

EXPERIMENTAL STUDY

Lawsonia inermis – An alternative treatment for hyperthyroidism?Zumrutdal E¹, Karateke F¹, Daglioglu K², Gulkaya M¹, Colak O³, Koksal F⁴

Numune Training and Research Hospital, Department of General Surgery, Adana, Turkey.

ezumrutdal@yahoo.com

Abstract: *Aim:* The goal of our study was to determine the effects of *Lawsonia inermis* (*L. inermis*) in mice, in which hyperthyroidism had been caused by thyroid stimulant hormone (TSH).

Material and method: The first phase of the study aimed to detect the effects of *L. inermis* on the amount of ionized hydrogen (pH) in cells. For this aim, the effect of *L. inermis* on pH levels in the liver tissues of mice, in whom *Escherichia coli* (*E. coli*) had caused peritonitis, was examined. In the second phase of the study, the effect of *L. inermis* on the serum T4 levels in the 24th and 48th hour in mice, whose thyroid cells showed an increased activity by TSH was measured.

Results: In the first phase, in mice, in whom *E. coli* had caused peritonitis, the pH in the liver tissue of the group that had been given *L. inermis* was found to be significantly alkaline ($p < 0.05$). In the second phase, in mice, in whom TSH had caused hyperthyroidism, it was noted that serum total T4 levels were significantly lower than in the group that had been given *L. inermis* in the 48th hour ($p < 0.05$).

Conclusion: In our study, we detected that *L. inermis* significantly decreased serum total T4 levels in the 48th hour in mice in whom TSH had caused hyperthyroidism. These results suggest that *L. inermis* can be used as an alternative treatment for the Graves' disease (Tab. 2, Fig. 1, Ref. 34). Text in PDF www.elis.sk.

Key words: *Lawsonia inermis*, hyperthyroidism.

L. inermis (henna leaf) is a popular natural dye to colour hands and hair in eastern cultures. *L. inermis* contains 2 OH 1.4 naphthoquinone's that has oxidant effects (1–3). It is also known for its benefits for medical treatment. In many studies, the antimicrobial, antitumoral and tuberculostatic effects of *L. inermis* were investigated (4–8). Graves' Disease is an autoimmune disease with hyperthyroidism. In this disease, thyroid stimulant immunoglobulin (TSI) attaches itself to thyroid stimulant hormone (TSH) and creates the thyroid hormone synthesis that is much more powerful than TSH. A need arises for more hydrogen ions (H⁺) for thyroid hormone synthesis in thyroid cells exposed to the disease in this way. The objective of this study was to determine whether *L. inermis*'s oxidant effects would decrease the thyroid hormone synthesis by decreasing the hydrogen ions in the hyperactive thyroid cells. According to our knowledge, this is the first study in English literature up to date.

Methods

H⁺ is important both for myeloperoxidase (MPO) and for thyroidperoxidase (TPO) enzymes. Oxidant molecules created by the neutrophils against microbial agents play a critical role in the host defense mechanism. The active phagocytes secrete the enzyme MPO and produce hydrogen peroxide. In the formation of

¹Numune Training and Research Hospital, Department of General Surgery, Adana, Turkey, ²Cukurova University, Faculty of Veterinary Adana, Turkey, ³Cukurova University, Department of Biology, Adana, Turkey, and ⁴Cukurova University, Department of Microbiology, Adana, Turkey

Address for correspondence: E. Zumrutdal, MD, Adana Numune Training and Research Hospital Department of General Surgery, Adana, Turkey

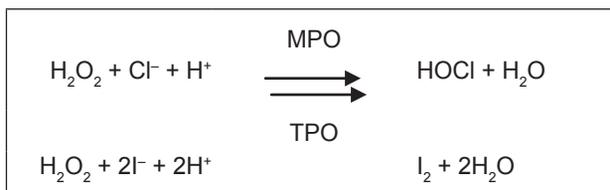


Fig. 1. The pathways of MPO and TPO enzymes.

thyroid enzymes, the enzyme TPO causes a reaction resembling that of the enzyme MPO.

Both MPO and TPO show the substrate and product similarities during the reactions (Fig. 1). TPO allows the formation of iodine (I₂) using H₂O₂ with iodide (I⁻). Iodine is given to tyrosine residues in the form of iodine and thyroid hormone synthesis starts. For this reason, the ion concentration is very important in thyroid hormone synthesis. A need arises for more H⁺ for thyroid hormone synthesis in thyroid cells exposed to Grave's disease.

