Ad7h (lane D). (B). Genotype Ad2 (lane E). DNA size markers (lane λ).

BamHI

 Table 1. Genotyping of human adenovirus isolates

Species	Genotype of genome variant	No. of isolates	Years of circulation
HAdV-B			
n = 9 (31.9%)	Ad7h	8	94, 95, 96, 97, 98
	Ad7b or Ad7 d21	1	98
HAdV-C			
n = 14 (60.8%)	Ad1D1 ² , AV1 New I	I ³ 4	95, 96, 97, 98
	AV2 New IV4, Ad2p	1 ⁵ 3	96
	AV2 New IV ⁶	6	97, 98
	AV5 New V	1	98

¹Based on *Bgl*II profile.

²Based on *Bam*HI profile.

³Based on *Sma*I and *Bgl*II profiles.

⁴Based on *Bgl*II profile.

⁵Based on *Sma*I profile. ⁶Based on *Bam*HI and *Bgl*II profiles.

Based on *Bam*FI and *Bgt*II promes.

Results and Discussion

A total of 23 epidemiologically unrelated adenovirus isolates from nasopharyngeal aspirates from children under 5 years of age hospitalized with acute lower respiratory disease between 1994 and 1998 in Uruguay were genotyped by digestion of viral DNAs with *Bam*HI, *Sma*I and *BgI*II. The resulting restriction profiles were assigned to corresponding genotypes according to the literature data.

The restriction analysis showed that 60.8% of the analyzed isolates belonged to *HAdV-C* and 39.1% to *HAdV-B* (Table 1). These results were in accordance with an earlier report from Brazil (Kajón *et al.*, 1999), where the incidence of *HAdV-C* was 64.1% and that of *HAdV-B* was 35.8%, but in contrast to other reports from South America (Kajón *et al.*, 1996; Videla *et al.*, 1999; Larrañaga *et al.*, 2000). However, no conclusion can be drawn due to the low number of isolates analyzed.

Of the nine isolates (39.1%) identified as *HAdV-B* eight corresponded clearly to the genotype Ad7h (Figs. 1–3, lanes D) (Hashido *et al.*, 1999; Kajón *et al.*, 1996; Videla *et al.*, 1999, Erdman *et al.*, 2002).

Single isolate displayed *Bam*HI and *Sma*I restriction profiles with a single fragment of about 2.322 bp instead of two expected for the genotype Ad7h (Figs. 1 and 2, lanes C) but a *Bgl*II restriction profile (Fig. 3, lane C) fully matching the genotype Ad7b or Ad7d2 (Li and Wadell, 1988; Hashido *et al.*, 1999; Erdman *et al.*, 2002). Ad7h turned out to be the genotype associated with severe fatal pneumonia and necrotizing bronchiolitis in small children in South America (Kajón and Wadell, 1992; Kajón *et al.*, 1994, 1996; Larrañaga *et al.*, 2000). In this study, 88.8% of the isolates belonging to *HAdV-B* were identified as the Ad7h genotype.

Of the fourteen isolates belonging to *HAdV-C* one was genotyped as Ad5 New V genomic variant (Figs. 1–3, lanes

A) using all the three restriction enzymes. The other thirteen isolates showed a marked variability. Four isolates were genotyped as Ad1D1 (Fig. 1, lane B) based on *Bam*HI profile (Kajón *et al.*, 1996; Videla *et al.*, 1999) but as AV1 New II genomic variant according to *Sma*I and *Bg*III profiles (Figs. 2 and 3, lanes B).

Three isolates were assigned to AV2 New IV genomic variant (Kajón *et al.*, 1996) based on *Bg*III profile but to Ad2p1 according to *Sma*I profile (Li *et al.*, 1999). The *Bam*HI digestion of DNAs of these isolates could not be evaluated due to obscure profiles.

Six isolates were genotyped as AV2 New IV genomic variant according to *Bgl*II and *Bam*HI profiles (Figs. 1–3, lanes E). However their *Sma*I profiles did not match the expected ones (Kajón *et al.*, 1996) (Fig. 2, lane E).

The higher genome variability of the isolates of *HAdV-C* compared to those of *HAdV-B* was in accord with the data of other authors (Avila *et al.*, 1990; Nascimento *et al.*, 1991). Several variants were found among the fourteen isolates belonging to *HAdV-C*. Thus, of fourteen Uruguayan isolates of *HAdV-C* thirteen did not match the profiles reported earlier for this virus species (Table 1). On the other hand, of nine Uruguayan isolates of *HAdV-B* only one did not match the expected profiles.

The data obtained in this study show the presence of human adenoviruses of species B (HAdV-B) and C (HAdV-C) isolated from epidemics between 1994 and 1998 in Uruguay. Although just a low number of serum samples as well as virus isolates (n = 23) was analyzed, genetic variability of human adenoviruses reflected in different species, genotypes and genotype variants has been demonstrated.

Acknowledgements. The authors thank Dr. A. Kajón, Lovelace Respiratory Research Institute, Albuquerque, NM, USA for critical reading of the manuscript, Dr. H. Chiparelli, and Mr. R. Camera from Departamento de Laboratorios de Salud Publica for respiratory virus diagnosis and technical assistance, respectively, Drs. C. Pirez and S. Dalmas, Clinica Pediatrica B of Hospital Pereira Rossell, Montevideo, Uruguay for collecting serum samples. This research was funded in part by a grant from CSIC-Universidad de la República.