It is known that the best indicator of H⁺ concentration is pH measurement. The thyroid glands in mice are so small, it was thought that the pH measurement in this tissue would not be trustworthy, so the study was planned in two phases.

In the first phase of the study, in order to better understand the effects of *L. inermis* on TPO and H⁺ concentrate in the hyperactive thyroid cells, we wanted to make use of the MPO enzyme, which is very similar in terms of substrate, product, and type of function to TPO. Yao et al showed that in the case of infection, the enzyme MPO became active in the liver in the 24th hour (9, 10). We created peritonitis in mice by administering *E. Coli* intraperitoneally to examine the effect of *L. inermis* on pH levels in the liver tissue

in the 24th hour. In the second phase of the study, we investigated the effect of *L. inermis* on the serum total T4 hormone level in the 24th and 48th hours in mice, in which hyperthyroidism had been caused by administering TSH intraperitoneally.

Subjects

Sixty female Swiss albino mice (25–30 g) were used in the first phase and 60 mice were used in the second phase. In order to ensure a natural environment before the experiment, the mice were kept in seclusion for 5 days at the same room temperature with 12 hours each of light and darkness every day. Throughout the entire duration of the observation and experiment, all the mice received the same food and water. Animal ethics approval for the entire project was obtained from the Medical Experimental Research Center of the University of Cukurova Animal Ethics Committee.

Creation of L. inermis solution

The *L. inermis* solution (2 %) was made by mixing the freeze-dried form of this plant (henna powder) with distilled water. The solution was heated to boiling using a magnetic stirrer, and was allowed to cool to room temperature and the sediment was removed by straining (8).

Formation of E. coli peritonitis in first phase

The *E. coli* culture, which had been formed in a Luria Bertani medium at 37 °C, was washed twice with sterile saline before injection. 108 CFU of *E. coli* was injected intraperitoneally inside of 200 µl of sterile saline. Following the inoculation of *E. coli*, its viability was checked with an emergency blood agar plate (11–15).

Preparation of liver tissue samples and measurement of pH

Mice were euthanized by cervical dislocation in the 24th hour following *E. coli* inoculation. A midline laparotomy was performed and approximately 1g of tissue from the right liver lobe was placed in a tube containing 1.5 ml 0.02 M EDTA and homogenized in a container of ice (Ultra Turrax T25). The pH of the homogenized liver tissue was assessed with a pH meter (InoLab) after centrifuging at 4 °C.

Work Plan

In the first phase, 60 mice were divided to 3 groups (n=20). In the 1st group, *E. coli* was inoculated intraperitoneally and 400 mg/kg/day of *L. inermis* orally by gavage was given in the 1st, 11th, and 22nd hours. In the 2nd group, *E. coli* was inoculated intraperitoneally and SF was given orally by gavage in the 1st, 11th and 22nd hours. In the 3rd group (control group), after SF administration intraperitoneally, SF was given orally by gavage in the 1st, 11th and 22nd hours. The pH was measured in 24th hour in liver for all groups.

In the second phase, 60 mice was divided to 6 groups (n=10). In the 1st group, TSH was given intraperitoneally and SF was given orally by gavage in the 1st, 11th and 22nd hours. In the 2nd group, TSH was given intraperitoneally and 400 mg/kg/day of *L. inermis* orally by gavage in the 1st, 11th and 22nd hours. The T4

levels of both these groups were measured in the 24th hour. In the 3rd group, TSH was given intraperitoneally and SF was given orally by gavage in the 1st, 11th and 22nd hours for two days. In the 4th group, TSH was given intraperitoneally and 400 mg/kg/day of *L. inermis* was given orally by gavage in the 1st, 11th and 22nd hours for two days. In the 5th group after SF administration intraperitoneally, SF was given orally by gavage in the 1st, 11th and 22nd hours for two days. In the 6th group after SF administration intraperitoneally, 400 mg/kg/day of *L. inermis* was given orally by gavage in the 1st, 11th and 22nd hours for two days. The T4 levels of groups 3, 4, 5, 6 were measured in the 48th hour.

The creation of hyperthyroidism

We caused hyperthyroidism in mice by administering TSH intraperitoneally using the method Chiu et al had described (16–18). To the groups 1 and 2, a single dose of 0.3 units (in 150 µl of sterile saline per dose), to the groups 3 and 4, two doses of bovine TSH 24 hours apart were administered intraperitoneally. The groups 5 and 6, which were not given TSH, were administered 150 µl of sterile saline in order to produce the same stress. The bovine TSH was purchased from Sigma (Germany).

Serum total T4 level measurement

After TSH application, following a chest wall incision under anesthesia (Xlazine 10 mg/kg + Ketamine 200 mg/kg), 1 cc of blood was obtained from the right atrium of each mice at 24 hours in the groups 1 and 2 and at 48 hours in the groups 3,4,5 and 6. Serum samples were obtained by centrifuging blood samples for 10 minutes at 5000 rpm. Serum total T4 hormone levels were detected by microelisa at 450 nm. The T4 kit was purchased from Alpha Diagnostics (adjusted test for mouse-serum T4 level measurement, Cat. No. 1100, USA). Results were determined as µg/dl by the standard curve, which was calculated from the samples.

Statistical analysis

Statistical analyses were performed using the statistical package SPSS v 12.0. For each continuous variable, normality was checked. The Student t-test was used for normally distributed data (pH) and the Mann–Whitney U test was used for non-normally distributed data (T4). p value <0.05 was considered as statistically significant.

Results

Phase 1 results

The liver tissue pH level of the mice in the group 1 were statistically significantly higher than in the group 2 (p<0.05). The pH levels of all groups in the Phase I study are shown in Table 1.

Phase 2 results

In the Phase 2, the mean serum total T4 level in the 24th hour in the group 1 was 7.91 µg/dL, while the T4 level in the group 2 was found to be 7.67 µg/dL (p>0.05). In the group 3, the mean serum total T4 level in the 48th hour was found to be 6.39 µg/dL, while it was found to be 5.00 µg/dL in the group 4 (p<0.05). Group

Tab. 1. pH values of liver tissue samples.

	Mean pH± SD	Median pH (Min–Max)
Group-1 Inoculated E.Coli and L.inermis	6.58±0.10*	6.59 (6.45–6.77)
Group-2 Inoculated E.Coli and water	6.53±0.11	6.53 (6.29–6.71)
Group-3 Control	6.52±0.08	6.52 (6.38–6.62)

Tab. 2. Serum total T4 levels (ug/dL) in mice with hyperthyroidism after getting L. Inermis or placebo.

Groups	24 hr T4 level	48 hr T4 level	p value
1 IP TSH + Water	7.91±1.93		=0.570
2 IP TSH + L. inermis	7.67±1.67		
3 IP TSH + Water		6.39±1.04	=0.041
4 IP TSH + L. inermis		5.00±1.59	
5 IP Saline + Water		4.03±0.85	=0.820
6 IP Saline + L. inermis		3.92±0.49	

IP – intraperitoneal

5 had a mean T4 level of 4.03 ug/dL, while the group 6 had T4 level of 3.92 ug/dL ($p>0.05$) (Tab. 2).

Discussion

Diseases associated with hyperthyroidism can occur with different mechanisms and its treatment is performed with different methods that are conservative and/or surgical. Hyperthyroidism model created in our study is an adaptation of the model performed by Chiu et al on TSH receptors (18). In the model of this study, the TSH administered intraperitoneally increases the production of the T4 hormone by connecting to the TSH receptors in the thyroid follicle cells.

It was reported by various researchers that *L. inermis* created serious hemolysis with the oxidant properties in children who suffer deficiency of the enzyme G6PD in the order of usages. G6PD is one of the basic sources that provide H⁺ to the cell with the help of NADPH+H⁺. It was shown that a minor oxidant effect related to *L. inermis* in a child who is G6PD deficient could lead to a serious illness.

In our study, we measured the effect of *L. inermis* on serum T4 levels in mice, in whom hyperthyroidism had been caused. This might be related to the oxidant effect of *L. inermis*. For this aim we wanted to measure pH in mice thyroid gland. However, the pH measurement might not be healthy due to the small size of the mice thyroid gland. So we benefited from the liver MPO enzyme, which resembles in both structure and function the thyroid TPO enzyme used in making thyroid hormones. Yao et al showed an increased MPO enzyme activation in liver tissue in the 24th hour in mice that had peritonitis (9, 10). In our study, in mice in whom peritonitis had been induced, the pH values in the liver tissues of mice in the group that had been given *L. inermis* shifted significantly to the alkaline side ($p<0.05$).

In the group of mice with peritonitis, which had not been given *L. inermis*, the pH was not different from that of normal mice in whom peritonitis had not been induced. These results showed that H⁺ levels had decreased with the oxidant effect of the *L. Inermis*.