References

- Avila M, Carballal G, Rovaletti H, Ebekian B, Cusminsky M, Weissenbacher M (1989): Viral etiology in acute lower respiratory infections in children from community. *Am. Rev. Respir. Dis.* 140, 634–637.
- Avila M, Salomón H, Carballal G, Ebekian B, Woyskovksy N, Cerqueiro MC, Weissenbacher M (1990): Isolation and identification of viral agents in Argentinian children with acute lower respiratory tract infection. *Rev. Infect. Dis.* 12, 974–981.

- Ballew H, Lyerla HC, Forrestier FT (1984): *Laboratory Methods* for Diagnosing Respiratory Infections. Atlanta, GA: US Department of Health and Human Services, Public Health Services.
- Berman S (1991): Epidemiology of acute respiratory infections in children in developing countries. *Rev. Infect. Dis.* 13, 454–462.
- Brandt CD, Kim HN, Vargosdo AJ (1969): Infection in 18000 infants and children in a controlled study of respiratory tract disease. I: Adenovirus pathogenicity in relation to serologic type and illness syndrome. *Am. J. Epidemiol.* **90**, 484–500.
- Erdman DD, Xu W, Gerber SI, Gray GC, Schnurr D, Kajon AE and Anderson L (2002): Molecular Epidemiology of Adenovirus Type 7 in the United States, 1966–2000. *Emerg. Infect. Dis.* 8, 269–277.
- Hashido M, Mukouyama A, Sakae K, Tsuzuki H, Yamashita T, Inada T, Inouye S (1999): Molecular and serological characterization of adenovirus genome type 7h in Japan. *Epidemiol. Infect.* **122**, 281–286.
- Horwitz MS (2001): Adenoviruses. In Knipe DM, Howley PM (Eds): *Fields Virology*. Fourth Edition. Lippincott-Williams and Wilkins, Philadelphia, pp. 2301–2326.
- Kajón A, Portes S, de Mello W, Nacimento J, Siqueira M (1999): Genome type analysis of Brazilian adenovirus strains of serotypes 1,2,3,5, and 7 collected between 1976 and 1995. *J. Med. Virol.* 58, 408–412.
- Kajón A, Mistchenko A, Videla C, Hortal M, Wadell G, Avendańo LF (1996): Molecular epidemiology of adenovirus acute lower respiratory infections of children in the South Cone of South America (1991–1994). J. Med. Virol. 48, 151–156.
- Kajón A, Wadell G (1992): Characterization of adenovirus genome type 7h: analysis of its relationship to other members of serotype 7. *Intervirology* 33, 86–90.
- Kajón A, Larrañaga C, Suárez M, Wadell G (1994): Genome type analysis of Chilean adenovirus strains isolated in a children's hospital between 1998 and 1990. J. Med. Virol. 42, 16–21.
- Kajón A, Suarez V, Avendańo L, Hortal M, Wadell, G (1993): Genome type analysis of South American adenoviruses of subgenus C collected over a 7-year period. *Arch. Virol.* 132, 29–35.
- Larrañaga C, Kajon A, Villagra E, Avendańo L (2000): Adenovirus surveillance of children hospitalized for acute lower

respiratory infection in Chile (1988–1996). *J. Med. Virol.* **60**, 342–346.

- Li QG, Wadell G (1988): Comparison of 17 genome types of adenovirus type 3 identified among strains recovered from six continents. J. Clin. Microbiol. 26, 1009–1015.
- Li QG, Henningsson A, Juto P, Elgh F, Wadell G (1999): Use of restriction fragment analysis and sequencing of a serotype-specific region to type adenovirus isolates. *J. Clin. Microbiol.* **37**, 844–847.
- Nascimento JP, Siqueira M, Sutmoller F, Krawczuck MM, Farias V, Ferreira V, Rodriguez MJ (1991): Longitudinal study of acute respiratory diseases in Rio de Janeiro: Occurrence of respiratory viruses during four consecutive years. *Rev. Inst. Med. Trop. de Sao Paulo* 33, 287–296.
- Pio A, Leowski J, TenDam HG (1984): The magnitude of the problem of acute respiratory infections. In "Acute Respiratory Infection in Childhood". *Proceeding of an International Workshop*, Sydney, August, 1984, pp. 3–16.
- Sharp IR, Wadell G (1995): Adenoviruses. In Zukerman AJ, Banatvalla J, Pattison JR (Eds): *Principles and Practices* of Clinical Virology. 3er ed. John Wiley, New York, pp. 287–308.
- Shinagawa M, Marsuda A, Ishiyana T, Goto H, Soto G (1983): A rapid and single method for preparation of adenovirus DNA from infected cells. *Microbiol. Immunol.* 27, 817–822.
- Suarez M, Wu E, Carrasco L, Acevedo C, Ramirez R, Peńa A, Larrañaga C, Morales T (1988a): Viral participation in the lower acute respiratory infections of infant. *Revista Chilena de Pediatría* **59**, 353–357 (in Spanish).
- Suarez M, Wu E, Carrasco L, Torrijos J, Massu M, Vildoso J, Cantos A, Hanke, M T (1988b): Viral detection in acute respiratory infections in hospitalized children. Serologic study. *Enfermedad Respiratoria y Cirugía Torácica* 4, 10–14 (in Spanish).
- van Regenmortel MHV, Fauquet CM, Bishop (2000): Virus Taxonomy: The Classification and Nomenclature of Viruses. The Seventh Report of the International Committee on Taxonomy of Viruses. Academic Press, San Diego-San Francisco-New York-Boston-London-Sydney-Tokyo.
- Videla C, Carballal G, Kajon A (1999): Genomic analysis of adenovirus isolated from Argentinian children with acute lower respiratory infections. J. Clin. Virol. 14, 67–71.