Austin et al showed that the pH (19) difference in the tissue is related to the changes in the intracellular pH. In this experimental study, we showed that oxidant effect of the *L. inermis* increased pH by decreasing the H ions in hypermetabolic cells.

The use of H⁺ increases in the hyperthyroid tissue that the TPO enzyme activated. In the second phase of our study, in the mice in which hyperthyroidism had been induced with the TSH stimulation, a significant decrease was noted in the serum T4 level in the 48th hour by *L.inermis*. These data, when they were evaluated with the first phase of our study, suggested that the oxidant effect of *L. inermis* decreased the production of T4 in the thyroid cells stimulated with TSH by connecting the hydrogen ions in.

In previous studies, it was shown that intracellular H₂O₂ increased as a response to TSH (20–23) and the increase in intracellular level of H₂O₂ (24–27) induce apoptosis through oxidative stress in the cell. The reduction of H₂O₂ in the cell under stimulation of TSH due to the H⁺ deficiency of both its use and its destruction, the increasing H₂O₂ concentration's killing of the diseased cell via apoptosis may be another mechanism related to the *L. inermis*'s oxidant effect.

There is still no definite information about substrate input and formation of the product during activation of TPO (28–33) in thyroid hormone synthesis. One of the intermediate molecules in the thyroid hormone synthesis is the hypiodous acid (HOI) (34), which shows an effect like the hypochlorous acid (HOCl). Iodine, an electronegative ion found in the 7A group of the periodic table, and which is missing an electron in its outer orbital, tries to become stable by taking on an electron. We can say that with this feature it will stabilize itself in the form of HOI when it cannot create iodine from iodide in thyroid hormone synthesis. By this way, T4 synthesis might be decreased.

In Grave's disease, TSI stimulate the increase in the H₂O₂ level and oxidative stress much more than TSH. With *L. inermis* decreases the reduction of H₂O₂ and by this way the increase of H₂O₂ may cause the cell to start apoptosis. We think that the cells under TSI stimulation might be apoptosized by *L. inermis*.

Conclusion

L. inermis may be effective as an alternative treatment of hyperthroidism and may be a therapeutic approach in Grave's disease by oxidant effects. Further studies are needed at the molecular level.

References

1. Grabley S, Thiericke R. Bioactive agents from natural sources: trends in discovery and application. *Adv Biochem Eng Biotechnol* 1999; 64: 101–154.
2. Konig GM, Wright AD, Sticker O. For new antibacterial sesterpenes from a marine sponge of the genus *Lufferiella*. *J Nat Prod* Feb 1992; 55 (2): 174–178.
3. Rojas A, Hernandez L, Pereda-Miranda R et al. Screening for antimicrobial activity of crude drug extracts and pure natural products from Mexican medicinal plants. *J Ethnopharmacol* 1992; 35 (3): 275–283.

4. **Ali BH, Bashir AK, Tanira MO.** Anti-inflammatory, anti-pyretic and analgesic effects of *Lawsonia inermis* L. (henna) in rats. *Pharmacology* 1995; 51 (6): 356–363.
5. **Habbal OA, AlJabri AA, El-Hug AH et al.** In-vitro antimicrobial activity of *Lawsonia inermis* Linn (henna). A pilot study on the Omani henna. *Saudi Med J* 2005; 26 (1): 69–72.
6. **Singh, VK, Pandey UK.** Fungitoxic studies on bark extract of *Lawsonia inermis* against ringworm fungi. *Hindustan Antibiot Bull* 1989; 31 (1–2): 32–35.
7. **Dasgupta T, Rao RA, Yadava PK.** Modulatory effect of Henna leaf (*Lawsonia inermis*) on drug metabolising phase I and phase II enzymes, lipid peroxidation and chemically induced skin and forestmach papillomagenesis in mice. *Mol Cell Biochem* 2003; 245: 11–22.
8. **Zumrutdal ME, Ozaslan M, Tuzcu M et al.** Effect of *Lawsonia Inermis* treatment on mice with sarcoma. 2008; 7 (16): 278–283.
9. **Yao V, Cooper D, McCauley R et al.** Bacterial translocation in a non-lethal rat model of peritonitis. *Colorectal Dis* 2001; 3: 338–344.
10. **Yao V, McCauley R, Cooper D et al.** Myeloperoxidase response to peritonitis in an experimental model. *ANZ J Surg* 2003; 73: 1052–1056.
11. **Hori Y, Nihei Y, Kurokawa et al.** Accelerated Clearance of *Escherichia coli* in Experimental Peritonitis of Histamine-Deficient Mice. *J Immunol* 2002; 169: 1978–1983.
12. **Knapp S, De Vos AF, Florquin S et al.** Lipopolysaccharide Binding Protein is an Essential component of the innate immune response to *Escherichia coli* peritonitis in mice. *Infection Immunol* 2003; 71 (12): 6747–6753.
13. **Haziot A, Hijiya N, Gangloff SC et al.** Induction of a Novel Mechanism of Accelerated Bacterial Clearance by Lipopolysaccharide in CD14-Deficient and Toll-Like Receptor 4-Deficient Mice. *J Immunol* 2001; 166: 1075–1078.
14. **Sewnath ME, Olszyna DP, Birjmohun R et al.** IL-10-Deficient Mice Demonstrate Multiple Organ Failure and Increased Mortality During *Escherichia Coli* Peritonitis Despite an Accelerated Bacterial Clearance. *J Immunol* 2001; 166: 6323–6331.
15. **Weijer S, Sewnath MH, de Vos, AF et al.** T. Interleukin-18 Facilitates the Early Antimicrobial Host Response to *Escherichia Coli* Peritonitis. *Infection Immun* 2003; 71 (10): 5488–5497.
16. **Sing-Yung WU, Reggio R, Florsheim H.** Characterization of thyrotropin-induced increase in iodothyronine monodeiodinating activity in mice. *Endocrinology* 1985; 116: 901–908.
17. **Gafni M, Gross J.** The effect of elevated doses of thyrotropin on mice thyroid. *Endocrinology* 1975; 97: 1486–1493.
18. **Chiu SC, Kubotak K, Kuzuya N et al.** Effect of prolonged administration of thyrotrophin on serum concentration, release and synthesis of thyroid hormones in mice. *Acta Endocrinol (Copenh)* 1983; 103 (1): 68–75.
19. **Austin C, Wray S.** Extracellular pH signals affect rat vascular tone by rapid transduction into intracellular pH changes. *J Physiol* 1993; 466: 1–8.
20. **Björkman, U.Ekholm R.** Accelerated exocytosis and H₂O₂ generation in isolated thyroid follicles enhance protein iodination *Endocrinology* 1988; 122: 488–494.
21. **Leseney AM, Deme D, Legue O et al.** *Biochimie* 1999; 81: 373–380.
22. **Kimura T, Okajima F, Sho K et al.** *Endocrinology* 1995; 136: 116–123.
23. **Kimura T, Okajima F, Kikuchi T et al.** *Am J Physiol* 1997; 273: E639–E643.
24. **Kim MH, Chung J, Yang JW et al.** Hydrogen peroxide-induced cell death in a human retinal pigment epithelial cell line. *Korean Ophthalmol* 2003; 17 (1): 19–28.
25. **Riou C, Remy C, Rabilloud R et al.** *J Endocrinol* 1998; 156: 315–322.
26. **Riou C, Tonoli H, Bernier-Valentin F et al.** *Endocrinology* 1999; 140: 1990–1997.
27. **Bretz JD, Rymaszewski M, Arscott PL et al.** *J Biol Chem* 1999; 274: 23627–23632.
28. **Magnusson RP, Taurog A, Dorris ML.** Mechanism of iodide-dependent catalytic activity of thyroid peroxidase and lactoperoxidase. *J Biol Chem* 1984; 259: 197–205.
29. **Magnusson RP, Taurog A, Dorris ML.** Mechanism of thyroid peroxidase and lactoperoxidase catalyzed reactions involving iodide. *J Biol Chem* 1984; 259: 13783–13790.
30. **Banerjee RK.** EDTA inhibits peroxidase-catalyzed iodide oxidation through interaction at the iodide binding site. *Biochim Biophys Acta* 1989; 15 (3): 393–396.
31. **Gavaret JM, Cahnmann HJ, Nunez J.** Thyroid hormone synthesis in thyroglobulin. The mechanism of the coupling reaction. *J Biol Chem* 1981; 256: 9167–9173.
32. **Taurog A, Dorris ML, Doerge DR.** Mechanism of simultaneous iodination and coupling catalyzed by thyroid peroxidase. *Arch Biochem Biophys* 1996; 330 (1): 24–32.
33. **Nakamura M, Yamazaki I, Nakagawa H et al.** *J Biol Chem* 1984; 259: 359–364.
34. **Czarnocka B.** *Thyroid International Thyroid peroxidase. Enzyme Antigen* 2006; 3: 1–16.

Received May 28, 2012.
Accepted October 27, 2013.